
Discovery of novel antimalarials through cell-based medicinal chemistry optimization of HTS hits

What is GNF?

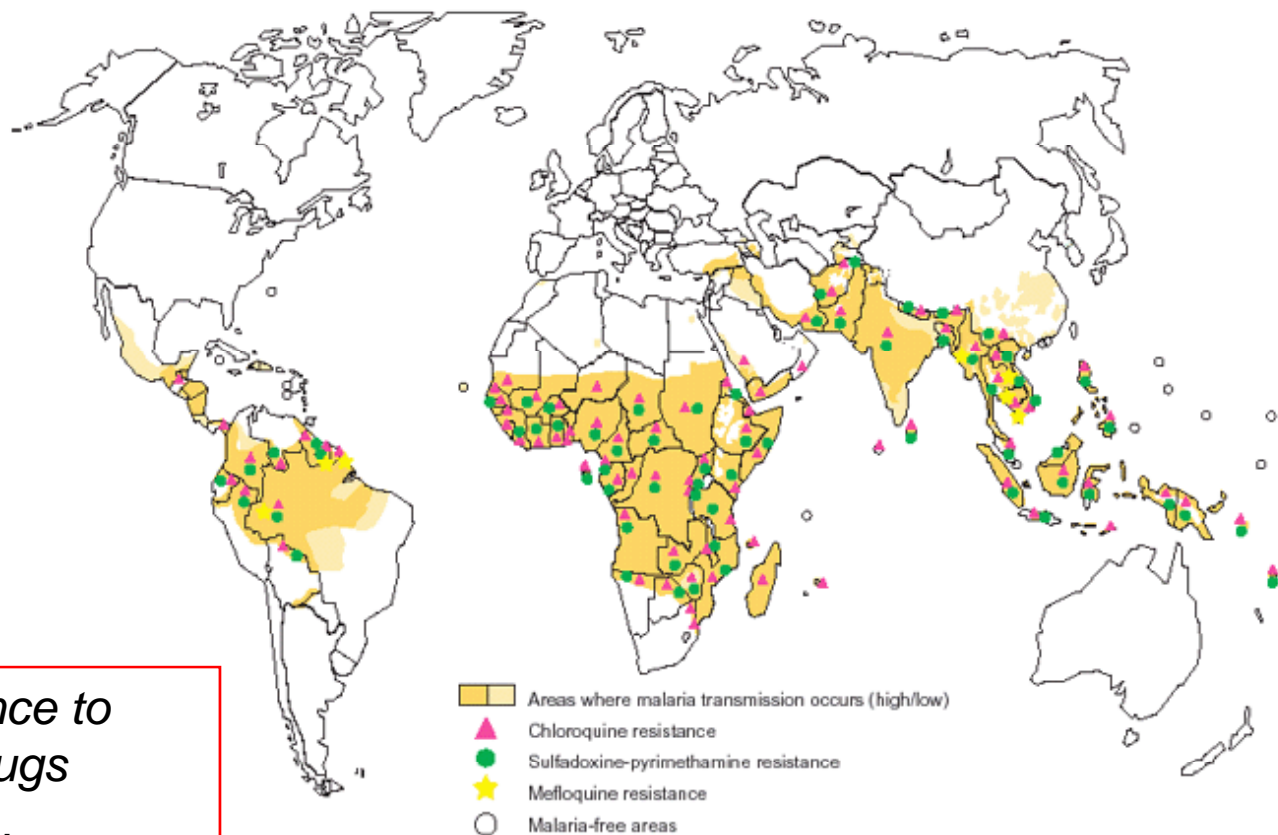


- Funded by the Novartis Research Foundation
- Moved into a temporary 40,000 square foot laboratory facility in 3Q 1999
- Moved into permanent 260,000 square foot research campus in 1Q 2002
- Additional 24,000 square foot manufacturing facility
- Present headcount ~580 FTE
- Currently, up to preclinical toxicity studies done at site, integrated with Novartis development

Malaria: Medical Need

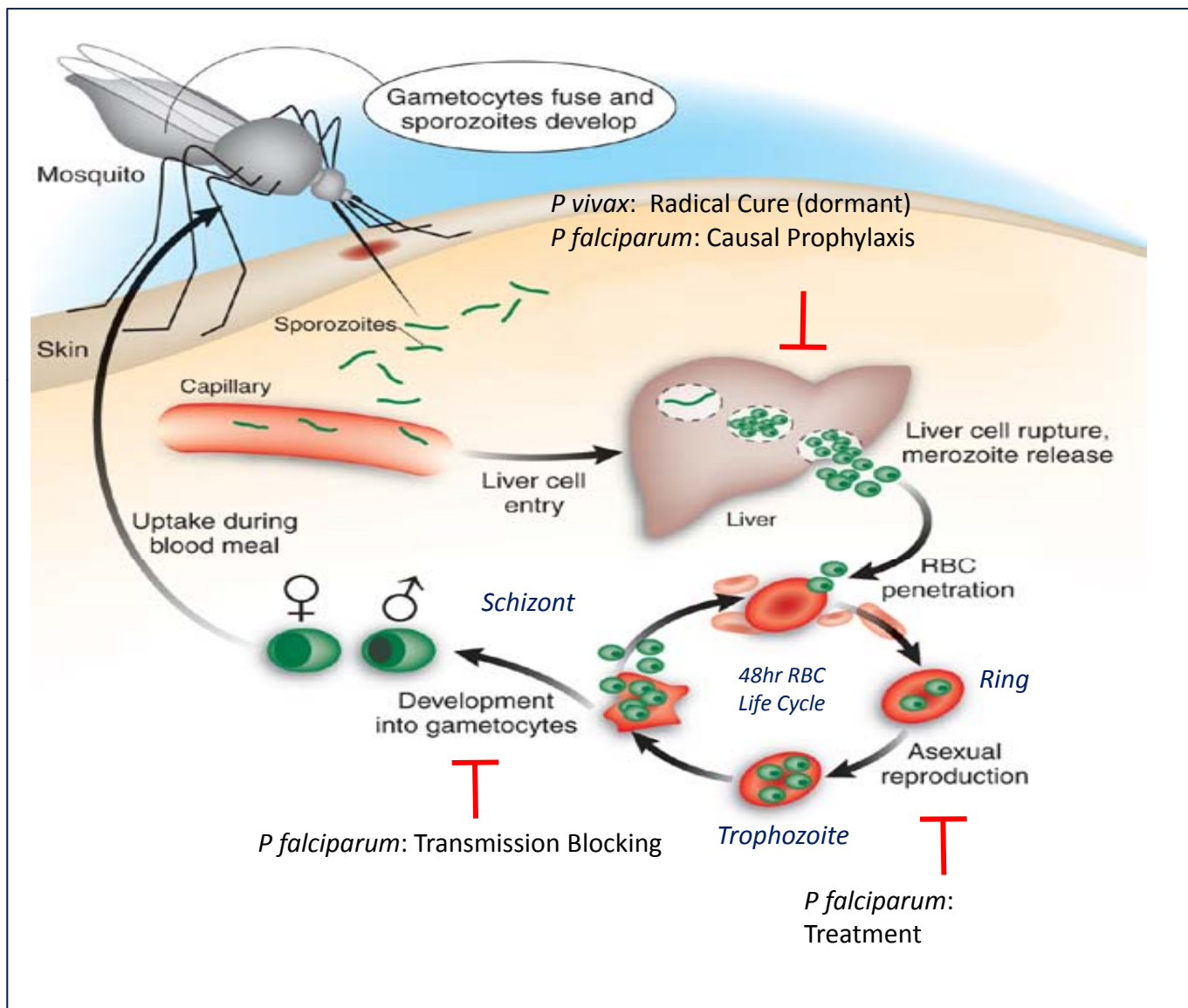
Growing resistance demands NCEs

- 500 million cases annually worldwide
- At least 1 million deaths per year, >75% under age of 5
- Resistance to current therapies widespread with exception of artemisinin derivatives
- Current artemisinin-based combination therapies (ACTs) contraindicated in 1st trimester



Drug resistance to marketed drugs widespread...

Complex Life-Cycle of the Parasite



Scope and Scientific Approaches

GNF scope is to use technology to develop novel assays with high throughput as well as translational component to develop novel antimalarial chemotypes through multiple parallel approaches

- **Cell-based screen approach**

- Majority of anti-infectives discovered through cell-based screening by facilitating parallel interrogation of druggable targets and also addresses compound permeability issues
- HTS on >2M compounds: *P. falciparum* infected human RBCs (proliferation inhibition)
- Liver-stage infection assay (*P. vivax* infection)

- **Target identification methods**

- Lab-evolved resistant strains upon compound treatment
- Tiling array analysis and whole-genome sequencing to help MoA determination
- Affinity chromatography/proteomics analysis

- **Target-based screen approach**

- Plasmodium kinases: such as CDPK1, CDPK5, GSK3, CK2 α
- Additional biochemical targets screening at GNF on cell-active compounds

The NGBS consortium

- **Novartis Institute for Tropical Diseases (NITD)**, Singapore
 - Team leaders: Bryan Yeung, Zou Bin, Christophe Bodenreider
- **Genomic Institute of the Novartis Research Foundation (GNF)**
 - Team leaders: **Arnab Chatterjee, Kelli Kuhlen and Elizabeth Winzeler**
- **Biomedical Primate Research Center (BPRC)**, Rijswijk (NL)
 - Team leader: Clemens Kocken
- **Swiss Tropical Institute (STI)**, Basel (CH) (now named Swiss TPH)
 - Team leader: Matthias Rottmann

- **The NGBS (NITD, GNF, BPRC, STI) consortium**
- Program Head: Thierry Diagana (NITD)

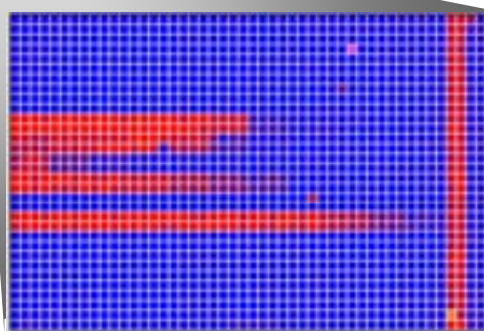
- NGBS is funded by:



GNF Malaria Cell-based Screening Summary



- > 2M compounds screened at 1.25 μM (3d7)
- 4851 hits < 1.2 μM EC_{50} vs W2 and/or 3d7
 - “Malaria Box” public resource for malaria research community
 - <https://www.ebi.ac.uk/chembl/db/index.php/compound>
- 1,256 < 200 nM EC_{50} vs 3d7
- >200 scaffolds represented in the reconfirmed compound set



**Tremendous amount of
chemical diversity from
screen**

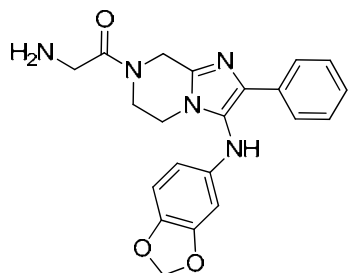
HTS reported in: Plouffe and co-workers *PNAS* **2008**, 9059.

Selection criteria for cell-based HLO at GNF

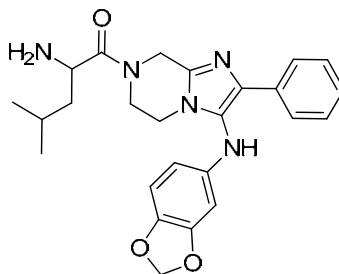
- ~5K reconfirmed blood-stage hits from HTS identified for further evaluation were determined by
 - Less than 1.25 μM EC_{50} with activity in 15 stain resistance panel
 - Less than 5-fold potency shift between strains
 - High selectivity in 6-cell line toxicity panel (SI > 20-fold)
 - Novel chemotype for malaria
 - Ease of synthesis (less than 7 steps)
 - Good solubility, metabolic stability and limited CYP450 inhibition
 - Multiple actives within a scaffold (if library diversity allows)
 - OPI algorithm used for clustering (developed at GNF) Yan, *et al. J. Chem. Inf. Model.* **2005** 45, 1784 -1790
 - Integrates SAR information into analysis of HTS data to prioritize interesting singletons as well as lower activity compounds that have good SAR from the screen

This presentation will focus on 1 series prioritized from this clustering

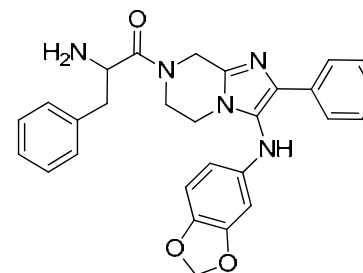
Imidazolopiperazines Hit series



GNF-Pf-5069



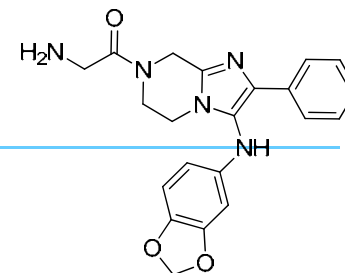
GNF-Pf-5179



GNF-Pf-5466

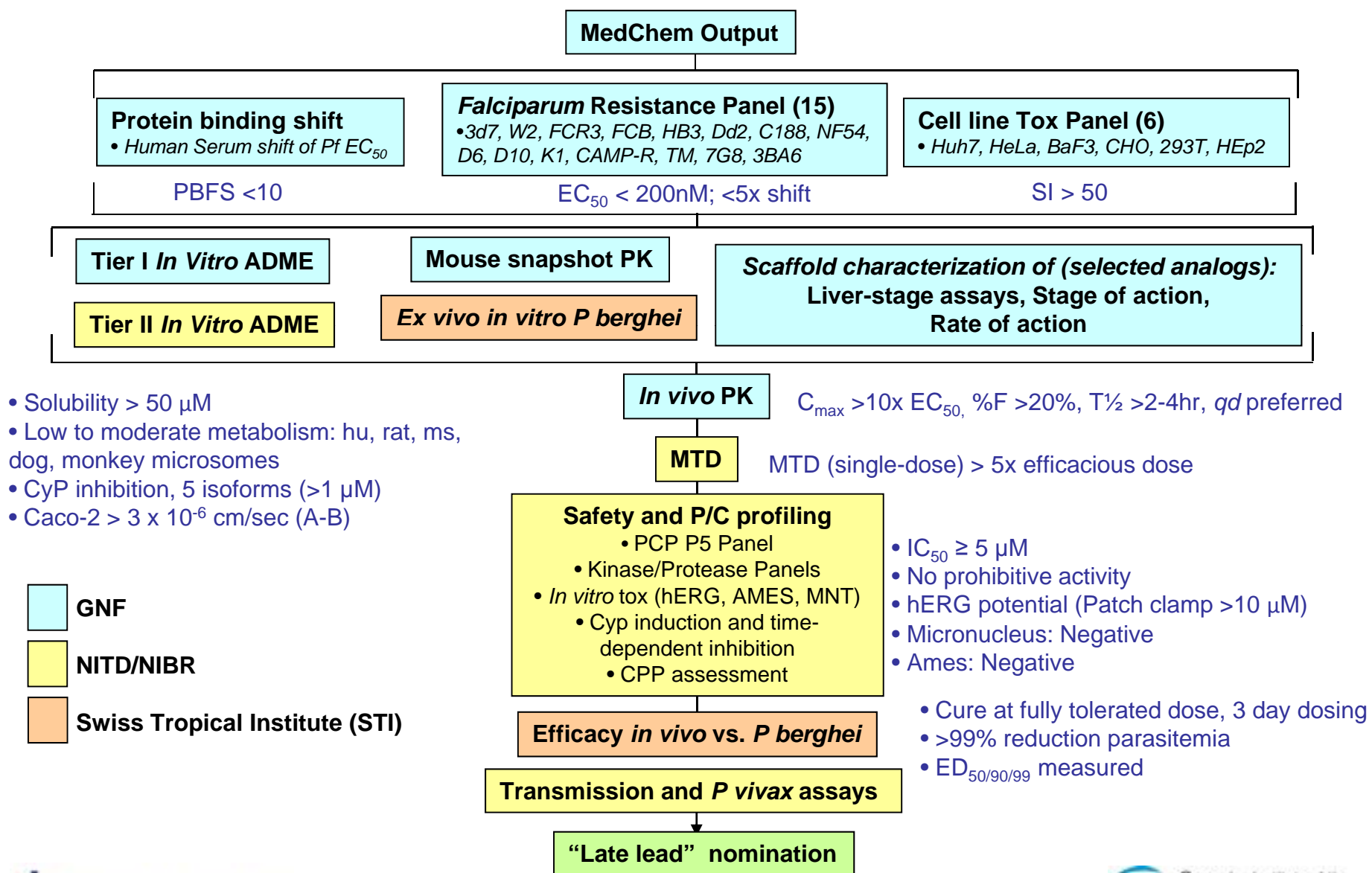
EBI ID	HTS W2 EC ₅₀ (μ M)	HTS 3D7 EC ₅₀ (μ M)	HTS Huh7 EC ₅₀ (μ M)	Powder W2 EC ₅₀ (μ M)	Powder 3D7 EC ₅₀ (μ M)	Powder Huh7 EC ₅₀ (μ M)
GNF-Pf-5069	0.097	0.063	>10	0.198, 0.473	0.161, 0.46	>100
GNF-Pf-5179	0.271	0.235	>10	0.122	0.119	>79
GNF-Pf-5466	0.119	0.116	>10	0.029	0.030	42.82

Imidazolopiperazines Hit Series

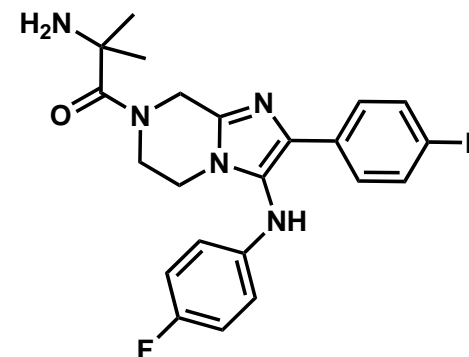
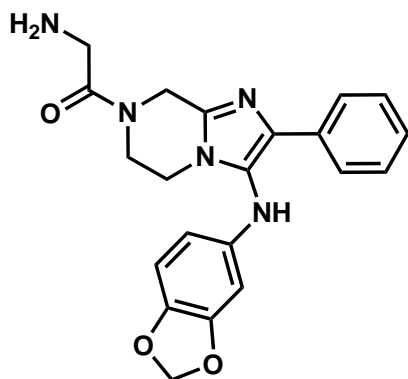


- Strengths
 - Unique scaffold for malaria, no biological activity annotation with structure
 - 4 step synthesis
 - Very good solubility; no activity on Cyp450s
 - Not identified in >100 HTS screens performed at GNF
- Issues
 - Moderate *in vitro* potency
 - *In vivo* snapshot PK: poor oral exposure
 - Likely due to catechol and glycinamide
 - hERG inhibition (35% at 10 μ M, patch clamp)

Compound Progression Scheme



Hit-to-lead optimization Summary



- GNF-Pf-5069
 - *Pf* 3D7 (EC_{50}): 460 nM
 - hERG (binding) IC_{50} = 19 μ M
 - Mouse metabolic stability (ER): 0.24
 - Solubility (pH 6.8): > 175 μ M
 - PO C_{max} (D.N.) = 16 nM / (mg/kg)
 - PO AUC_{0-5h} (D.N.) = 48.6 h*nM / (mg/kg)Poor exposure limited any further work

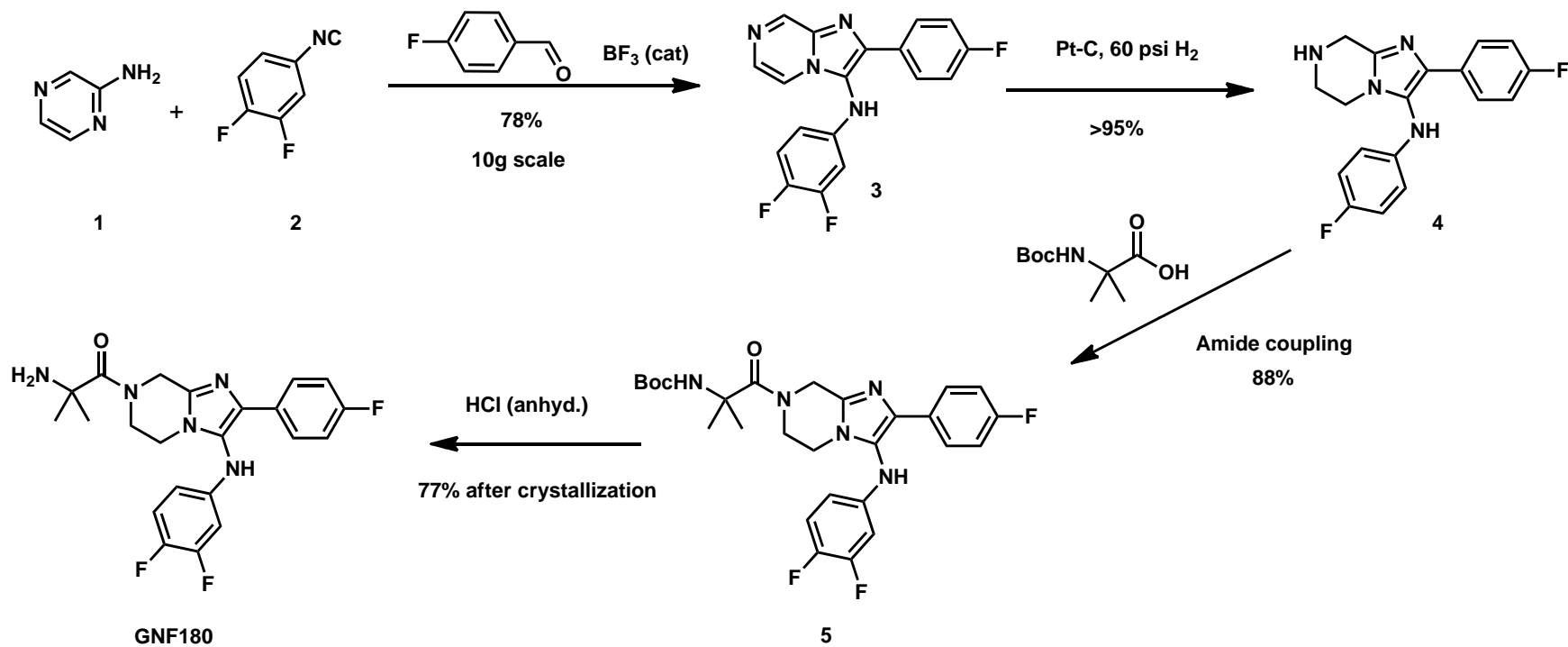
- Compound GNF776
 - *Pf* 3D7 (EC_{50}): 20 nM
 - hERG (binding) IC_{50} = 3.95 μ M
 - CL = 60 mL/min/kg; V_{ss} = 23.7 L/kg; $T_{1/2}$ = 6.2 h
 - PO C_{max} (D.N.) = 60 nM / (mg/kg)
 - PO AUC_{0-inf} (D.N.) = 596.5 h*nM / (mg/kg)
 - F (%) = 87
 - Efficacy achieved in *p berghei* mouse model

(*D.N.: dose normalized)

Still need to improve hERG and potency

J. Med. Chem.
2011, 54, 5116.

Groebke-Blackburn Cyclization Route

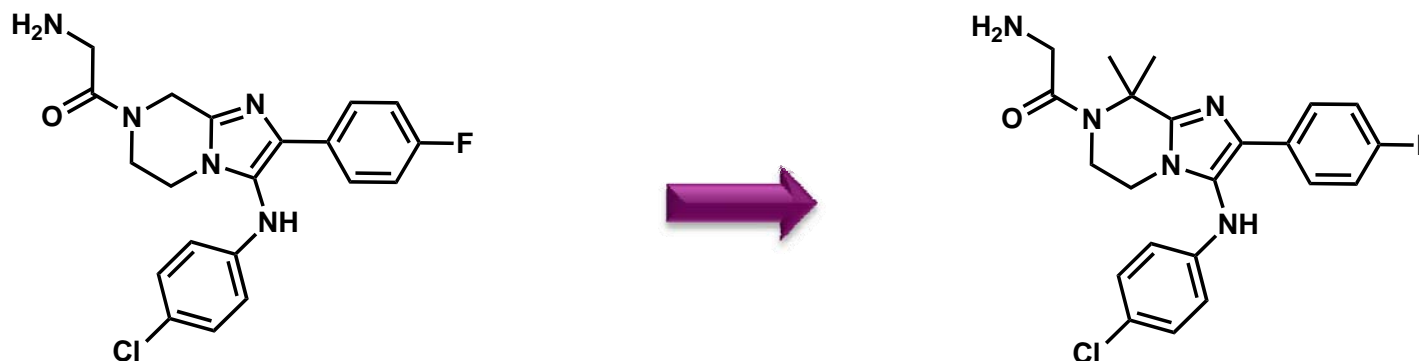


P falciparum 3D7
EC₅₀ = 30 nM

- 4 steps and 53% overall yield
- No column chromatography

Imidazolopiperazine Core Optimization

8,8-dimethyl substitution significantly improves profile



- GNF268
 - PfW2 (EC₅₀): 9 nM
 - hERG (binding) IC₅₀ = 6.6 μM
 - CL = 1179 mL/min/kg; V_{ss} = 106.76 L/kg
 - Oral T_{1/2} = 2.2 h
 - PO C_{max} (D.N.) = 3.3 nM / (mg/kg)
 - PO AUC_{inf} (D.N.) = 30 hr*nM
 - F (%) = 17
 - 30x cross-resistance to GNF-Pf-5069 lab evolved resistance strain

(*D.N.: dose normalized)

- GNF179
 - PfW2 (EC₅₀): 6 nM
 - hERG (binding) IC₅₀ = 7.2 μM
 - CL = 21.92 mL/min/kg; V_{ss} = 11.8 L/kg
 - Oral T_{1/2} = 8.4 h
 - PO C_{max} (D.N.) = 60.5 nM / (mg/kg)
 - PO AUC_{inf} (D.N.) = 1035 hr*nM (mg/kg)
 - F (%) = 58
 - 7x cross-resistance to GNF-Pf-5069 lab evolved resistance strain

gem-Dimethyl group on piperazine improves potency and PK in mice

SAR Summary

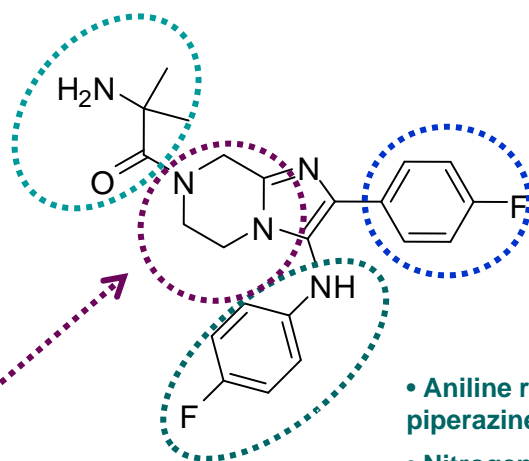
- Optimization of the Imidazolopiperazine scaffold:
 - Introduction of 8,8-dimethylgroup on piperazine improves *in vitro* and *in vivo* potency by 10-fold
 - Required glycine functionality is primary cause for hERG activity

Amino acid SAR

- NH₂ preferred; but also can use NHR, NR₂, OH and alkyl groups
- 1-2 carbon spacer to NH₂ preferred
- Dimethylglycine improves *in vivo* PK relative to glycine (mouse and rat)
- Glycine least active on hERG (2-3 fold), while all other amino acids in similar range

Piperazine ring SAR

- 8-substitution helps with potency and mouse exposure (dimethyl > valine, alanine > hydrogen)
- 5-dimethyl and 6-dimethyl moderately helps potency, no real effect on PK
- Cyclic lactams (6-oxo) derivatives active with 8,8-dimethyl substitution and reduces metabolism



- Aryl required
- *Para* and *meta* substituted aromatic preferred
- *Ortho* substituted aromatic not preferred
- Strong electron-withdrawing group not preferred

- Aniline required with unsubstituted piperazine series
- Nitrogen required, but N-H is not in unsubstituted piperazines
- Variety of aromatics help potency (*para* and *meta* preferred)
- Heteroaromatics and cycloaliphatics not active in unsubstituted piperazine series, but active with 8-dimethyl groups
- Can modestly effect hERG activity

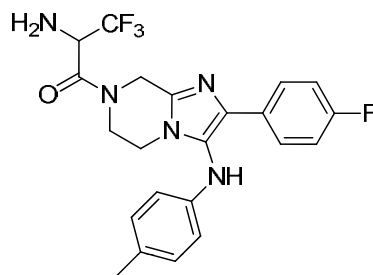
hERG Optimization

Targeting the amine functionality

Modulate amine basicity:

pKa = 3.3
Glycine amine pKa = 7.5

GNF824

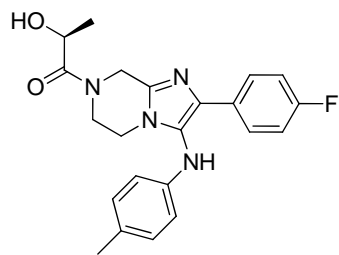


P. falciparum 3D7 EC₅₀ = 50 nM

hERG (Dofetilide binding)
IC₅₀ = 12.47 μM

Solubility (pH 6.8) = 34 mg/L

Replacement of amine with alcohol:



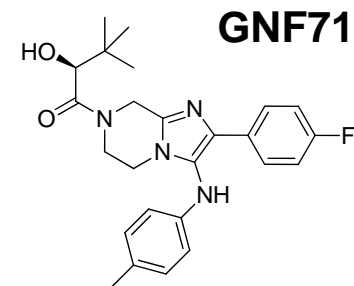
GNF557

P. falciparum 3D7 EC₅₀ = 300 nM

hERG (Dofetilide binding)
IC₅₀ > 30 μM

Solubility (pH 6.8) = 110 mg/L

Optimize
P. falciparum
potency

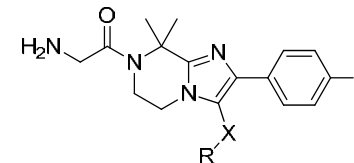


GNF711

P. falciparum 3D7 EC₅₀
= 20 nM

Unable to progress due to poor metabolic stability and Log D

SAR of the 3-position (aniline group)



Cmpd	X	R	<i>Pf</i> 3D7 EC ₅₀ (nM)	pH 6.8 Solubility (μM)	Rat ER	hERG (binding) (μM)
GNF156	NH	4-F-Ph	10	>1000	0.52	6.4
GNF877	CH ₂	4-F-Ph	8	>1000	0.49	2.7
GNF767	O	4-F-Ph	3	>175	0.13	4.0
GNF765	S	4-Me-Ph	8	42	ND	4.2
GNF772	SO	4-Me-Ph	42	>175	0.37	26.0
GNF771	SO ₂	4-Me-Ph	5210 (NF54)	ND	ND	ND
GNF713	SO	3-F-Ph	481	ND	ND	12.0
GNF714	SO	4-F-Ph	70	ND	ND	>30
GNF715	SO	3,5-Me ₂ - Ph	381	ND	ND	15.0
GNF716	SO	3,4-F ₂ -Ph	89	ND	ND	27.0

Sulfoxide linkage is a good balance of potency, solubility and hERG

Profile of Preclinical candidate: GNF156

- ***In vitro* Activity Profile of Imidazolopiperazines**

- *P. falciparum* W2 potency: EC₅₀ = 0.006 μM
- Compound exhibits < 5-fold shift across 15 strains
- Selectivity vs. Huh7 cells: >10,000 fold
- Complete transmission blocking at 500 nM

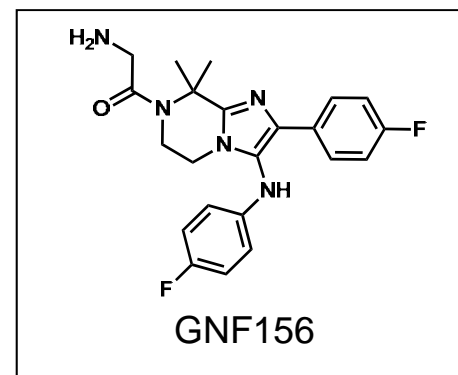
- ***In vivo* Profile**

- Good oral PK/bioavailability and exposure; T_{1/2} > 4 h
- *In vivo* efficacy in *P. berghei* mouse model: ED₉₉ = 2.2 mg/kg
- Causal prophylaxis data: Fully protective at 20 mg/kg

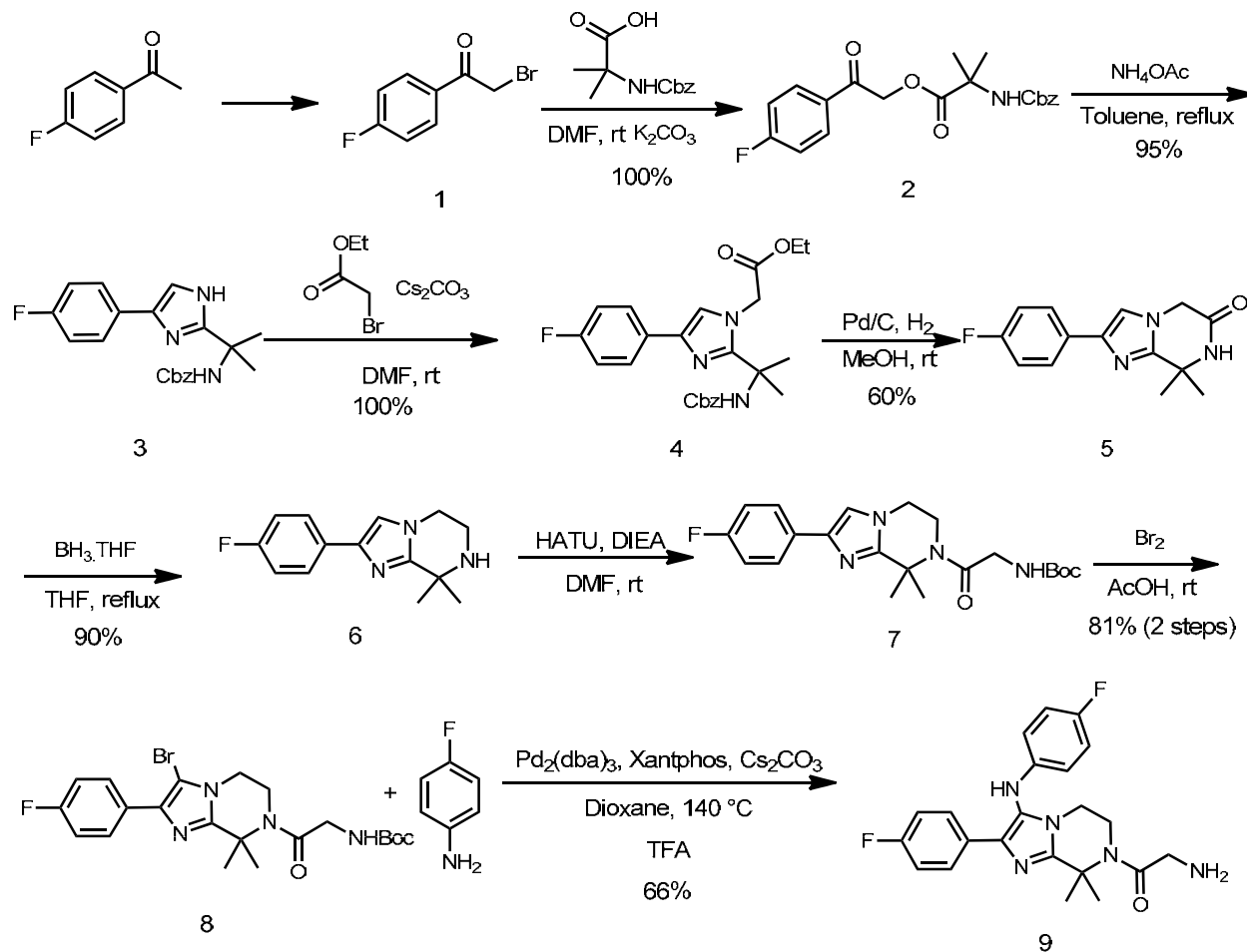
- **ADME/Safety/Physicochemical Profile**

- ADMET: solubility: >1000 μM @ pH1 and @ pH 6.8; no GSH trapping; no Cyp induction or phototoxicity flags
- Clean PCP receptor panel, Ames, MNT and good metabolic stability
- Formulation suitable for tox studies identified (35x exposure multiples achieved in rats and 16x in dogs)
- hERG activity at 6.6 μM in binding assay; 13 μM in automated patch clamp assay

- **Unique activity for blood/liver-stage infections and transmission blocking**



GNF156: Current Synthesis Route



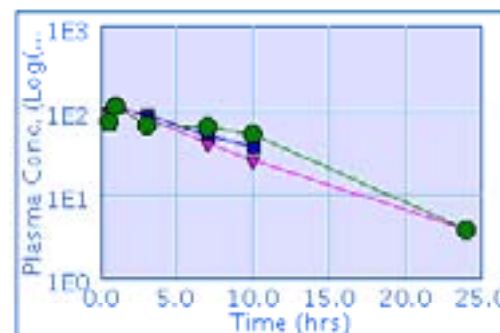
- 10 step route, but minimal purifications
- 27% overall yield
- Low cost of goods for starting materials

GNF156: *In vivo* efficacy in *P. berghei* model

Compound	# of animals per dose	ED ₅₀ (mg/kg)	ED ₉₀ (mg/kg)	ED ₉₉ (mg/kg)
Chloroquine	5	1.9	5.2	8.4
Mefloquine	5	3.8	5.0	8.6
Artesunate	5	5.0	20.5	119
Imidazolopiperazine GNF156	5	0.48	1.13	2.2

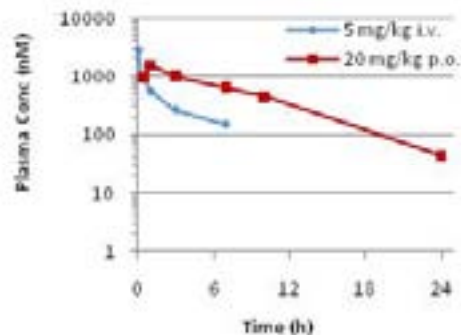
Oral Exposure in mouse at efficacious dose (2 mg/kg)

$C_{\max} = 112 \pm 5 \text{ nM}$
 $T_{\max} = 1 \text{ h}$
 $T_{1/2} = 4.8 \pm 0.8 \text{ h}$
 $AUC_{\text{inf}} = 945 \pm 110 \text{ nM}\cdot\text{h}$
 $\%F = 32 \pm 4$

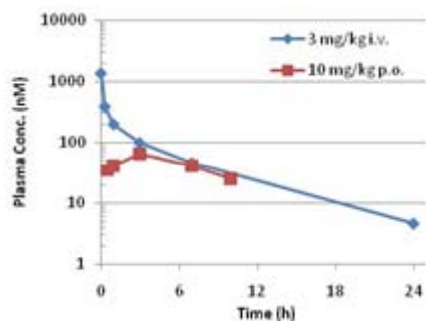


GNF156 PK Properties in Mouse, Rat and Dog

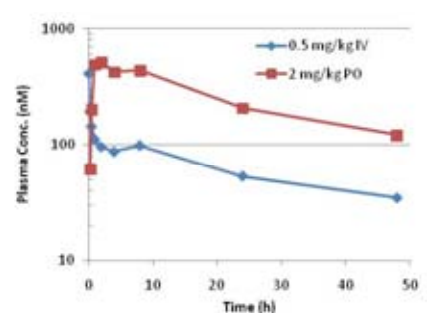
GNF156 Single Dose PK in Mice



GNF156 Single Dose PK in Rat



GNF156 Single Dose PK in Dog



Mouse PK (5 / 20 mg/kg, n=3)	
CL (ml / min•kg)	49 (mod)
V _{ss} (l/kg)	10 (high)
t _{1/2term.} (h)	6.7
AUC (h•nM) p.o.	12156
C _{max} (nM) p.o.	1539
T _{max} p.o. (h)	1.0
Oral BA (%F)	74

Rat PK (3 / 10 mg/kg, n=3)	
CL (ml / min•kg)	24 (low)**
V _{ss} (l/kg)	26 (high)
t _{1/2term.} (h)	4.9
AUC (h•nM) p.o.	975
C _{max} (nM) p.o.	91
T _{max} p.o. (h)	1.6
Oral BA (%F)	20

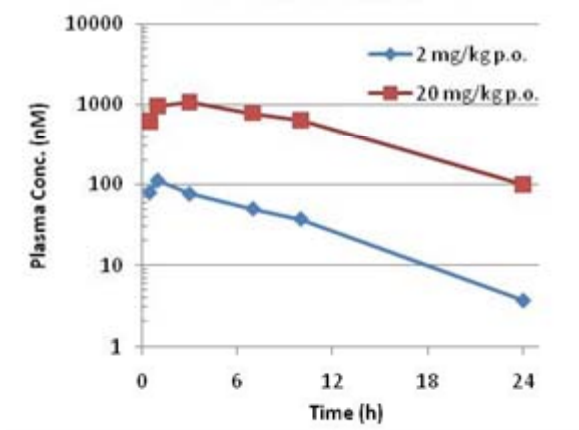
** Adjusted for RBC partitioning

Dog PK (0.5 / 2 mg/kg, n=3)	
CL (ml / min•kg)	5 (low)
V _{ss} (l/kg)	11 (high)
t _{1/2term.} (h)	28.5
AUC (h•nM) p.o.	16142
C _{max} (nM) p.o.	553
T _{max} p.o. (h)	1.7
Oral BA (%F)	89

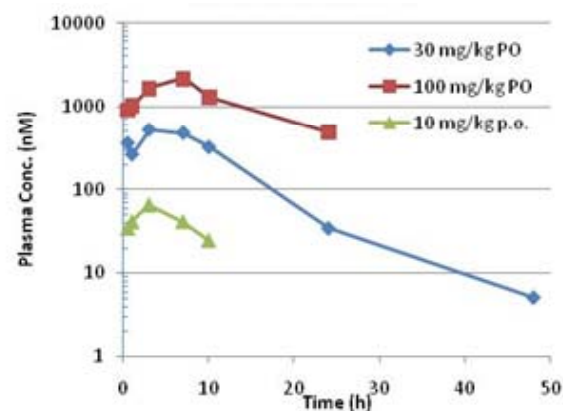
- In rat whole blood, GNF156 showed moderate RBC partitioning with blood to plasma ratio of 3.5.
- GNF156 exhibits low to moderate blood CL, high V_{ss}, and reasonable bioavailability across species
- Exposure multiple of 16 achieved in dogs (AUC_{eff} ~1000 h•nM at 2 mg/kg in mice). Long T_{1/2} in dogs.

GNF156: Dose Linearity PK in Mouse and Rat

Mean Mouse Plasma Conc.



Mean Rat Plasma Conc



Formulation	Mouse		Rat		
	Solution		Solution		Suspension
Dose PO (mg/kg)	2	20	10	30	100
$t_{1/2}$, po (h)	4.9	4.4	4.7	6.2	8.4
AUC (h•nM) p.o.	945	12156	975	7338	34885
AUC fold change		12.9		11.6	35.8
C_{max} (nM)	112	1539	91	581	2233
C_{max} fold change		13.7		6.4	24.5
F (%)	32	74	20	40	57

- AUC increase more than dose proportional in mice
- AUC and C_{max} increase more than dose proportional in rats
- Exposure multiple of 35 achieved in rats at 100 mg/kg ($AUC_{eff} \sim 1000 \text{ h} \cdot \text{nM}$ at 2 mg/kg in mice)

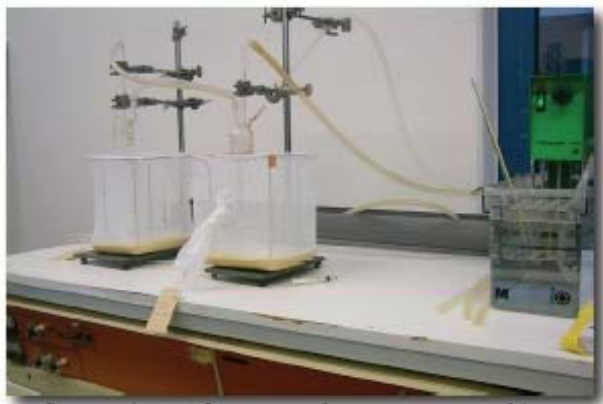
GNF156: Human efficacious dose projections

Species	Units	Mice	Rat	Dog	Human
Body weight	kg	0.025	0.25	8.7	70
Dose	mg/kg	5	3	0.5	1
Vss	L/kg	10.2	25.7	10.9	10.2 to 25.7
CL	mL/min/kg	49.2	84.4	4.8	0.9 to 4.5
T1/2	h	2	4.9	28.5	14.5 to 71
F	%	66	47	89	67

- Efficacy : ED₉₉ in mice for KAF156 is 2.2 mg/kg.
- Bioavailability : F was estimated to be 67% in humans
- Clearance : 0.9 to 4.5 ml/min/kg using various methods.
- Human efficacious dose (to show similar efficacy to the mouse 99% parasitemia reduction) was predicted to be **2.3 to 48mg** based on the following assumption
 - **Efficacy is similar in P.b infected mice and P.f infected human**
 - **PK is similar in normal mice and P.b infected mice**

Imidazolopiperazines Block Transmission

Novel activity for a potent blood-stage antimalarial



Stage V gametocyte transmission mosquito feeding experimental procedure:

1. The compounds are added to a bloodmeal with NF54 gametocytes
2. Six days after the fed mosquitoes are dissected and checked for oocysts
3. Compounds can have an effect on mosquitoes, therefore the mortality of the mosquitoes is checked every day

	% infected mosquitoes	# Oocysts/ mosquito
Control	95	49
GNF776 (0.7 μ M)	60	1.5
GNF776 (7 μ M)	0	0

Data furnished by Sauerwein lab

Conclusions and path forward:

- GNF776 (lead cmpd) inhibits oocyst development dose-dependently
- Optimized GNF156 blocks transmission completely at 500 nM
- This novel activity is critical for potential eradication goals

GNF156 preclinical safety summary

No significant tox at up to 30-45 (rat) or 135-250 (dog) fold over ED₉₉ exposure

- **Genotoxicity**

Mini Ames mutagenicity assay: **negative**

- **Phototoxicity**

3T3 NRU assay: **negative** (PIF = 1.4)

- **2-week rat toxicity at 10, 30, and 100 mg/kg/day**

- 100 mg/kg/day: salivation, respiratory distress, ↑ lymphocyte counts and WBC, mild thyroid follicular hypertrophy/hyperplasia, and possibly testicular degeneration (1/5 rats but not in controls)

≥30 mg/kg/day: ↑ platelets (17-20%), in 3/5 rats at 30 and all rats at 100 mg/kg/day, mild-mod ↑ creatinine (up to 2-fold); mild (15%) ↑ globulin

All doses: ↓ CK (-16- 56%) and glucose (-6 - 15%) in several rats per group

- **Dog rising dose with telemetry at 2, 10, 30, 100, 250 mg/kg**

250 mg/kg: Transient heart rate increase without ECG changes

Lessons learned

- Molecular weights greatly effect the cellular optimization
 - Forces the chemistry to be very atom economical (high ligand efficiencies)
- SAR can be challenging, but getting hits from HTS with good SAR is critical
 - Better to have SAR than absolute potencies off the deck
 - Potencies can be optimized relatively quickly compared to other parameters
- Weekly cytotoxicity, protein binding shift potency, and cross-resistance data to other scaffold lab-evolved resistance strains really allows for better optimization of other parameters

Acknowledgements (Research)

Chemistry

- Advait Nagle
- Tao Wu
- Tomoyo Sakata
- Robert Moreau
- Jason Roland
- Pranab Mishra
- David Tully
- Valentina Molteni

Biology

- Kelli Kuhen
- Carolyn Francek
- Zhong Chen
- Kerstin Henson
- Rachel Borboa
- James Gilligan
- Tae-gyu Nam
- Neekesh Dharia
- David Plouffe
- Case McNamara
- Stefan Meister
- Elizabeth Winzeler

Pharmacology/Analytical

- Tove Tuntland
- Perry Gordon
- Jonathan Chang
- Matthew Zimmerman
- Liang Wang
- Todd Groessl
- Barbara Saechao
- Bo Liu
- Chun Li
- David Jones
- Wendy Richmond
- Kevin Johnson
- Tom Hollenbeck
- Lucas Westling
- Michael Kwok
- Tiffany Chuan
- John Isbell

Informatics

- John Che
- Yingyao Zhou

Swiss Tropical and Public Health Institute

- Matthias Rottmann
- Christoph Fischli
- Sonja Maerki

NITD

- Bryan Yeung
- Zou Bin
- Anne Goh
- Suresh B. Lakshminarayana
- Veronique Dartois
- Thomas Keller
- Thierry Diagana

External collaborators

- Montip Gettayacamin (AFRIMS - Bangkok)
- Robert Sauerwein (NCMLS - The Netherlands)
- Ian Bathurst (MMV)

GNF Management

- Jennifer Taylor
- Richard Glynne
- Martin Seidel
- Peter Schultz

Acknowledgements (Development)

Core team

Margaret Weaver
Xingmei Han
Giancarlo Francese
Sreehari Babu
Rita Ramos
Karen Beltz
Bo Han
Wen Shieh
Markus Baenziger

Novartis Tropical Medicines

Heiner Grueninger
Anne-Claire Marrast
Paul Aliu
Gilbert Lefevre

Clinical team

Jens Praestgaard, Ruobing Li

External support and advice

Marcel Tanner, Swiss Tropical
Institute,
Nick White, Oxford University