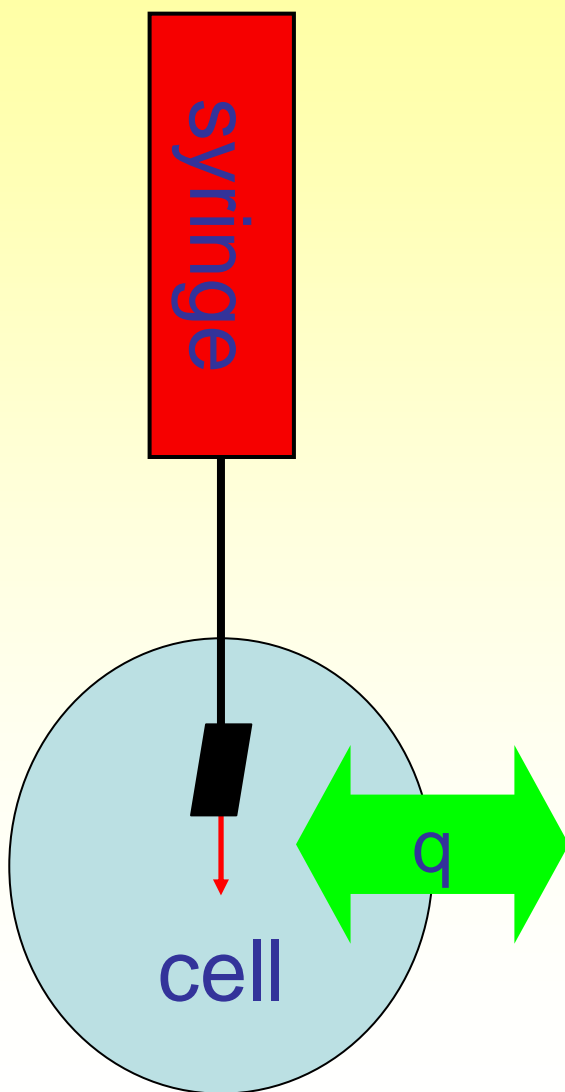


Techniques for Data Collection calorimetry

Calorimeters

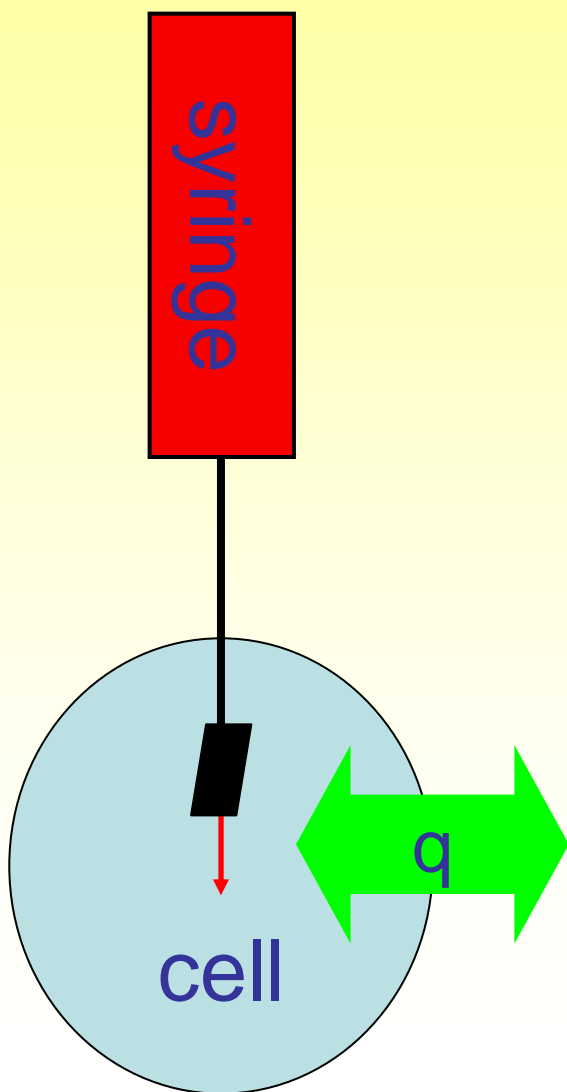
- Technique described using automated commercially available calorimeters but other setups feasible

Isothermal titration calorimetry



- Often used to study binding:
 - Guest in syringe is titrated into host in calorimeter cell
 - Heat effects of guest binding to host are measured and interpreted to give thermodynamic binding parameters

Kinetics by calorimetry



- Kinetics:
 - Reactant in syringe is titrated into solution containing other reactant or catalyst (enzyme) in calorimeter cell
 - Reaction heat effects are measured and interpreted in terms of rate equations

Kinetics by calorimetry

- Advantages
 - Measures rates
 - Label free (requires no chromophores or solution transparency)
 - High sensitivity: $\mu\text{cal s}^{-1}$ (reaction heats typically kcal mol^{-1} , requires rates typically $1\text{-}10 \text{ nM s}^{-1}$)
 - Automated data collection

Kinetics by calorimetry

- Disadvantages
 - High sensitivity (measures everything)
 - Requires matched solvents in syringe and cell
 - Cell is typically metal: less inert than glass
 - Cell is not fully closed (inert atmosphere difficult)

Kinetics by calorimetry

- Are you sure you know what your products are?
(also true for, e.g., UV-visible)

heat flow linear with reaction rate

$$power = \frac{dQ}{dt}$$

$$Q = n \cdot \Delta H_{app} = [P]_{total} \cdot V_o \cdot \Delta H_{app}$$

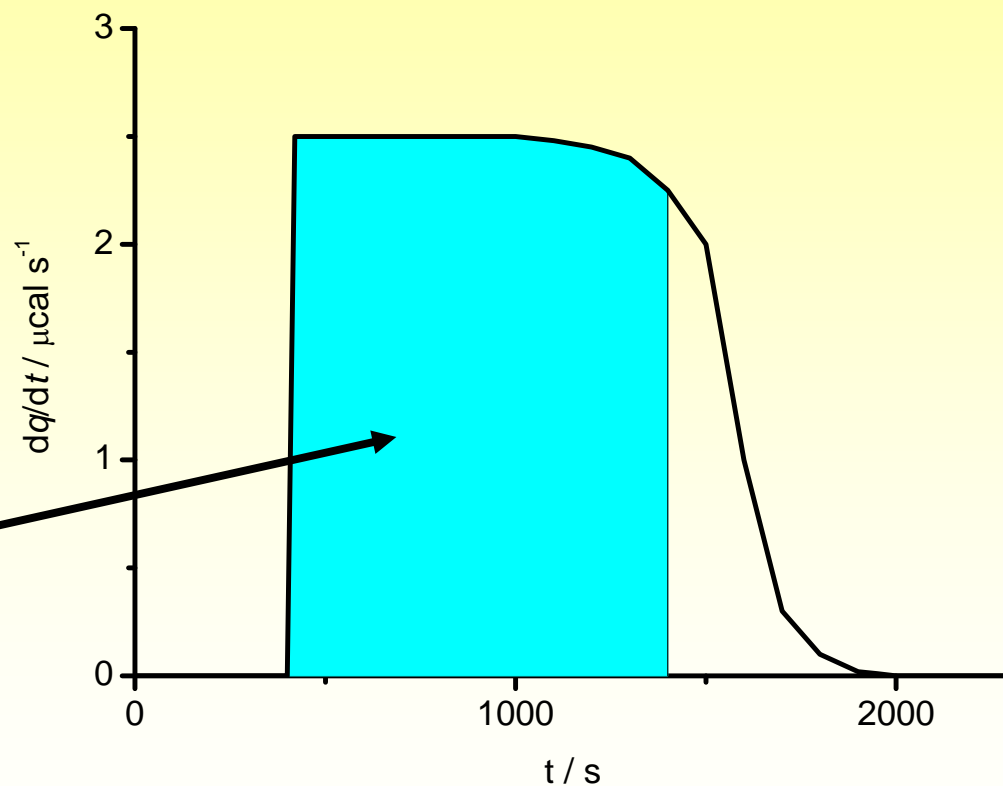
$$power = \frac{d[P]_{total}}{dt} \cdot V_o \cdot \Delta H_{app}$$

$$\frac{d[P]_{total}}{dt} = \frac{1}{V_o \cdot \Delta H_{app}} \cdot \frac{dQ}{dt}$$

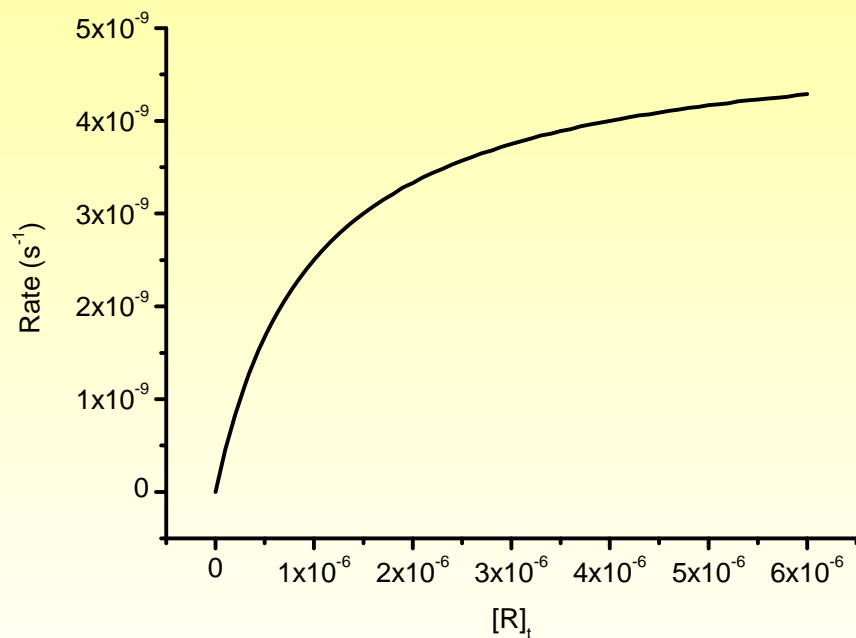
$$[R]_t = [R]_{t=0} - \frac{\int_0^t \frac{dQ}{dt} dt}{V_o \cdot \Delta H_{app}} \cdot$$

- remaining reactant concentration from extent of reaction

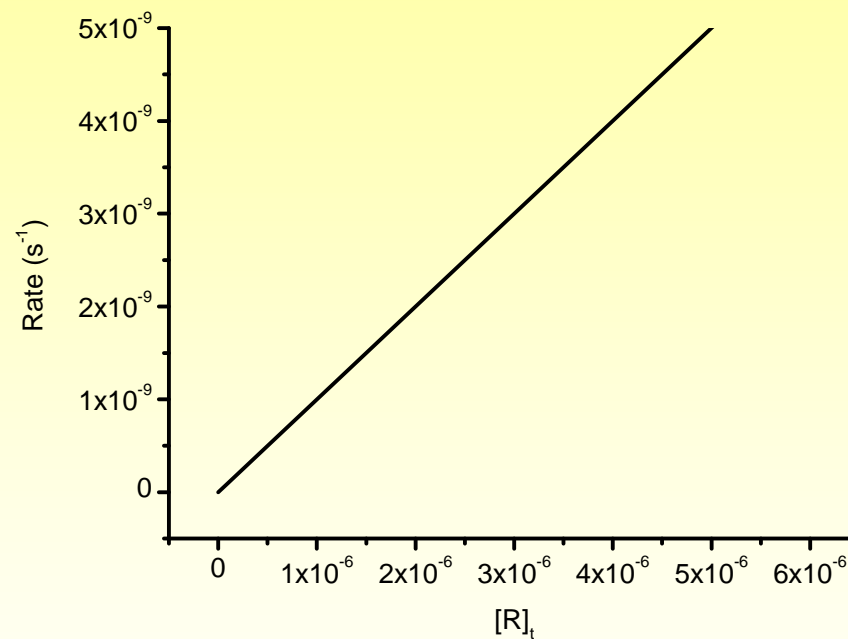
$$[R]_t = [R]_{t=0} - \frac{\int_0^t \frac{dQ}{dt} dt}{V_o \cdot \Delta H_{app}}$$



Rate as function of [reactant]



Michaelis Menten



$$\frac{d[P]}{dt} = k[A]^a[B]^b[C]^c$$