

Micro scale-down technologies for high throughput development of chromatographic separations

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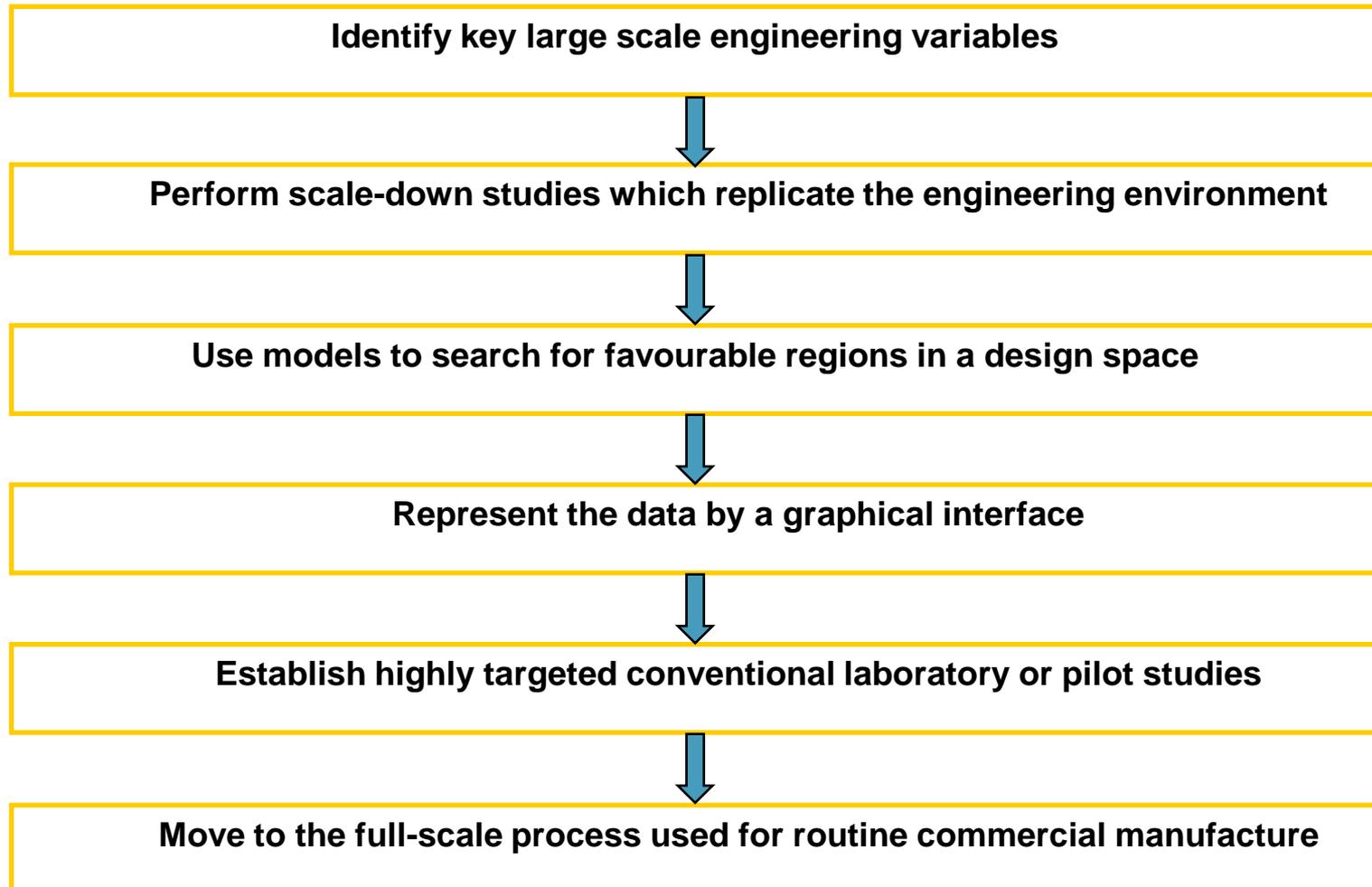
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 - UCL strategy for micro biochemical engineering
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What is micro biochemical engineering?

- Addresses limitations of the current development paradigm
- Combines scale-down assessment and large scale verification
- Reduces feed requirements
 - Operation at the millilitre-scale or smaller
- Uses automation as far as possible
 - Enables parallel experimentation

Micro biochemical engineering workflow



Challenges in chromatography development

- Many operating modes, chemistries and backbones
 - Affinity
 - Ion-exchange
 - Hydrophobic interaction
 - Multi-modal
 - Buffer selection
- Laboratory studies may require large feed volumes
- Many conflicting output metrics
 - Yield
 - Quality
 - Time
 - Economics

Micro-scale approach to chromatography

Micro-scale
(1 μ L – 5 mL resin)

Rapid scouting of large numbers of options and measurement of relative performance

Laboratory-scale
(5 – 100 mLs resin)

Evaluation of scale-translation effects and improved definition of a design space through a smaller experimental set

Pilot-scale
(0.1 – 1 L resin)

Verification of outputs and fine-tuning of operation for limited sets of optimal conditions identified earlier

Full-scale
(1 L+ resin)



Automation of microscale chromatography

- Can run many separations simultaneously
- Unattended operation
- Simplifies preparation and clean-up
- Reduces manual workload to a manageable level
 - Removes some potentially time consuming and labour intensive tasks
- Can integrate with other equipment
 - Orbital shakers
 - Solid–liquid separation devices
 - Plate readers

Improved process understanding

- Screen many different resins and buffer conditions
 - Dynamic binding capacity, yield and purity
- Obtain kinetic, equilibrium and breakthrough data
 - Study how feed concentration and residence time affect separations
- Improve process characterisation
 - QbD, validation and support greater post-approval process flexibility

Microscale chromatography formats



Microlitre batch
incubation

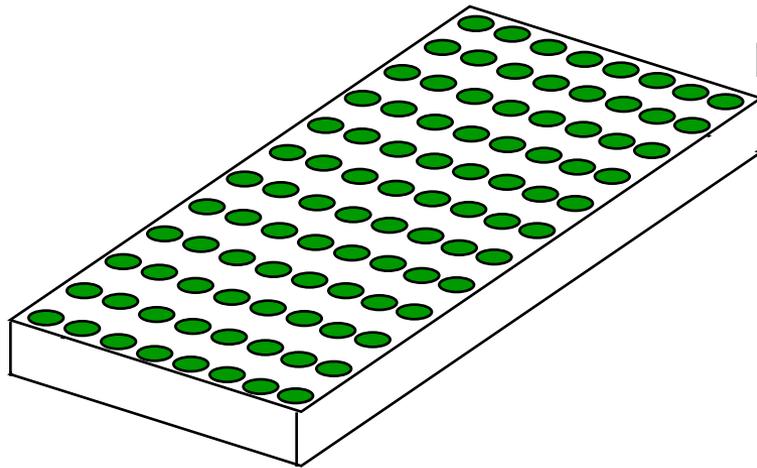


Miniature
columns

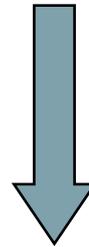


Pipette tip
chromatography

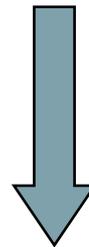
Microlitre batch incubation format



Filter plate containing up to a few hundred microlitres of matrix and incubated with feed material



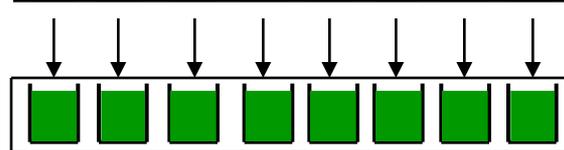
Agitate at a defined rate and for a defined time period



Centrifugation or
vacuum filtration

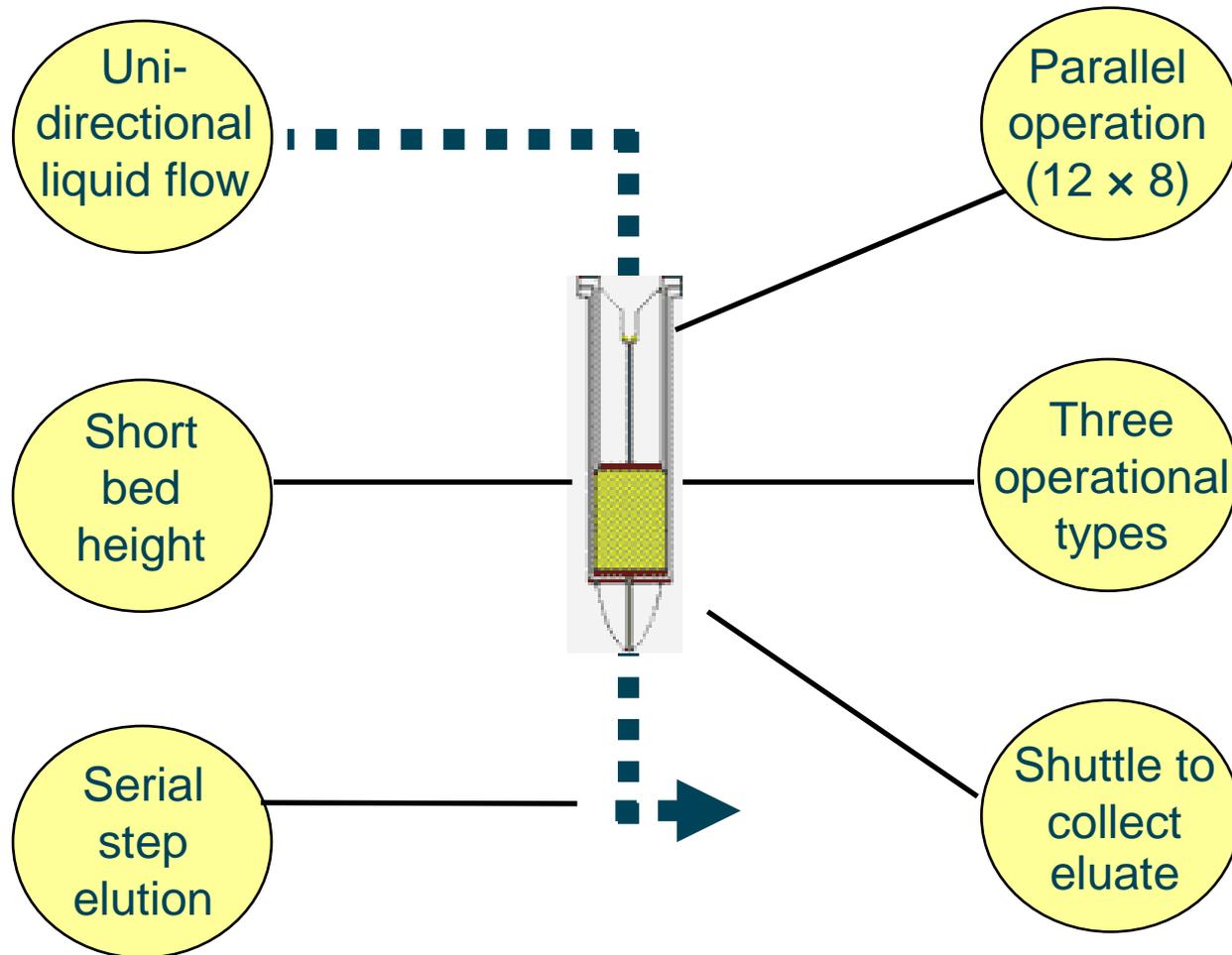


Resin containing filter-plate

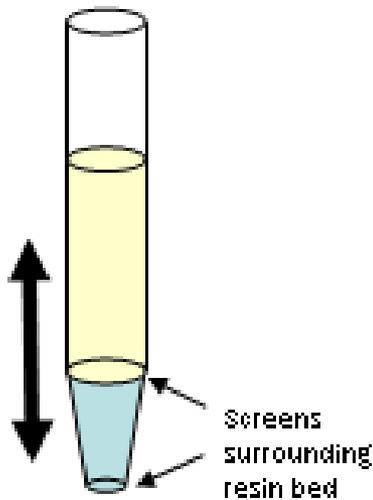


Collection plate

Miniature column format

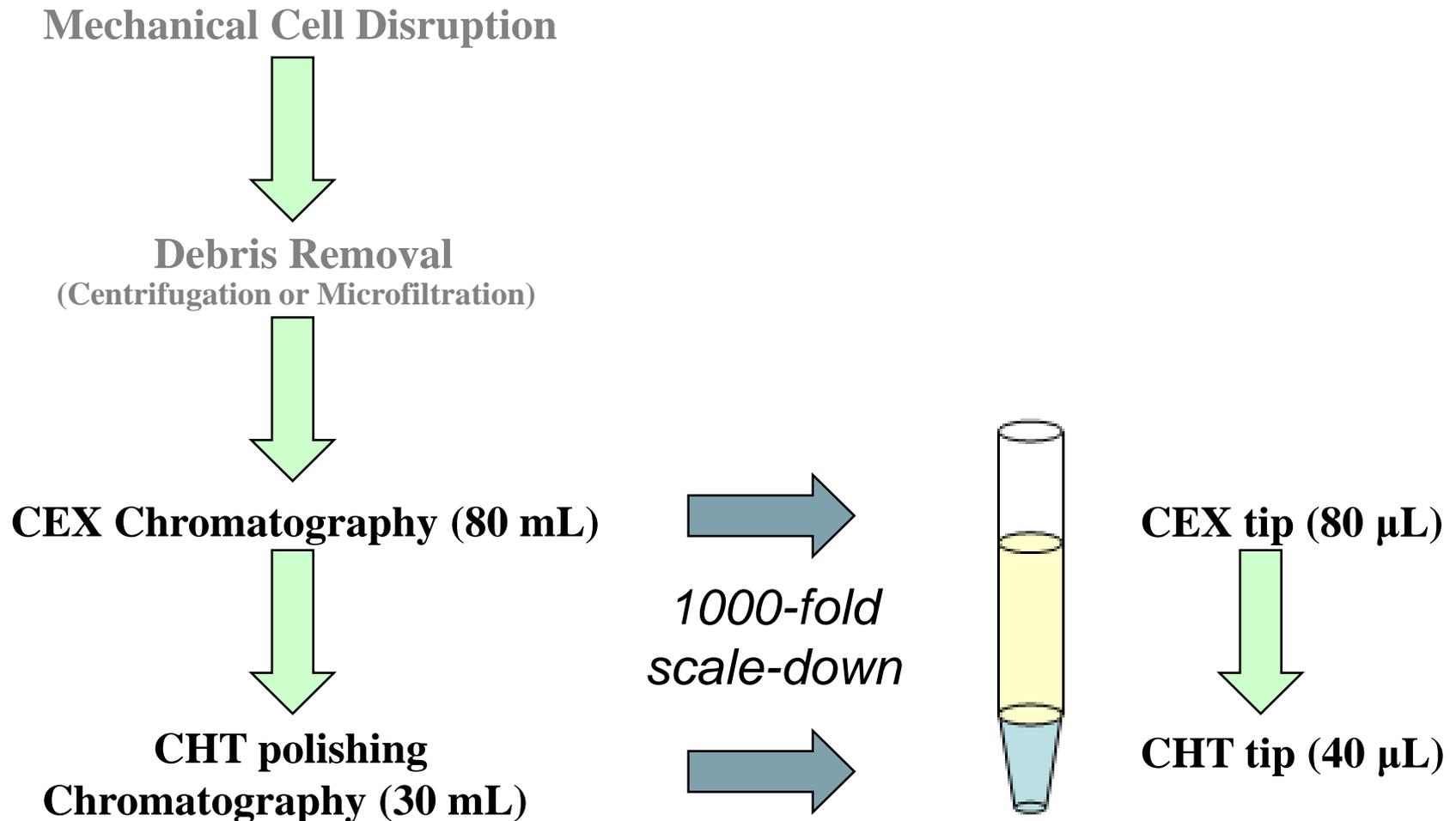


Pipette tip chromatography format

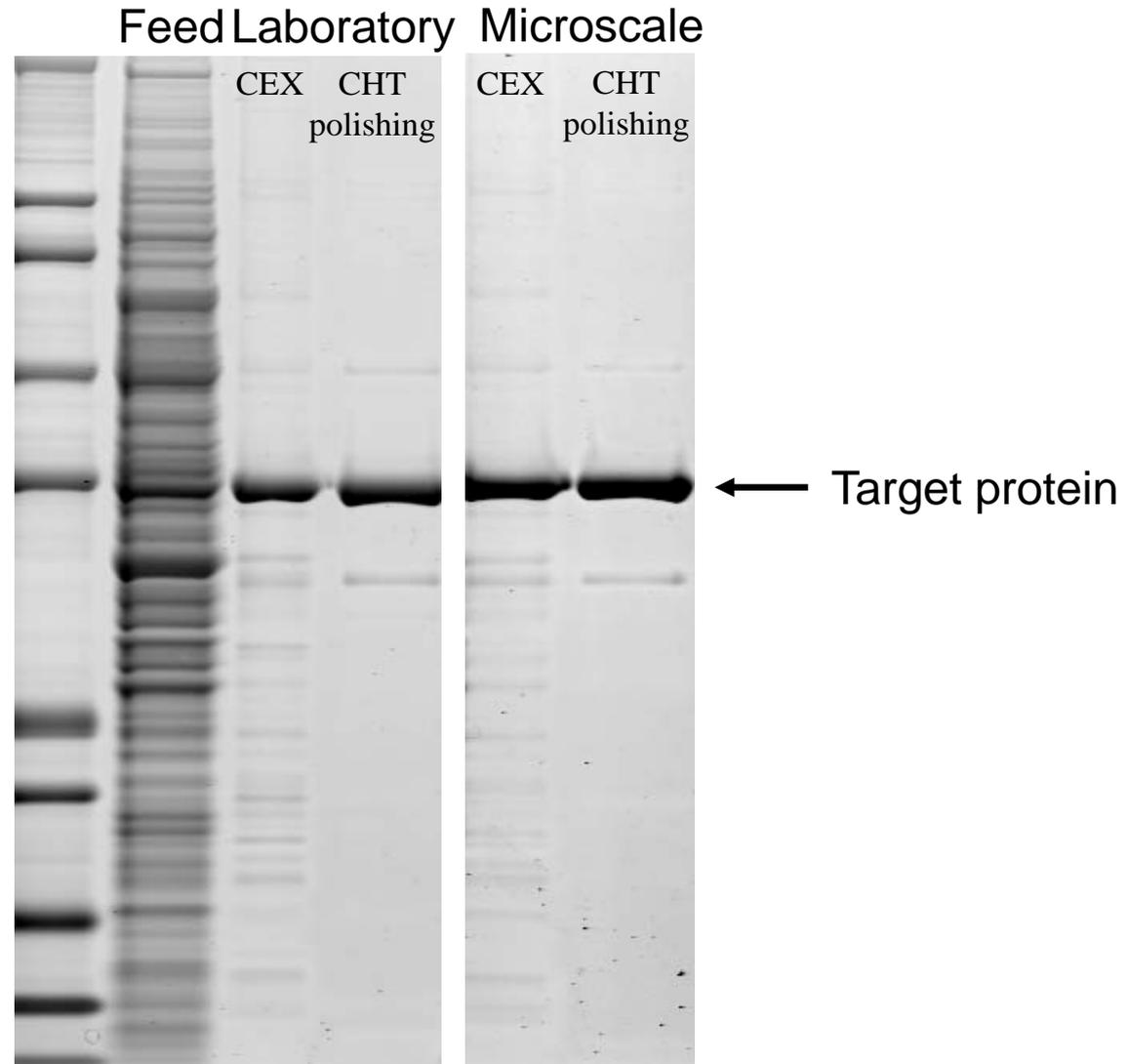


- Supplied pre-packed for any custom or off-the-shelf resin by PhyNexus
- Resin is packed into the base of the tip and held between two screens
- Feed or buffers are held in a 96-well plate
- These are aspirated and dispensed repeatedly in a bidirectional fashion for the required residence time
- Optimise robotic parameters
 - Flowrate (v)
 - Number of bidirectional cycles (N)

Pipette tip chromatography example 1: VLP purification (UCL and Merck)

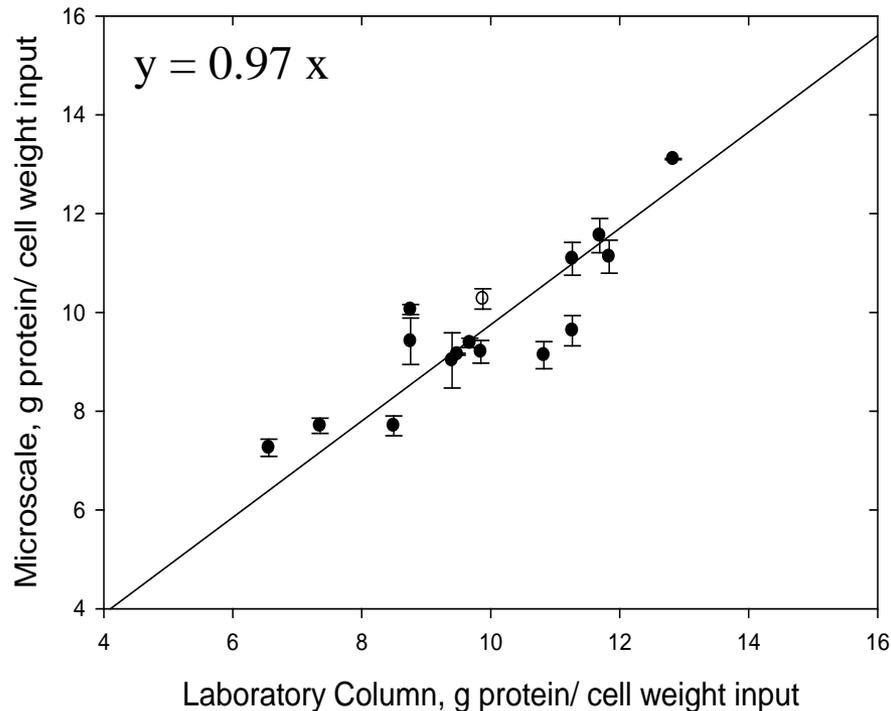


Comparability of microscale chromatography



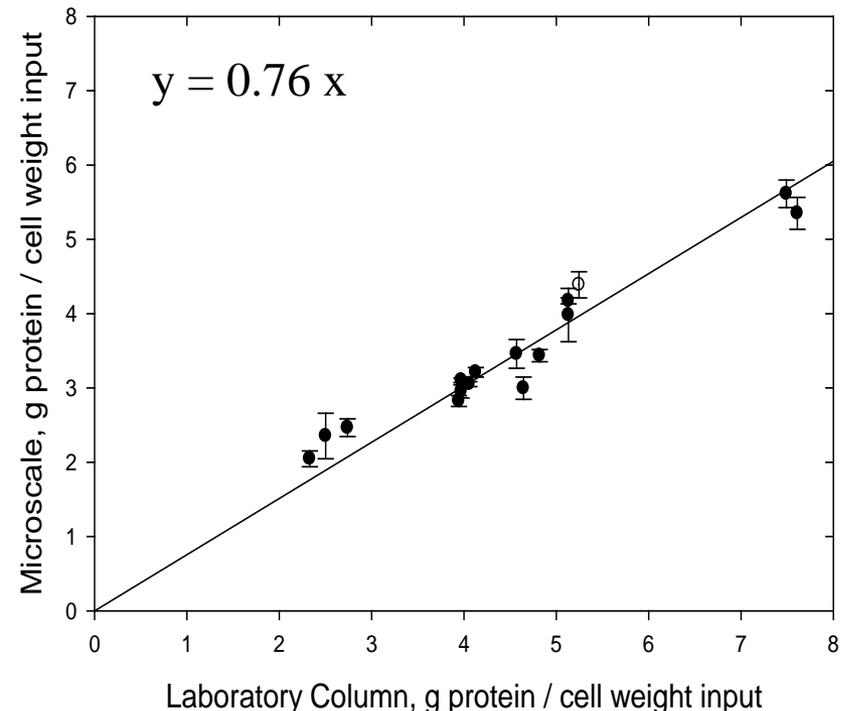
Correlation of microscale yield to lab scale

CEX Chromatography



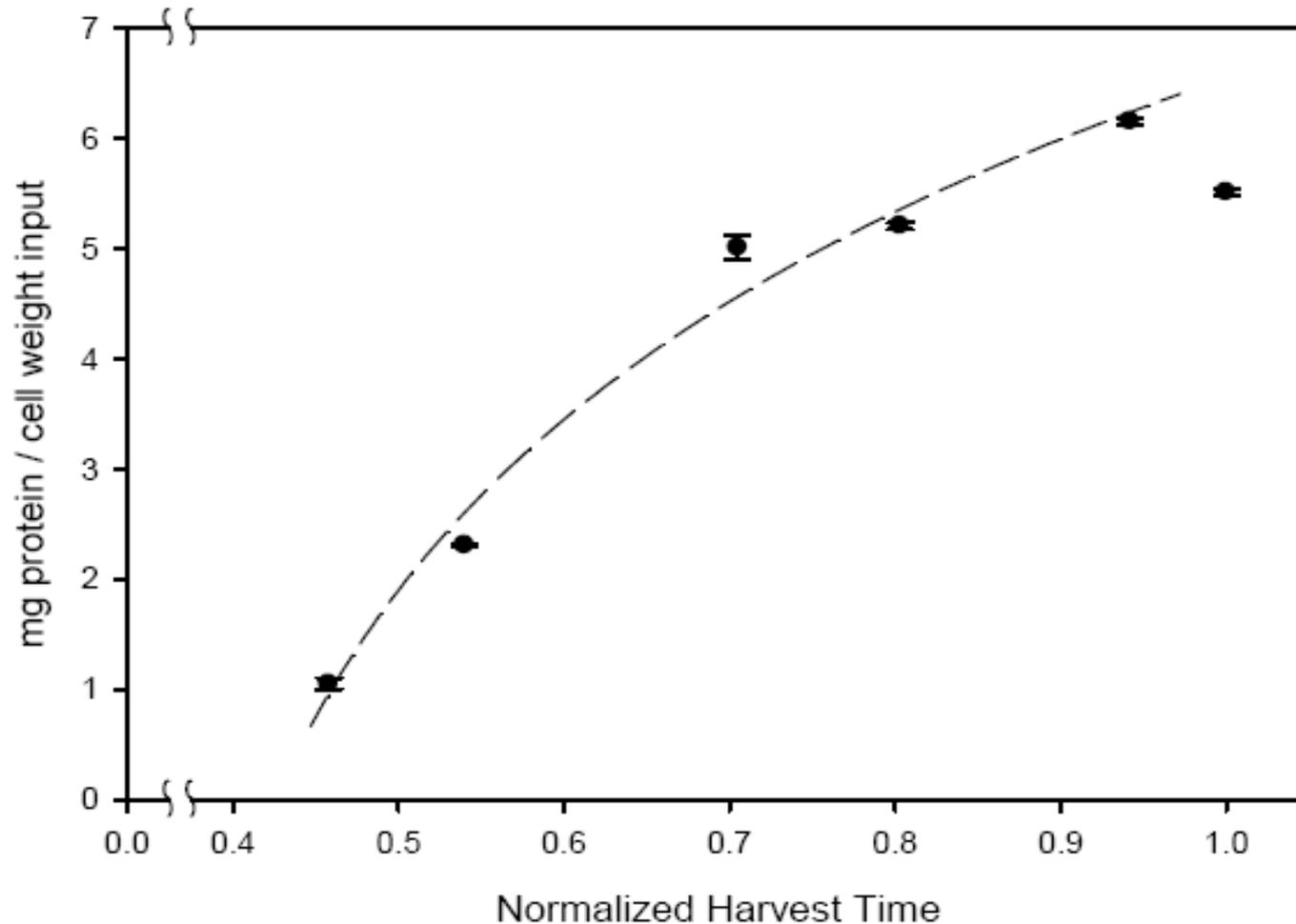
Strong one-to-one correlation between microscale and laboratory scale

Polishing Chromatography



Consistent offset following polishing chromatography allows use of a correction factor

Determination of Harvest Time



Wenger et al. (2007), An automated microscale chromatographic purification of virus-like particles as a strategy for process development, *Biotechnology and Applied Biochemistry*, 47, 131–139

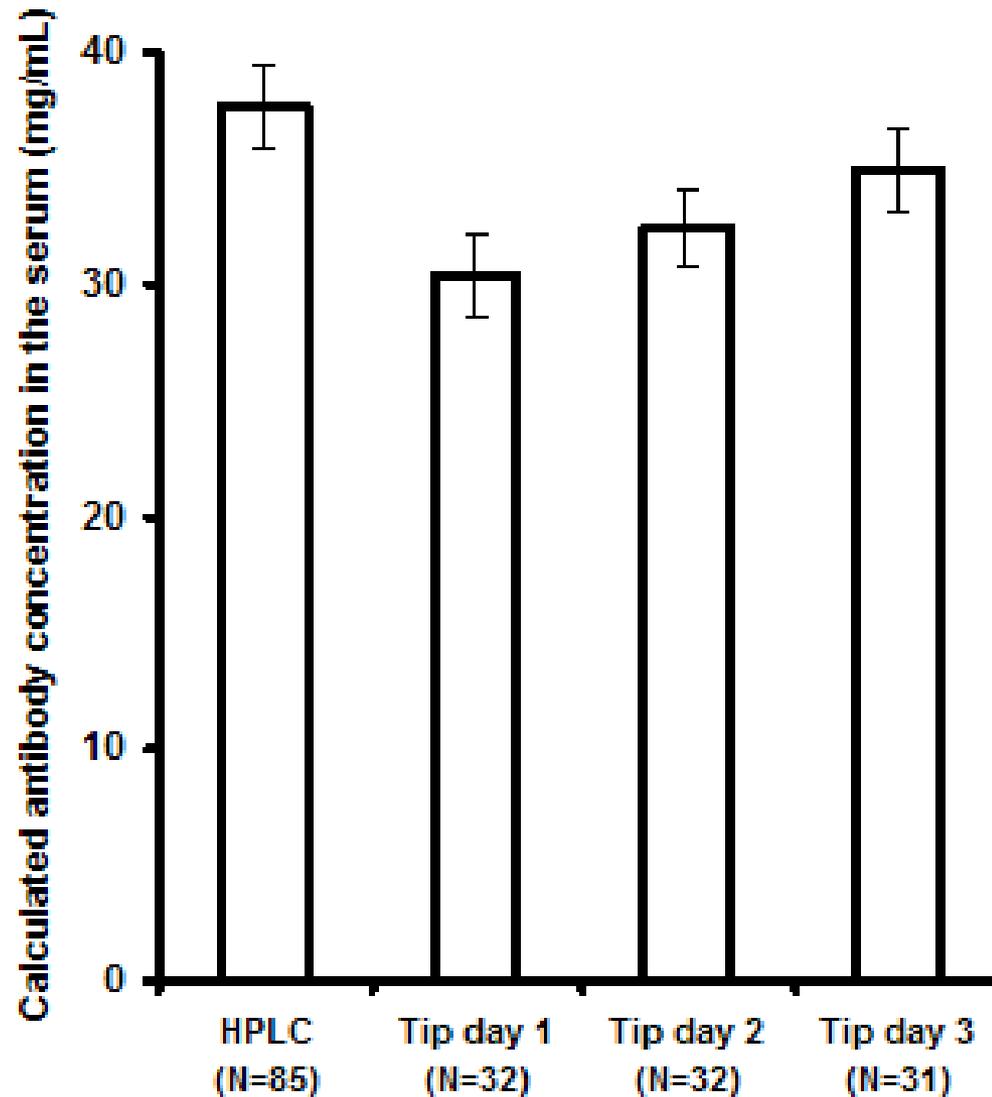
Pipette tip example 2: antibody analysis

- Robotic tip screening for ovine polyclonal antibody capture
 - Maximising dynamic binding capacities
- Protein G HPLC for antibody quantification
 - Manually-intensive
 - Time-consuming
 - Required litre scale- buffer volumes
 - Conducted on separate system to the robot
- Required a faster, more integrated alternative

Robotic tip analysis

- Protein G tip method operated on Tecan robot
 - 8-channel pipetting system
 - 40 μ L packed resin bed
- Protocol set-up to integrate with main experiment
- Evaluated properties of method with ovine pAb
 - Linearity
 - Range
 - Specificity
 - Accuracy (versus reference Protein G HPLC)
 - Precision

Sample data – accuracy and precision



Tip versus HPLC comparison

Characteristic	Protein G tips	HPLC
Samples processed simultaneously	8	1
Analysis run time / sample (minutes)	5	13
Buffer volumes	mL-scale	L-scale

Tip versus HPLC comparison

Robotic pipetting capacity [Number of liquid handling channels]	% tip – HPLC time saving
8	62
12	69
96	94

Chhatre et al. (2010), An automated packed Protein G micro-pipette tip assay for rapid quantification of polyclonal antibodies in ovine serum, *Journal of Chromatography B*, 878, 3067–3075

Continuing challenges at microscale

- **Potential for data overload**
 - Require efficient ways to store and manage data
 - Need to select test points carefully to minimise analysis
 - Prevent analytical bottlenecks
- **Need to minimise sample volume**
 - Microscale unit operation linkage must account for volume consumed in hold-up or analysis

Potential solutions

- Mathematical tools for efficient design space search
 - Simplex algorithm for early development or where analysis is time-consuming
- Smart deployment of current range of analytical methods
 - Use fast assays (e.g. total protein) at first for coarse screening
 - Leave time-consuming assays (e.g. ELISA) until a good region has been identified
- Microfluidic assays developments
 - Reduce sample volume requirements
 - Lab-on-a-chip (e.g. Agilent Bioanalyser for electrophoresis)

Conclusions

- **Microscale chromatography**
 - Rapid data generation
- **Automation**
 - Increased throughput
 - Reduced manual intervention
- **Requires rapid integrated assay techniques**
 - Avoid shifting bottleneck over to analysis