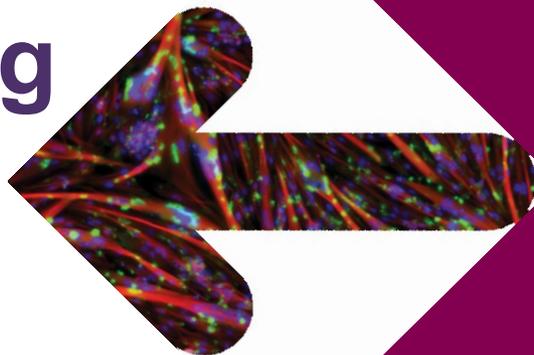


Phenotypic vs target screening approaches – black or white or shades of grey?

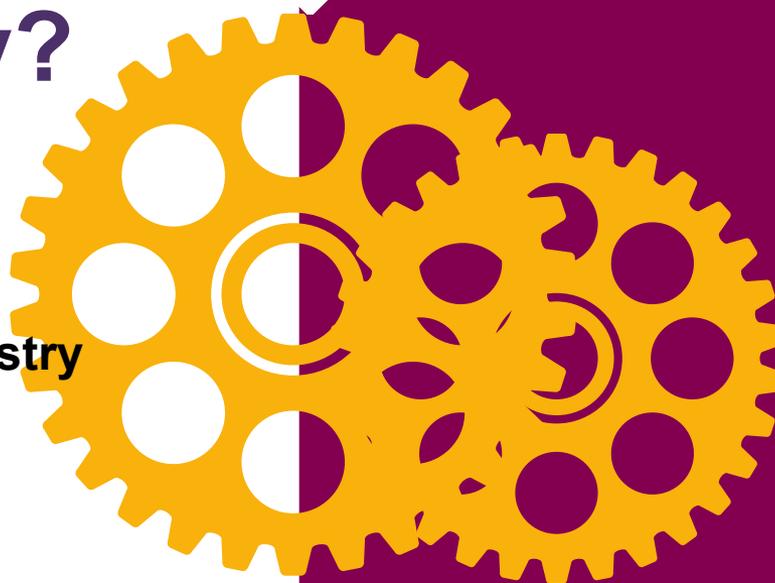


Graeme Walker

**The Society of Chemical Industry
Choosing the Right Target in
Drug Discovery
London, May 15th, 2013**

AstraZeneca 

Discovery Sciences | Reagents & Assays



Discovery Science competencies

How can we maximize value and impact into drug discovery projects?

- Crystallography; first structures, iterative structures
- NMR and other biophysical techniques

- Fragment Chemistry
- Chemical Biology

- Computational biology
- Cheminformatics
- Predictive Chemistry
- Compound collection enhancement



- Compound management
- Biobanks (clinical samples)
- Hit generation
- SAR screening
- Ion channel centre of excellence
- Proteins
- Cells
- Transgenics
- Antibodies

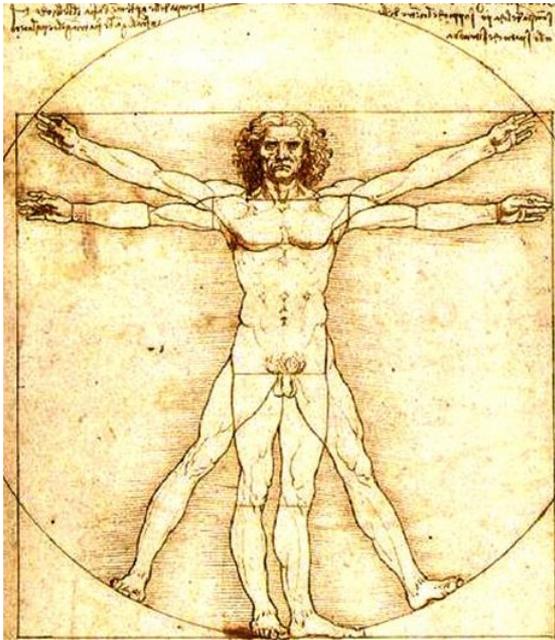
- Project support
- Statistical qualification

- Assay development
- High content biology assays



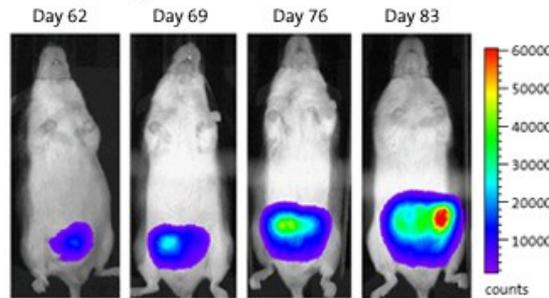
Phenotypic versus Target approaches

What do we mean?

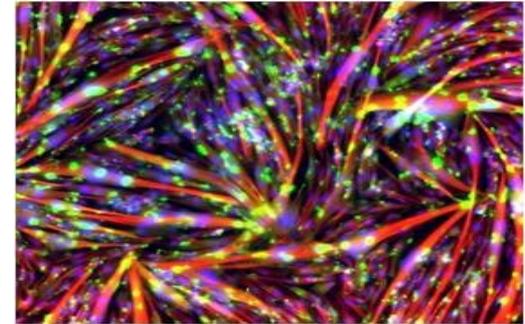


Patient

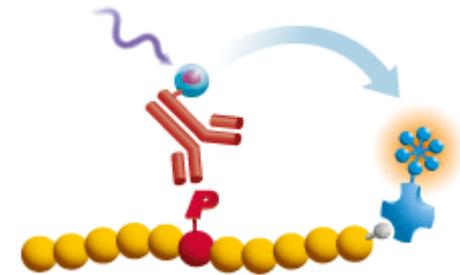
Glowing Prostate Tumors
PSA-Luc/iPB-TAg mice



Animal Models



Cellular Systems

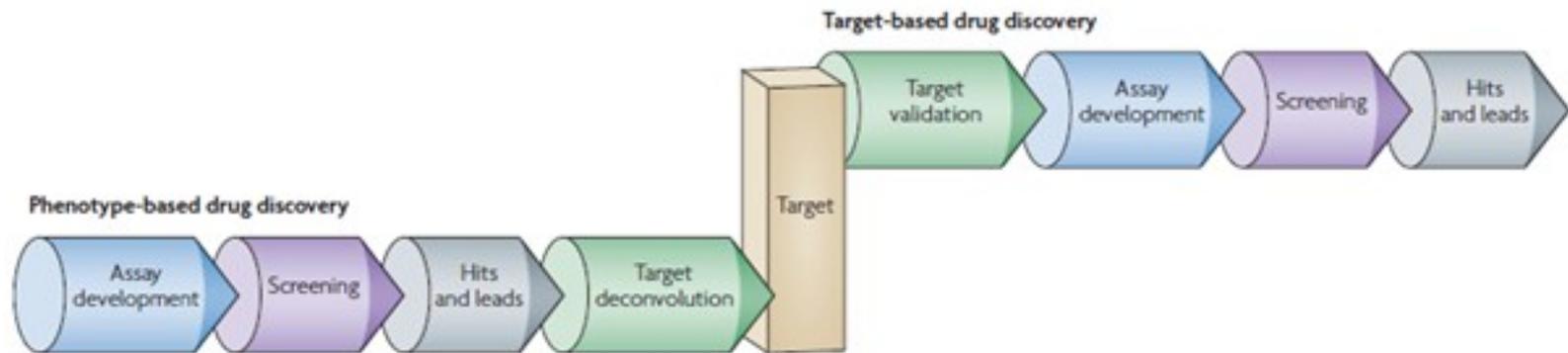


Isolated Enzyme



Target Directed (TDD) vs Phenotypic Drug Discovery (PDD)

- Targets are identified and validated (?)
- Typically use recombinant proteins or cells over-expressing the target of interest
- Assay throughput is usually high
- Screens used are to measure the compound's effect on the target of interest
- **Need to confirm compound effects in biological effect assay**



Terstappen *et al*, Nature Reviews Drug Discovery, 2007, 6, 891-903

- Targets are unknown
- Ideally use native human cells
- Assay throughput is usually low
- Screens used to measure the desired biological effect in cells, tissues or whole organisms where multiple, biologically relevant targets and pathways are simultaneously interrogated
- Activity in phenotypic screening might be translated to human disease more effectively than that in target-based screens
- **Need to do target deconvolution to identify target**



Target-driven/directed Drug Discovery (TDD) VS Phenotypic Drug Discovery (PDD)

TDD: The ability of compounds identified in target-driven approaches to modify disease progression in patients is not known *a priori* and may not be related to the biochemical activity of the compound *in vitro*

PDD: Simultaneously interrogating multiple, biologically relevant molecular targets and pathways to discover compounds that modulate relevant biological processes in a target/mechanism agnostic fashion

- novel functions for well-studied proteins
- discover new pathways of therapeutic value

Novel target discovery

Increase chemical diversity

Novel compound mechanism of action

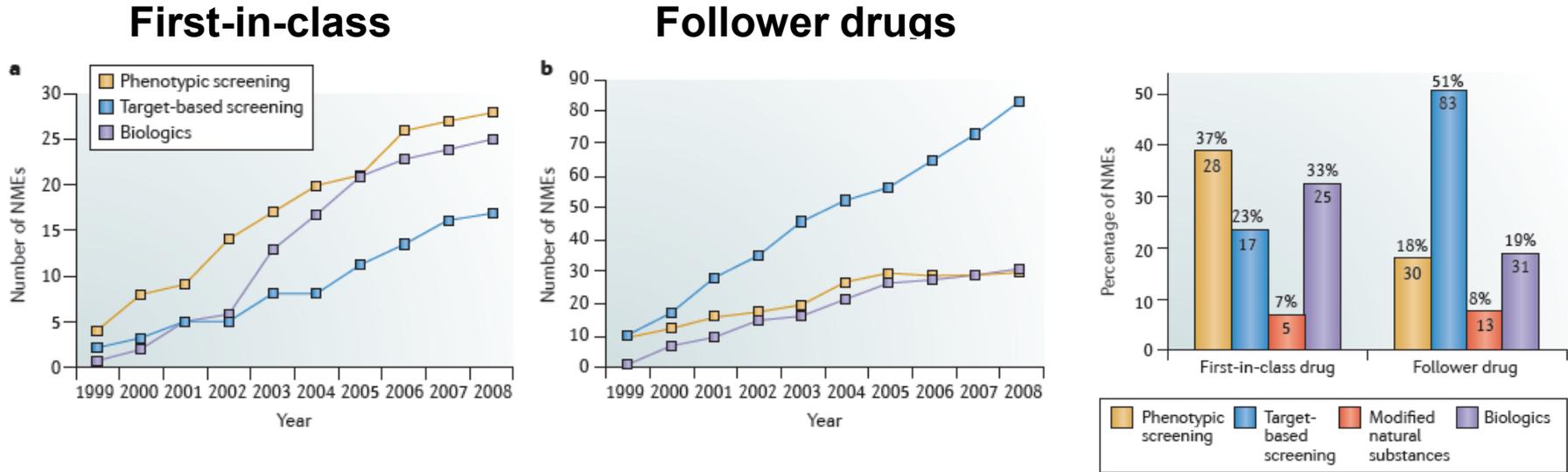


Why Phenotypic Discovery?

- Analysis of drug discovery over the last ten years is a story of massively increasing expenditure with less delivery of new drugs to the market
- Some of this can be explained by tougher regulatory policies but the fact remains that unless this improves we will not be able to replace the loss of previous “blockbusters”
- One possible explanation is that target-directed drug discovery driven by molecular biology and HTS has not delivered to expectations
- Over the last ten years most First In Class (FIC) small molecule drugs have still come from phenotypic screens rather than target directed screening
- Efficiency vs effectiveness?
- Followers (fast or slow)/Best In Class (BIC) tend to use target based approaches



Drugs discovered by target-based and phenotypic-based approach between 1999 and 2008



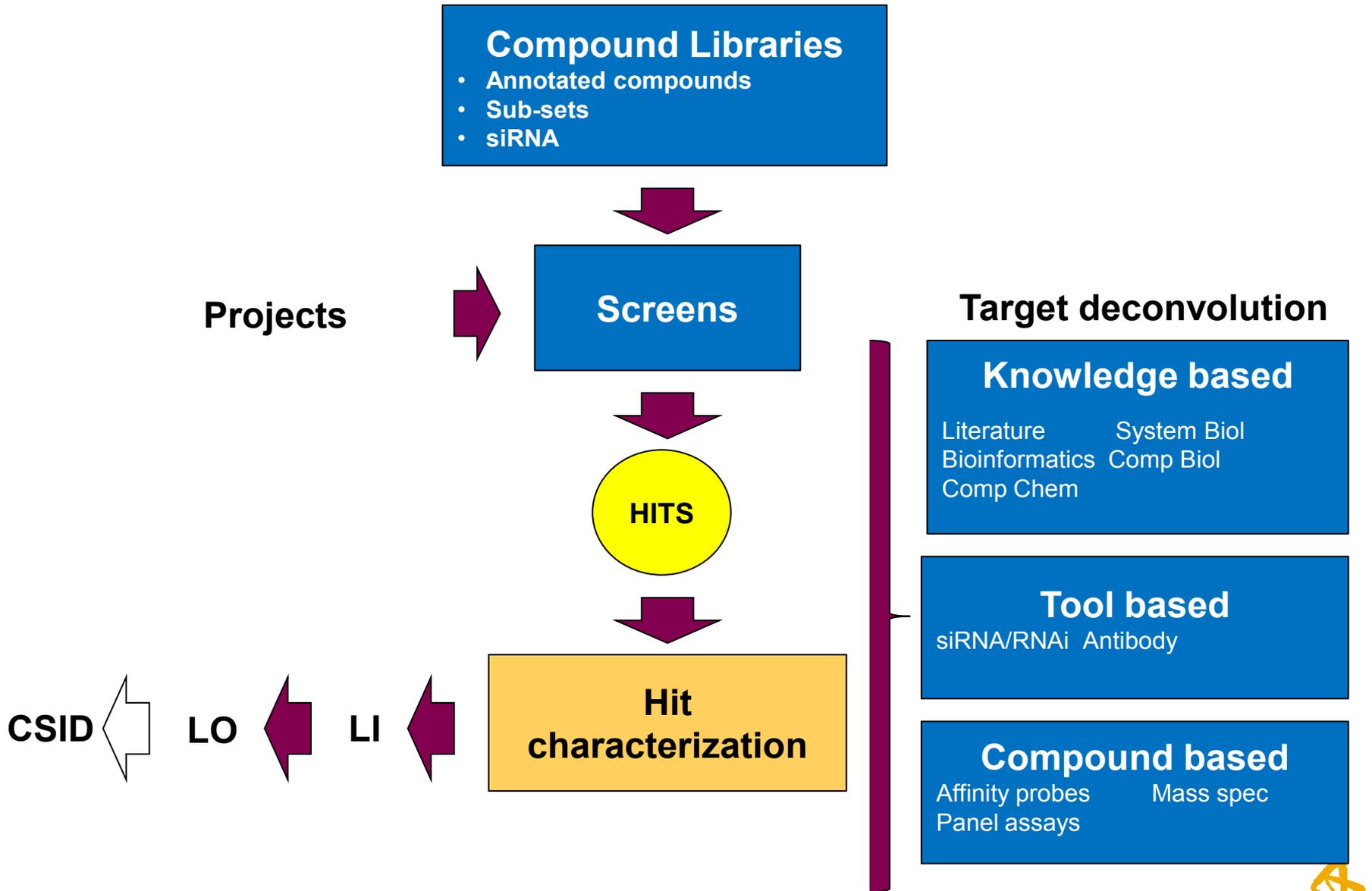
Swinney and Anthony, Nature Reviews Drug Discovery, 2011, 10, 507-519

Main problems with phenotypic screening are:

- Phenotypic assays have generally been low throughput
- This necessitates deconvolution of target for HTS
- Cell based (phenotypic assays) are now much better – good enough to drive Med Chem?



Phenotypic Discovery Cascade



Bridging the gap

Grey area between Target and Phenotypic approaches?

- Pathway specific approaches
 - Monitor specific pathways then identify upstream target(s)
- Other approaches to improve chances of technical success in a project
- Build cellular assays which are more predictive of in-vivo and clinical response
 - Use of primary and stem cells
 - Use of 3D cell culture systems
 - Combine target and phenotypic end-points
- Design cascades to identify compounds with a specific molecular mechanism of action with greater disease relevance
 - Covalent inhibitors
 - Down-regulators of targets



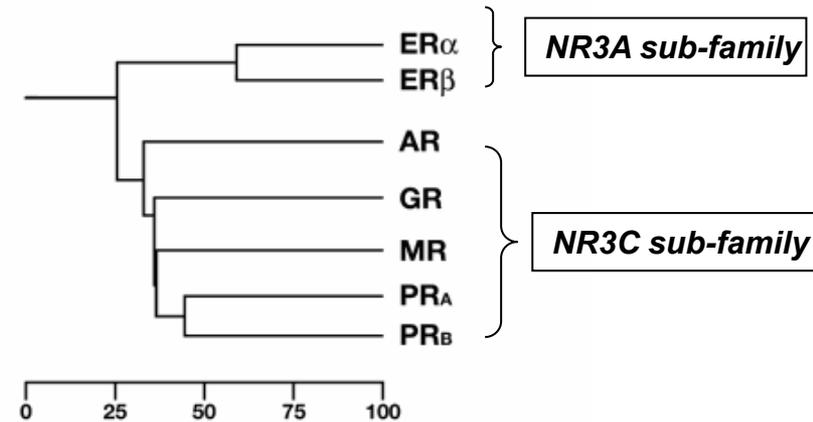
Selective Estrogen Receptor Down-Regulator (SERD) Project

Identifying compounds with a different molecular mechanism of action



ER α basics and link to breast cancer

- Member of the nuclear receptor superfamily
 - Steroid hormone receptor (ER α , ER β , PR, GR, MR and AR)
- Ligand-activated transcription factor which regulates expression of estrogen responsive genes
- Natural ligand is estradiol
- Normal role is in female reproductive function and maintaining bone density.
- Role in breast cancer - ~75% of cancers are ER and /or PR +ve in postmenopausal women
 - Candidates for endocrine treatment
- ER α is a key transcriptional regulator in driving ER+ve breast cancer proliferation
- Estrogen link first identified by Sir George Beatson in the 19th century
 - Ovariectomy leads to a reduction in breast tumour size



ER α versus ER β
DBD – 95% homology
LBD – 53% homology



Evolving endocrine treatments

- Current treatments
 - block ER α signalling using antagonists such as Tamoxifen
 - inhibit synthesis of Estrogens - aromatase inhibitors (Anastrozole)
 - removing ER α with an ER α -downregulator (SERD) (Fulvestrant)
- Tamoxifen has been the mainstay of endocrine treatment for many years
 - Antagonist on breast; partial agonist on bone and endometrium
 - Selective Estrogen Receptor Modulator (SERM)
- Subsequent 2nd generation SERMS
 - Raloxifene, Lasofoxifene, Bazedoxifine
- Third generation aromatase inhibitors
 - Anastrozole
- Fulvestrant - SERD
 - Pure anti-estrogen; no agonist effects; novel mode of action



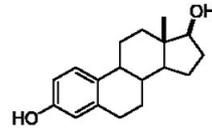
SERMS and SERDs

Mechanistically different classes of ER modulators

**SERM =
Selective Estrogen
Receptor Modulator**

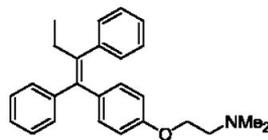
Agonists:

Estradiol

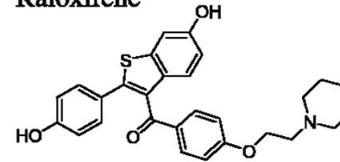


SERMs:

Tamoxifen

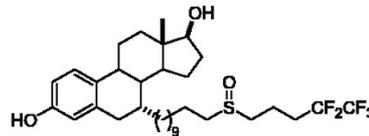


Raloxifene

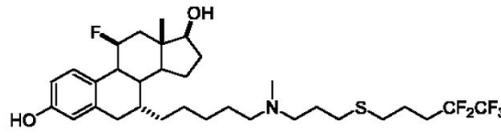


SERDs:

ICI 182,780

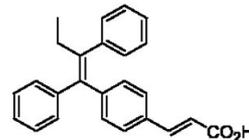


ZK-703



Mixed Function SERM / SERDs:

GW5638

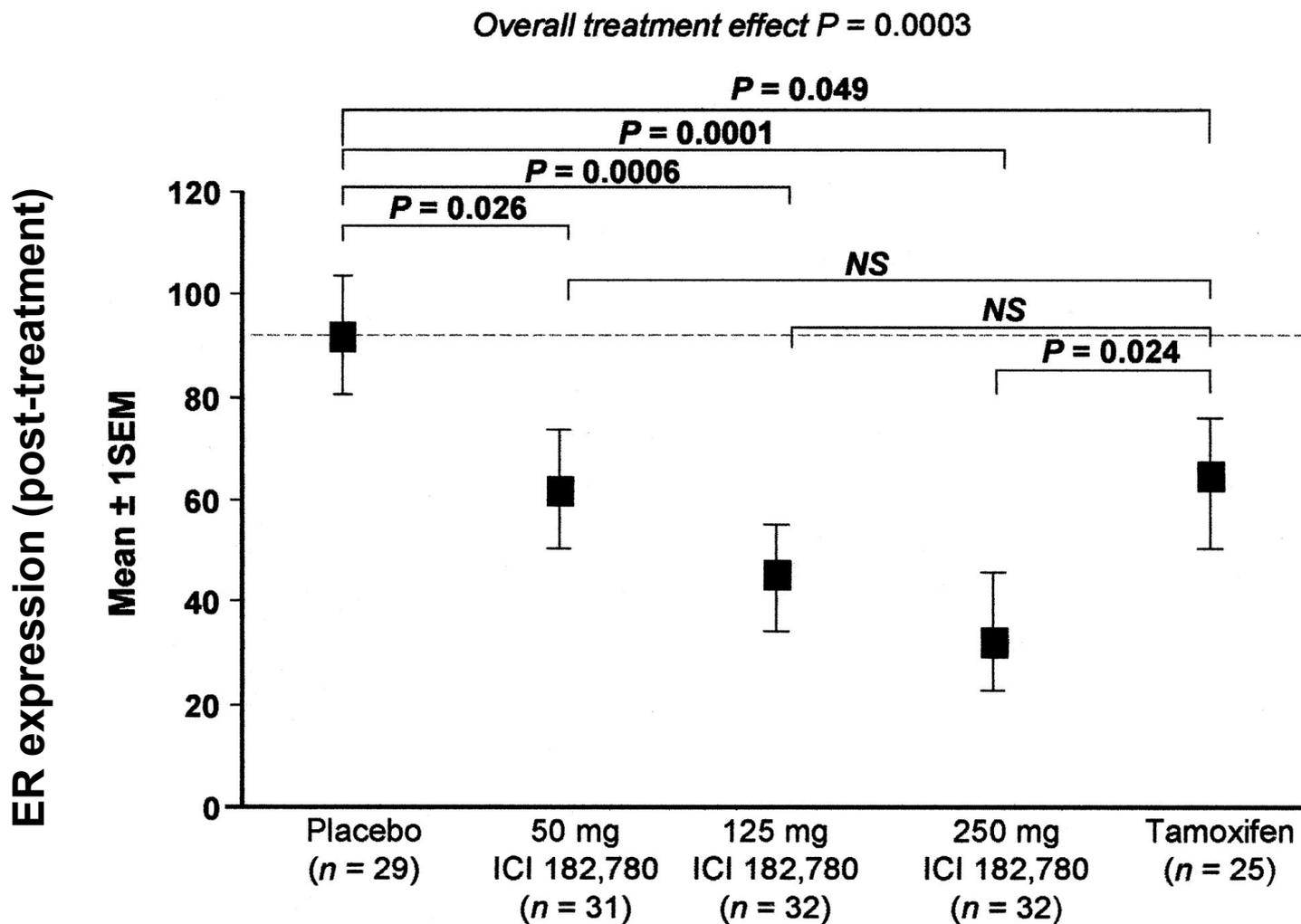


Issues with
SERMS led to a
desire for a “pure
anti-estrogen”
agent

**SERD =
Selective Estrogen
Receptor Down-
regulator**



Fulvestrant binds the ER, blocks hormone signalling and increases receptor degradation

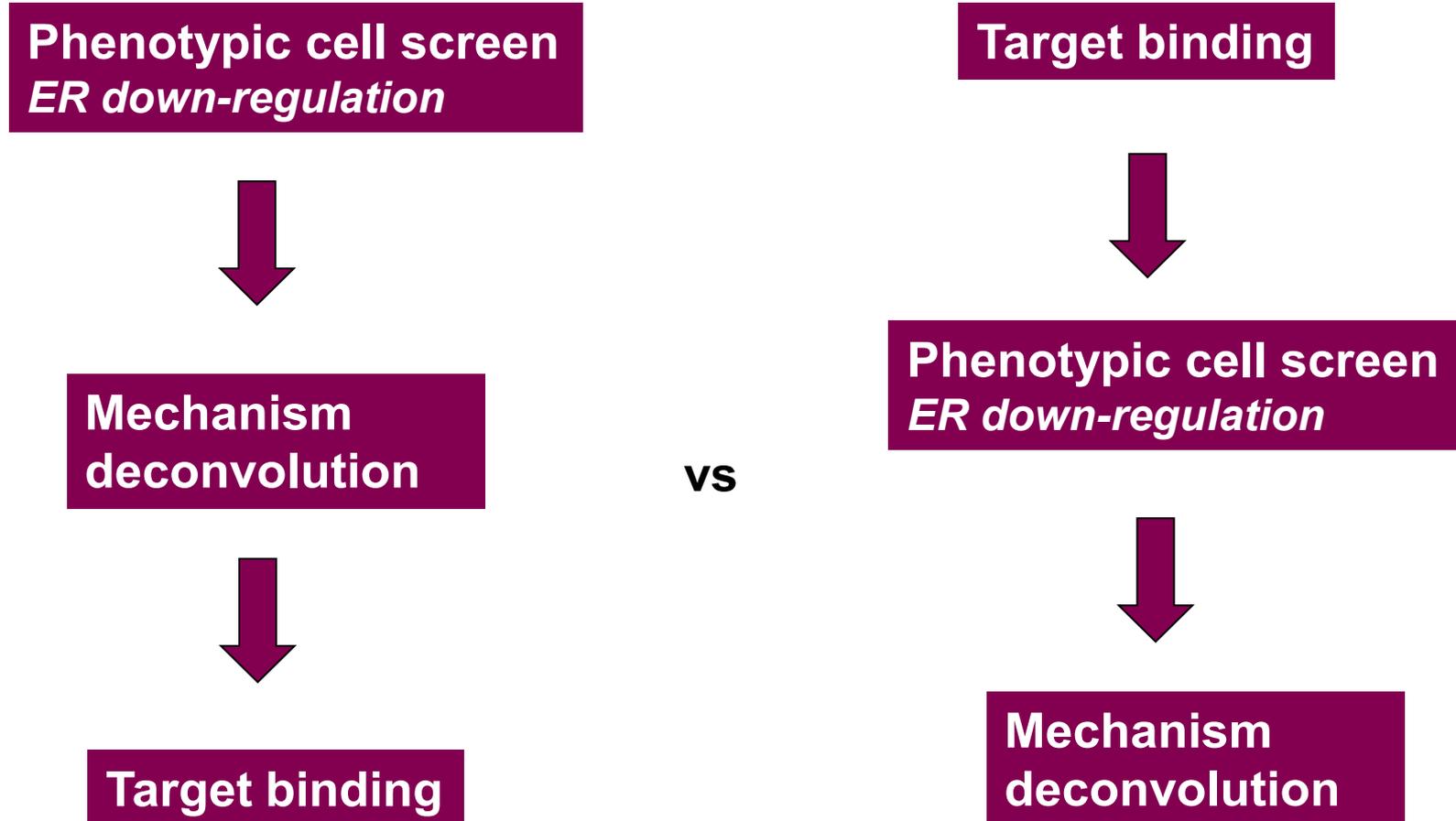


Why a novel SERD?

- Fulvestrant is approved as a second line therapy in ER positive PM women after anti-estrogen therapy failure
- Evidence that down-regulation is linked to efficacy
 - Higher dose regimens recently approved
 - \$160m sales in 2006 (Anastrozole \$1.7b) - 250mg dose
 - 500mg dose approved Sep 2010 – 2011 sales \$546m
- Once monthly 5ml injection – oral route preferred
- Ongoing clinical trials to evaluate dose scheduling and combinations with Anastrozole
- Novel oral agent with greater efficacy desired



Lead Generation strategy?



Lead Generation Strategy

- Within AZ many previous efforts had targeted ER α , or β , as well as estrogen-related receptors
 - External literature and competitors
 - Analyse collection to identify non-steroid (non-tamoxifen), non-phenol ER binders - early start-points for chemistry
- Structural information is available, and steroid-pocket binding hypotheses are available for ER and GR
 - Establish ER α structural system in house
- Sub-set HTS biochem screen
 - Identify novel binders then modify them into down-regulators
- Build cellular cascade to drive SAR and understand mechanism of action of compounds
 - Multiple mechanisms of ER α -downregulation exist
 - Cellular cascade assays are required which can differentiate down-regulation via direct binding to the ER α , agonist feedback or off target mediated down-regulation



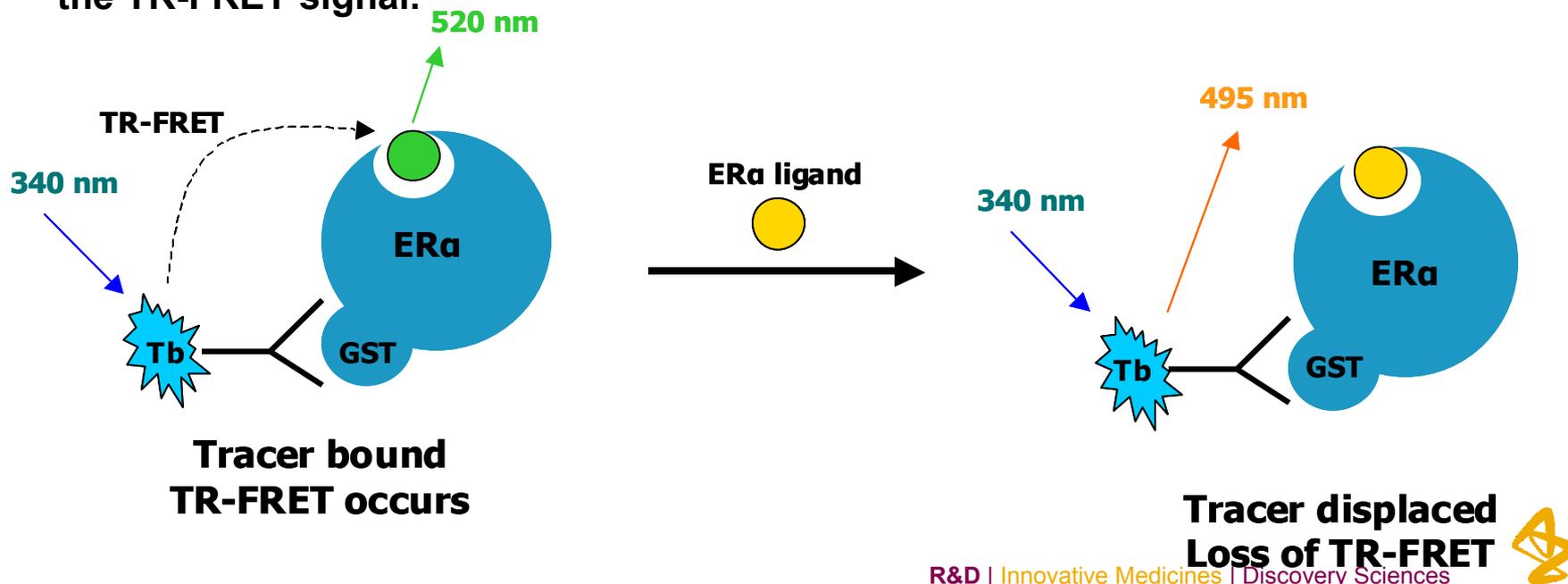
ER α Ligand Binding Domain (LBD)-GST Assay Overview

ER α LBD-GST Time Resolved-Fluorescence Resonance Energy Transfer (TR-FRET) Competitive Binding Assay - Invitrogen

A tracer & antibody-based HTS method for identification of ER α ligands.

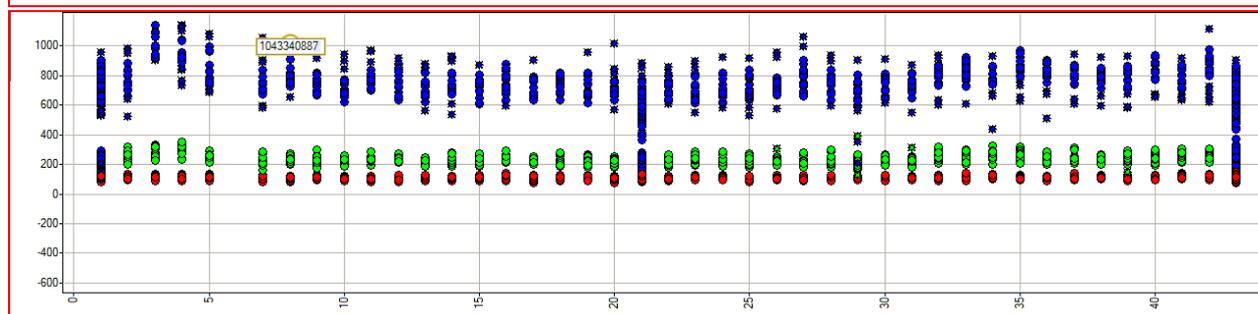
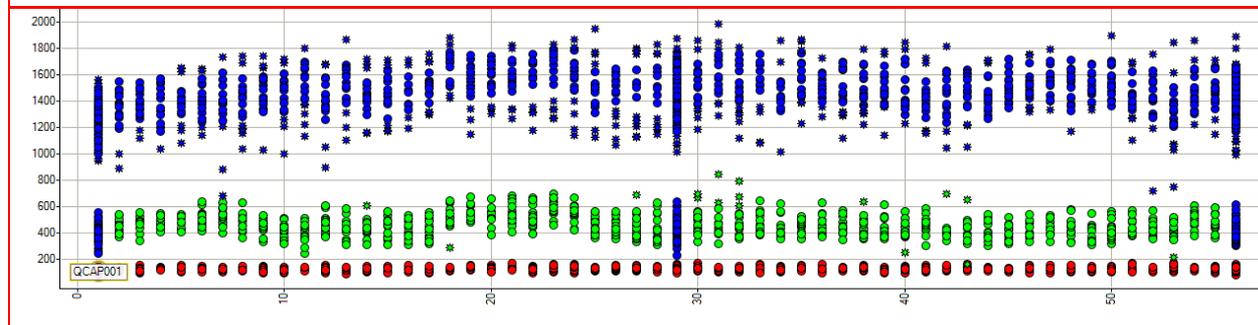
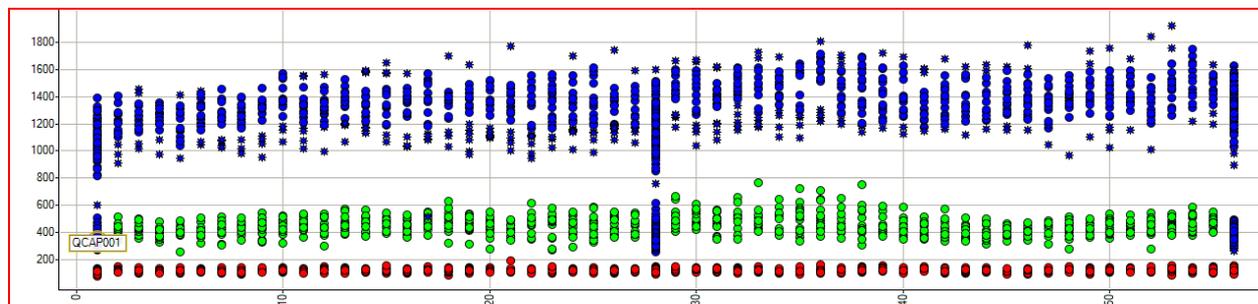
Binding of the tracer (fluorescent steroid) to ER α is detected by TR-FRET from the terbium-labeled antibody (donor) to the tracer's fluorophore (acceptor).

ER α ligands can be identified by their ability to displace the tracer and disrupt the TR-FRET signal.



HTS follow up – control well data

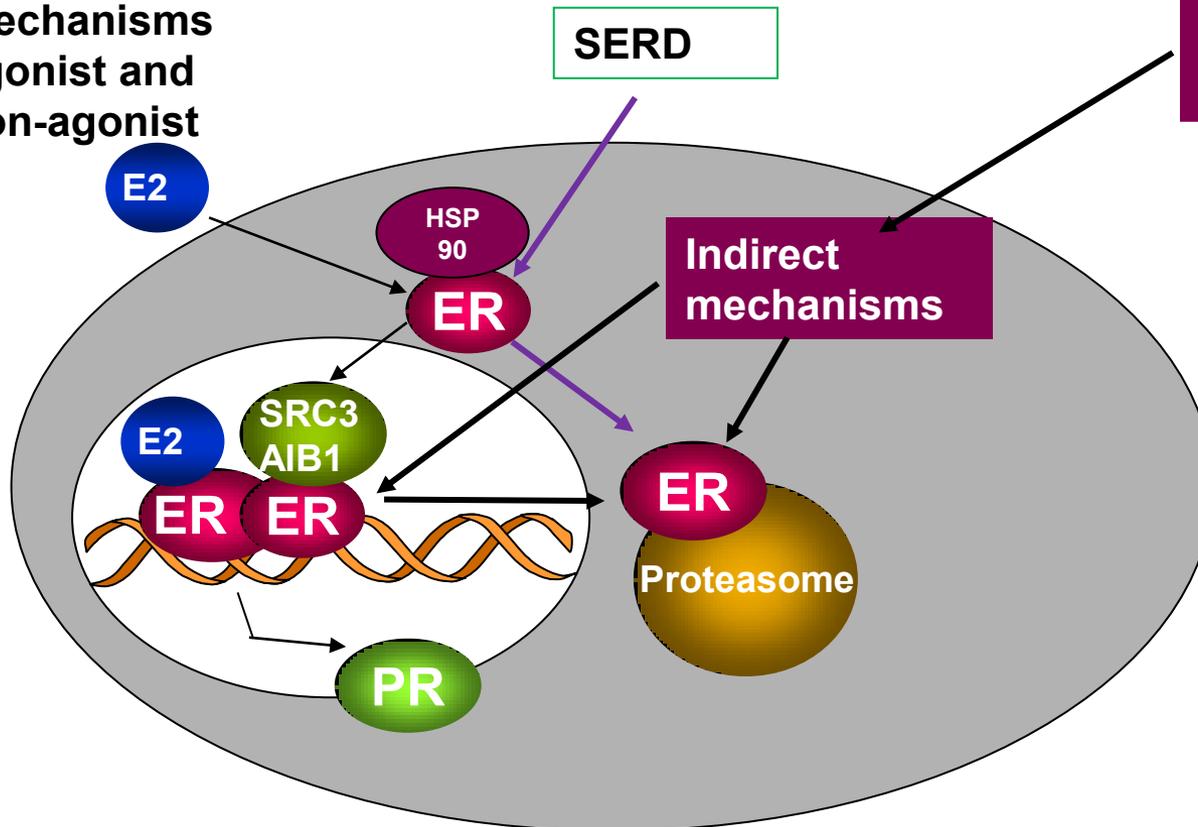
100k sub-set screen ; 6k followed up in concentration response format
0.5k < 1 μ M IC₅₀ ; 2.5k < 10 μ M ; 15 distinct series ; 7 confirmed by X-ray



Multiple mechanisms of ER α regulation exist – Mediated via ligand binding to ER α or other targets

Post HTS – identifying novel ER α down-regulators was feasible but vast majority had “agonist” profile

Direct mechanisms
agonist and
non-agonist

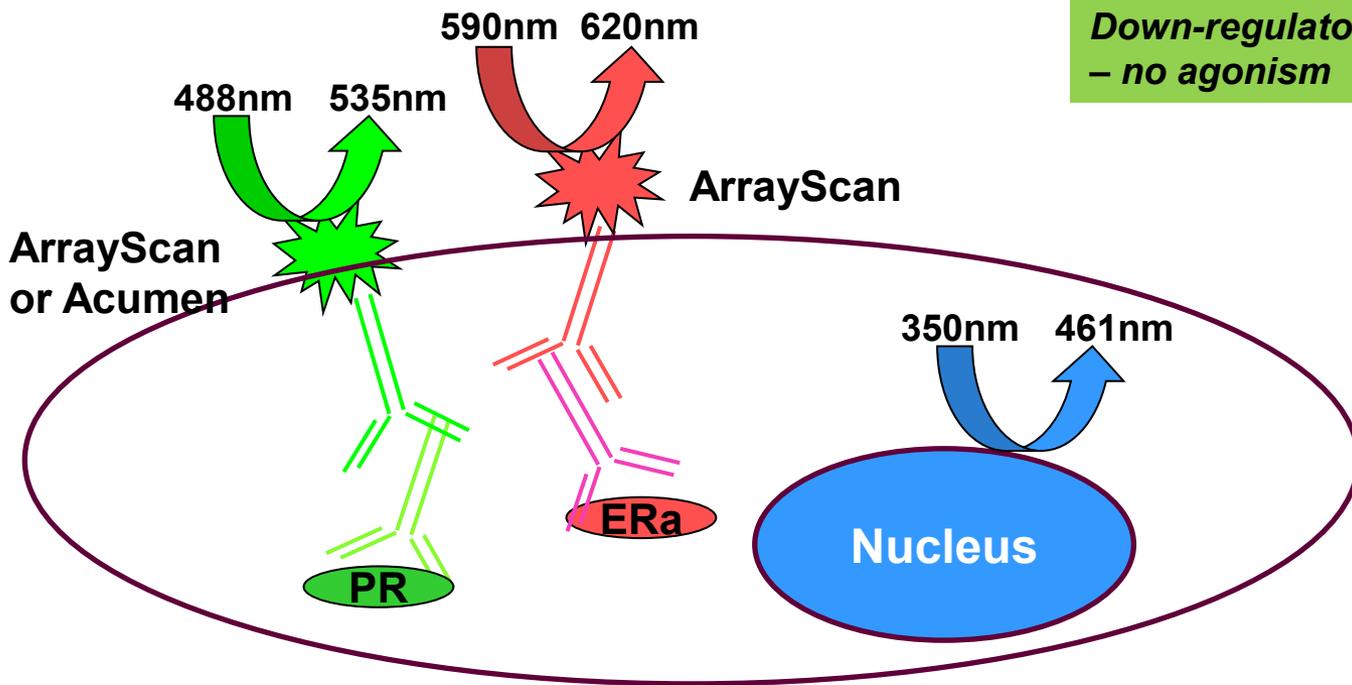


HDAC inhibitors
HSP90 inhibitors
Kinase inhibitors



Multiplexed MCF7 cell assay

- Immunofluorescence quantified on the ArrayScan (ER) and Acumen (PR) platforms
- Total levels of ER α and PR are detected by specific antibodies then by labelled secondaries
- Detection after 24 hour cpd treatment to enable PR signal to be induced
- Finalized assay uses cryopreserved cells, 384 well format and automated antibody staining
- Has been modified to read-out as a functional antagonism assay – pre-dose with E2
- Full validation package completed successfully.

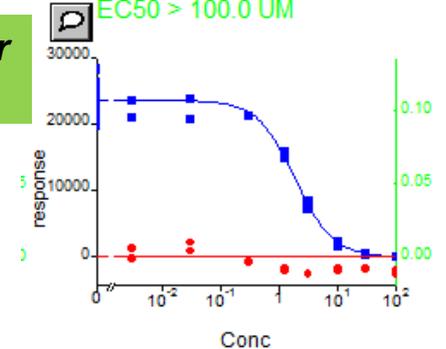


**Down-regulator
– no agonism**

Literature cpd.

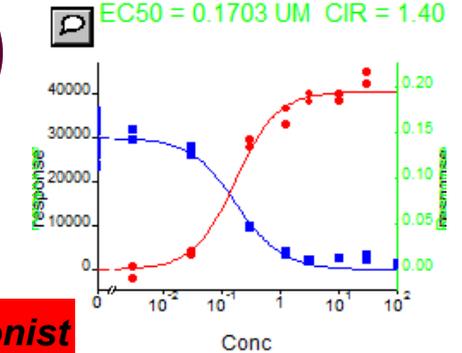
IC50 = 1.857 μ M

EC50 > 100.0 μ M



IC50 = 0.1750 μ M CIR = 1.71

EC50 = 0.1703 μ M CIR = 1.40



Agonist



Assay Overview - ~300 cpds/run in 12 point duplicate concentration response format

Day 1



Day 2



Day 3



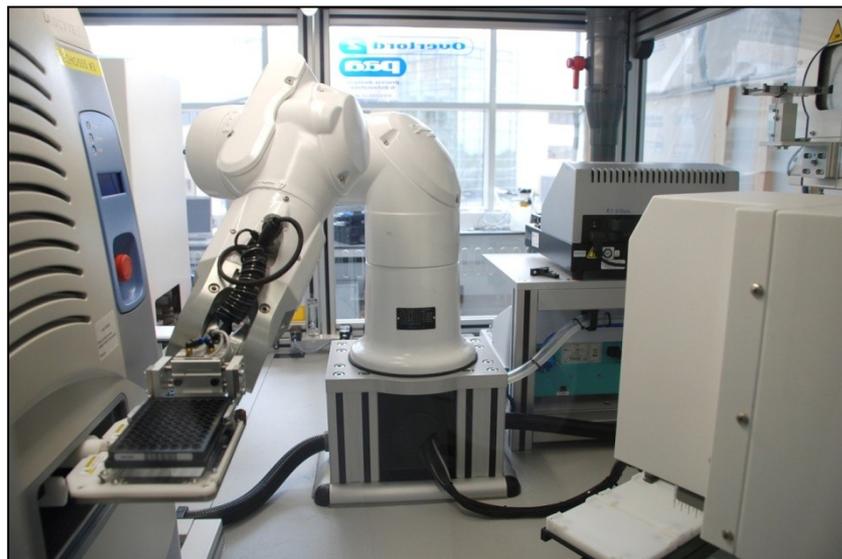
Day 4

Seed cryopreserved MCF-7 cells in 384 well plates.

Automated compound dispensing using Integrated Echo workcell followed by 24 hr incubation.

Fix cells (20mins). Automated antibody staining using AutoElisa workcell (1hr permeabilization & incubate with primary antibodies to ER and PR overnight)

Automated antibody staining using AutoElisa workcell (1hr incubation with secondary antibodies & Hoechst). Read plates on Acumen for PR detection and Arrayscan for ER α detection



Mechanism of Action assay

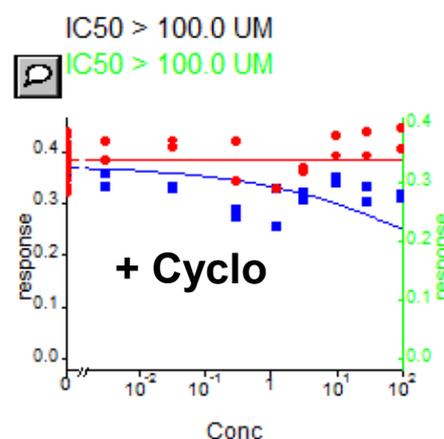
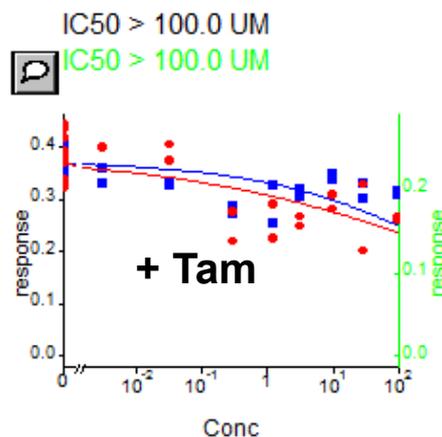
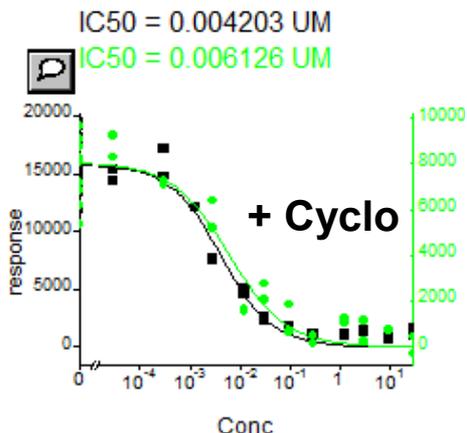
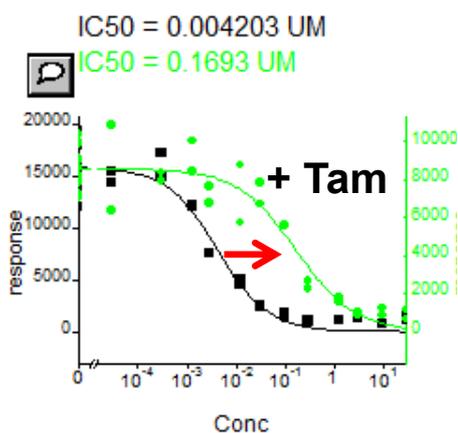
- Comprises 3 tests in parallel to distinguish direct binding down-regulators of ER α from off target mechanisms.
- Tamoxifen binding stabilises ER α
- Cells are pre-treated with 250nm Tamoxifen or DMSO (control) or 25ug/ml Cycloheximide for 1hr prior to compound dosing and then incubated for 5 hrs.
- ER α only is detected on the Acumen or ArrayScan platform – timecourse is too short to have measureable PR induction.
- Potential SERD compounds will compete with Tamoxifen for binding to ER α and become less potent (0.5 log shift in potency for control compounds).
- No shift in potency is seen with off-target down-regulators and indeed some off-target compounds are inactive at 5 hours.
- Protein synthesis is implicated in the agonist induced down-regulation mechanism
- The cycloheximide treatment blocks protein synthesis in the cell which helps further to discriminate indirect and undesirable mechanisms of down-regulation.
- For on target compounds, the potency is largely unchanged in the presence of cycloheximide and this arm also verifies that activity is seen against pre-existing ER α .



MCF7 MOA cell assay

ER detected following DMSO, Cycloheximide and Tamoxifen treatment plus compound

- Immunofluorescence quantified on the Acumen or ArrayScan platform
- Total levels of ER α are detected by a specific antibody then by labelled secondary antibody
- 5 hour timepoint – 1 hour pre-incubation - 3 conditions, vehicle, + 250nm Tamoxifen, + 25ug/ml Cycloheximide
- On target – +Tam IC50 shifts ; Off target - +Tam IC50 no shift or inactive at 5 hours
- + Cycloheximide – compound active versus extant ER α and IC50 non significantly shifted



On - target – Novel SERD

Off - target – HDAC inhibitor



Impact on project – effective and timely compound triage

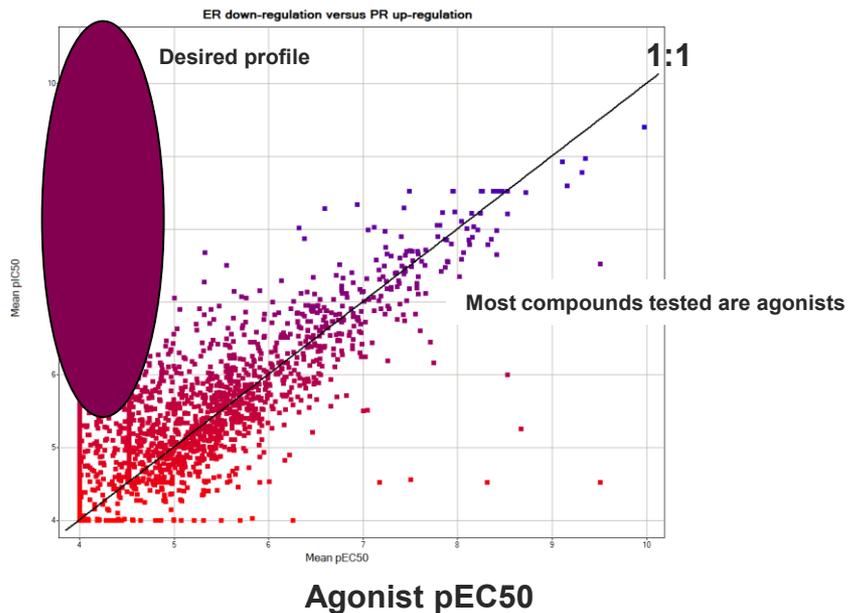
5k compounds

ER α down-regulation ; PR up-regulation

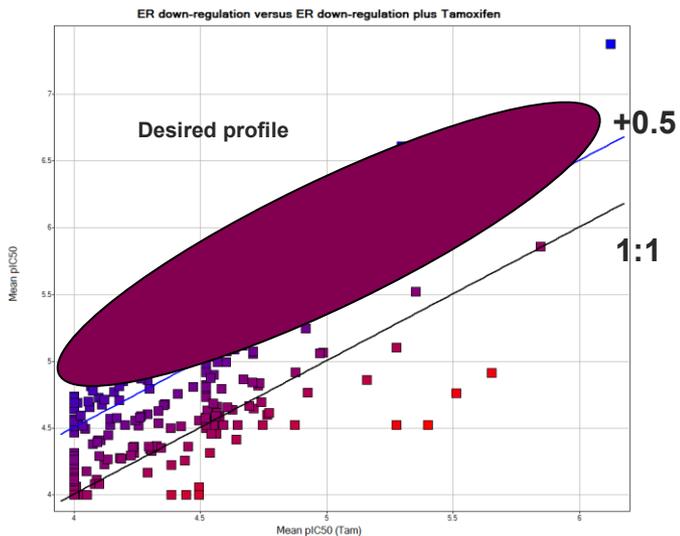
300 compounds

ER α down-regulation
Plus Tamoxifen/cyclohex/DMSO control

Down-regulation pIC50



Down-regulation pIC50

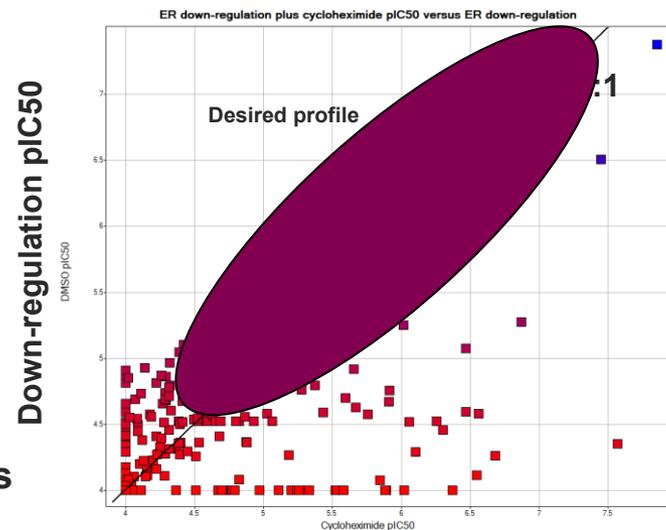


Down-regulation pIC50 + Tamoxifen

Desired profile
~100 compounds

Novel confirmed SERDs

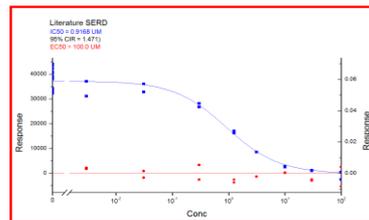
Down-regulation pIC50



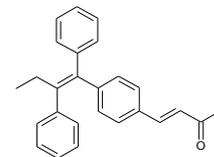
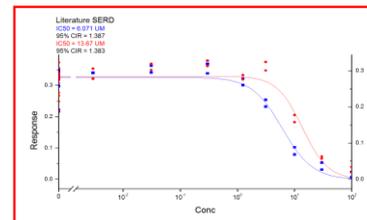
Compound Profiles

Literature SERD compound
 ER down-regulator ✓
 PR up-regulator ✗
 Competitive with tamoxifen ✓
 Active at shorter time point ✓
 The desired profile for a novel SERD

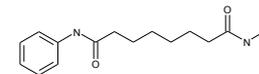
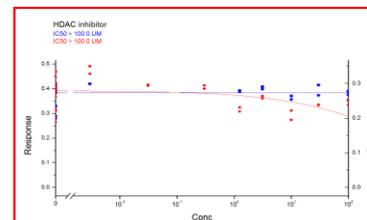
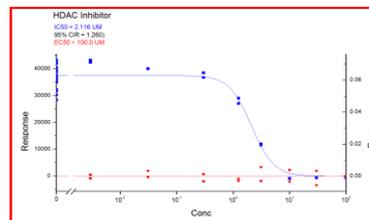
Overlay of ER α down-regulation
 with PR up-regulation after 24 hrs



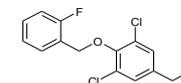
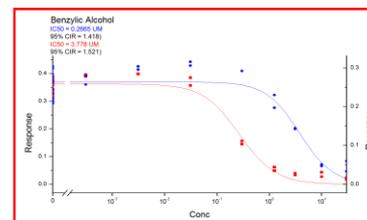
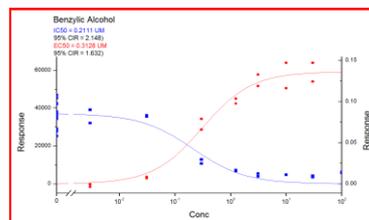
Overlay of ER α down-regulation in presence
 and absence of 250nmTamoxifen after 5 hrs



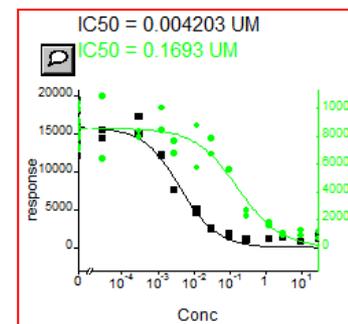
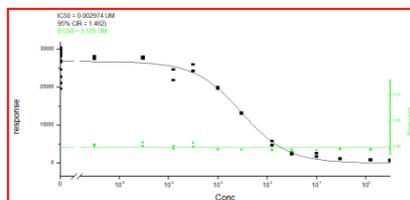
HDAC Inhibitor
 ER down-regulator ✓
 PR up-regulator ✗
 Competitive with tamoxifen ✗
 Inactive at shorter time point ✗
 An undesired profile for a SERD



Benzyl Alcohol
 ER down-regulator ✓
 PR up-regulator ✓
 Competitive with tamoxifen ✓
 ER agonist down-regulator profile
 An undesired profile for a SERD

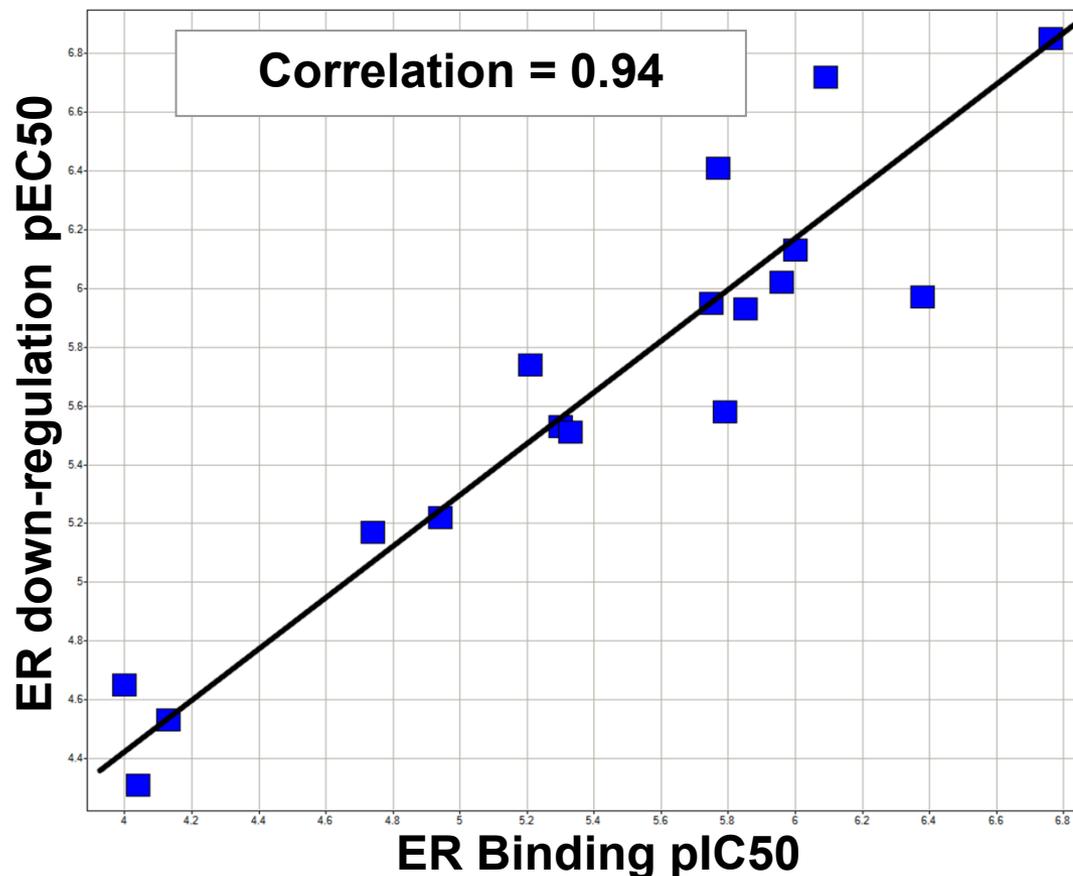


Novel SERD 1
 ER down-regulator ✓
 PR up-regulator ✗
 Competitive with tamoxifen ✓
 Active at shorter time point ✓
 Desired profile for a novel SERD



Correlation of ER α Binding and Down-regulation

The ER α down-regulation assay developed shows a very good correlation with ER α binding for the lead novel series with a correlation coefficient of 0.94.



Correlation among cellular down-regulation, antagonism and anti-proliferative effects for the novel series – some literature SERDs were better antagonists than SERDs



Novel SERD versus Fulvestrant

Multiplex ER α PR assay

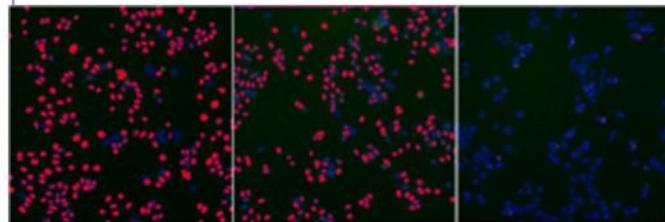
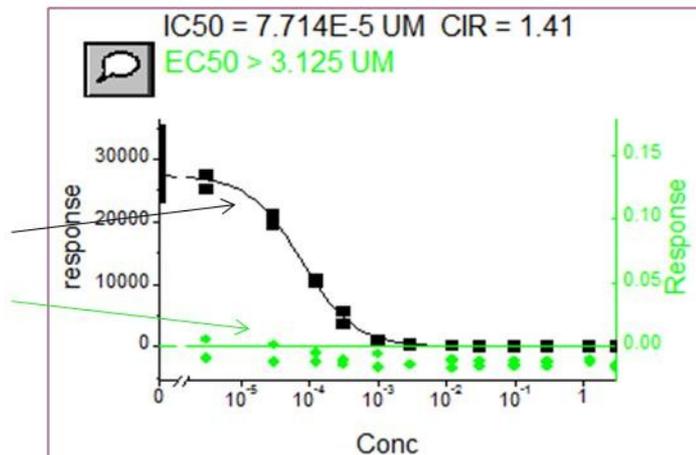
Multiplexed ER α PR assay

Black – Decrease in ER levels (down reg)

Green – No increase in PR levels (agonism)

ArrayScan images illustrating level of ER α staining

Fulvestrant

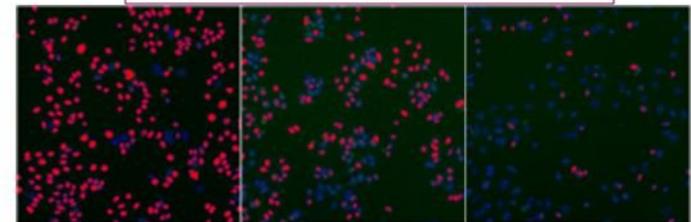
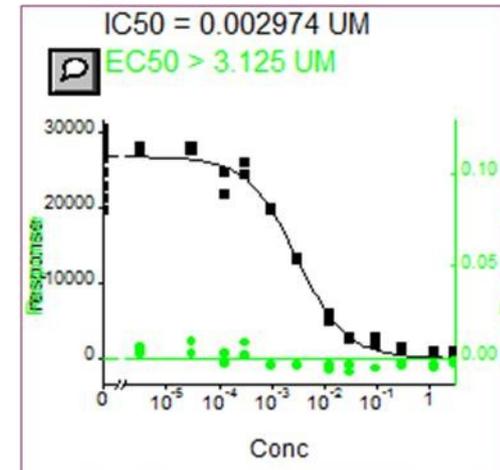


Max ER α response

50% effect

100% effect

Novel SERD



Max ER α response

50% effect

100% effect



Summary - Cell assays available to discriminate different compound profiles and drive SAR chemistry

Do compounds reduce ER α levels in MCF7 cells?

Are they acting via an agonist mechanism?

Are compounds active at 5 hours as well as 24 hours?

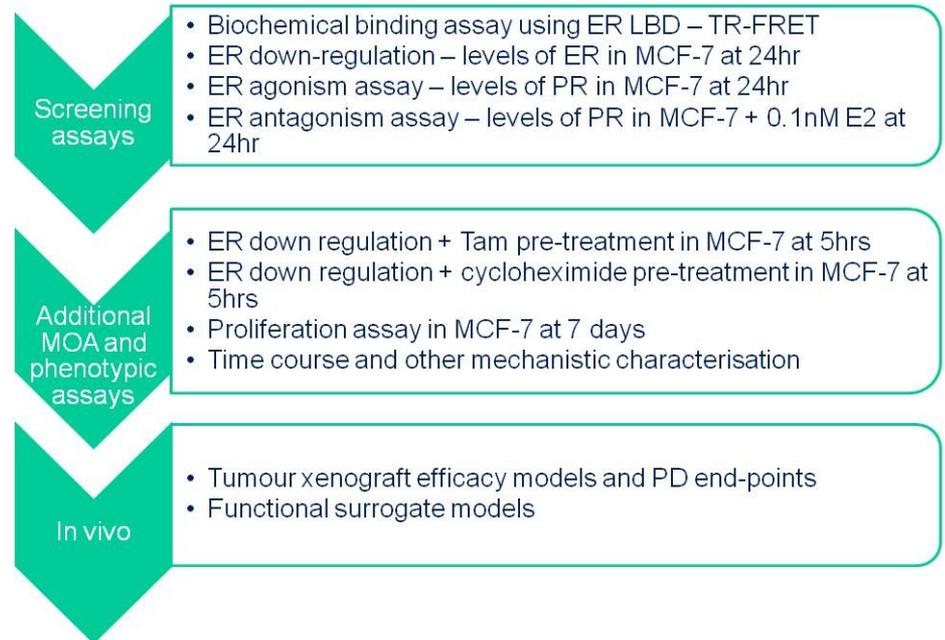
Do compounds work by binding to ER α ?

Are compounds able to reduce levels of existing ER α ?

Do compounds antagonise ER α mediated signalling?

Are compounds anti-proliferative in an ER α driven cell line?

How do novel SERDs compare to Fulvestrant?



Summary and Conclusions

- Resurgence in interest in phenotypic screening approaches
 - Relative lack of success using target directed approaches perceived to be primarily due to a lack of target validation
 - Developments in cellular reagents and assay platforms enable more predictive and robust phenotypic assays
- Phenotypic approaches require significant investment in target deconvolution
 - Projects are uncomfortable not knowing the molecular target
 - Target identity not necessary for drug registration
- Advances in cell reagents mean that target directed approaches can utilise assays which more accurately reflect disease biology and may ultimately lead to greater project success
 - Examples where human stem cell derived assays predict in vivo and adverse effects better than recombinant cell assays
- Opportunities exist to identify Best In Class molecules with different molecular mechanisms of action



Acknowledgements

- **Phenotypic Discovery Initiative**
 - Per-Erik Stromsted and Ryan Hicks
- **SERD Project Team Past and Present**
 - Michael Tonge (HTS and Biochemical assays)
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 - Rowena Callis (Cell Assays)
 - Iain Simpson (Chemistry)
 - Benedicte Delouvrie (Chemistry)
 - Al Rabow (Comp Chem)
 - Rob Bradbury (Chemistry)
 - Chris De Savi (Chemistry)
 - David Andrews (LG Project Leader)



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