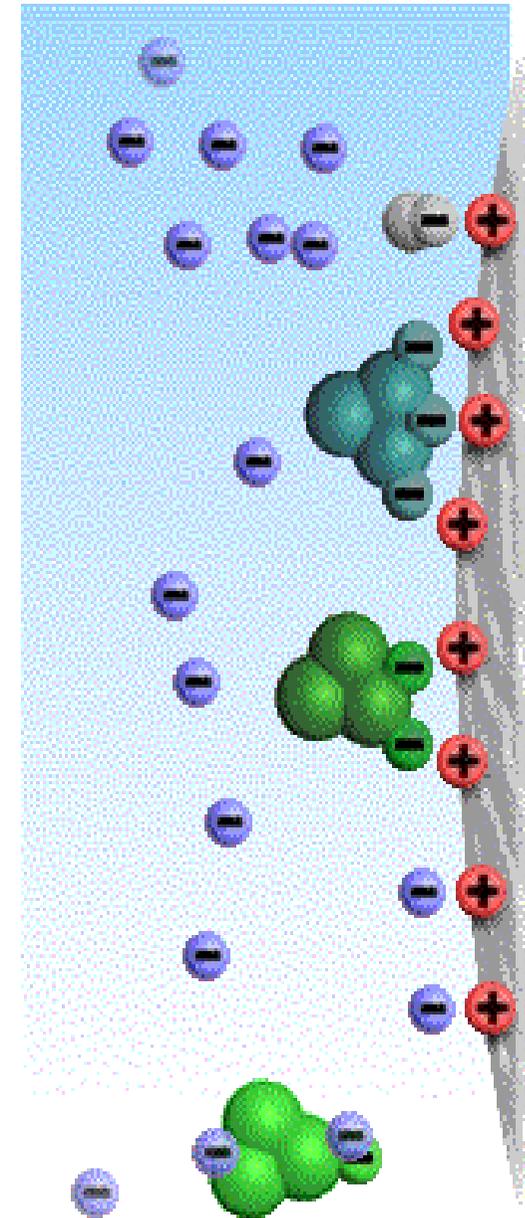


Types of Ion Exchange Chromatography Media

Andy Masters
GE Healthcare, Life Sciences
Sales Development UK and Ireland
Chromatography Resins
www.gelifsciences.com/protein-purification



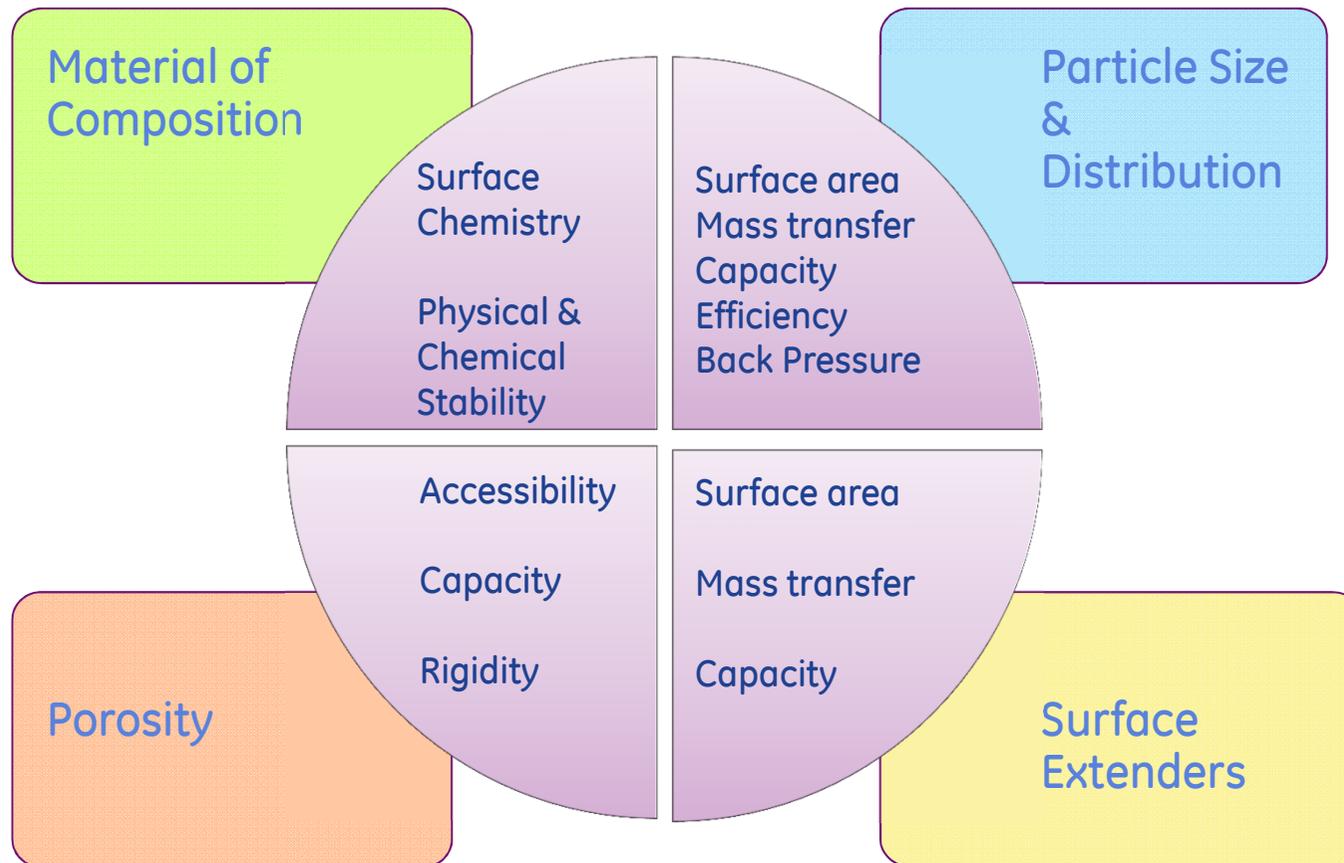
SCI IEX 2012: Technical Training
Ion Exchange Theory and Practice
for Bioprocessing
Tuesday 18 September 2012
Queens' College, University of Cambridge, UK



Content

- Properties of IEX chromatography media
- Classic ligands
- Strong and weak IEX and atypical behaviour
- Mixed mode ligands
- Future developments for IEX and MM
- Media selection

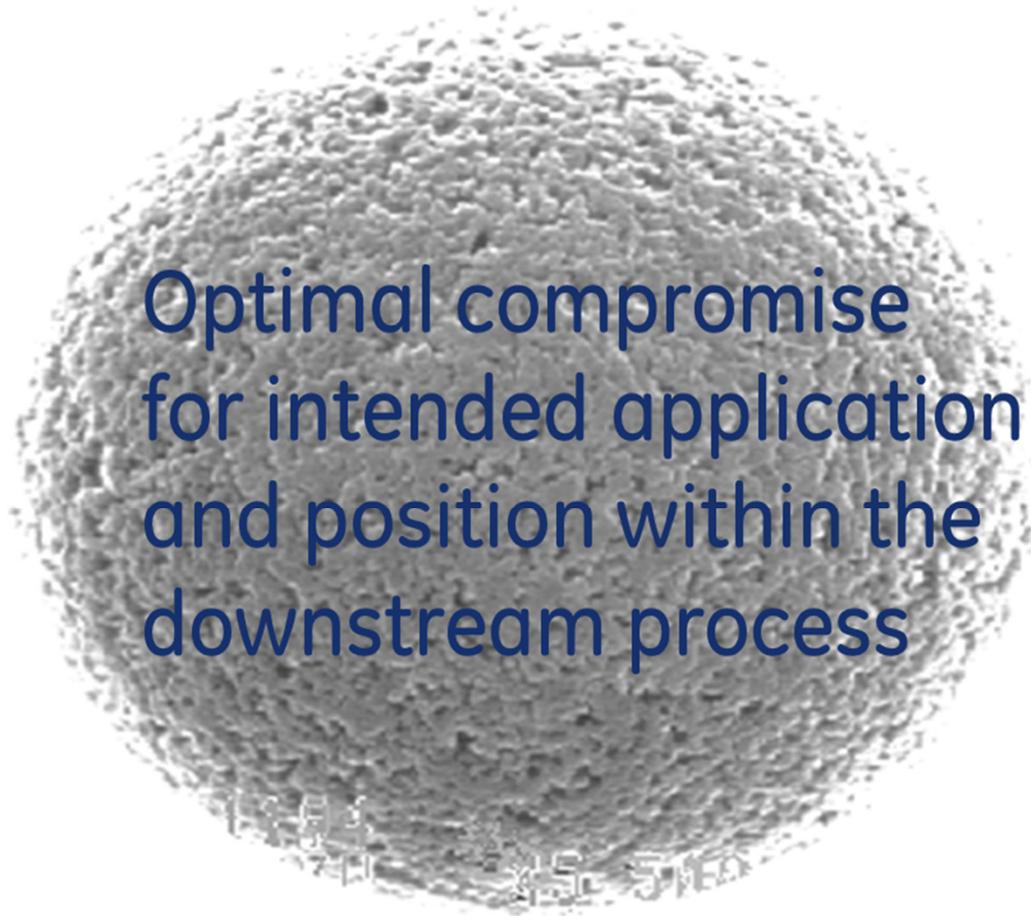
Base matrix key to properties of all chromatography media



Base | chron

s of all

M
C



Optimal compromise
for intended application
and position within the
downstream process

ze
on

P

Base matrix composition

The ideal matrix for IEX

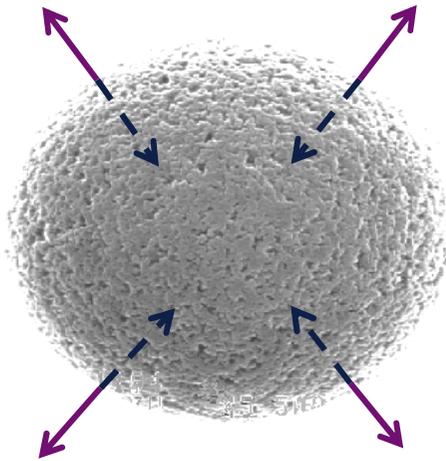
- Hydrophilic
- Large pore size/surface area
- Spherical (mono-sized) particle
- Rigid – optimal pressure/flow
- Easy to functionalize
- Chemically stable – SIP/CIP

Typical materials

- Agarose
- Cellulose
- Ceramics
- Dextran
- Polystyrene
- Polyacrylamide
- Silicas
- Synthetic/Organic Polymers

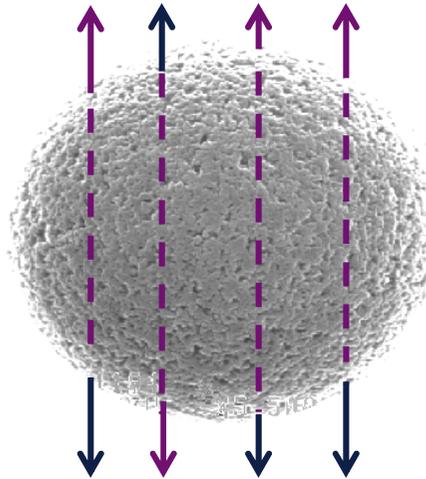
Porosity/surface area

Porous media



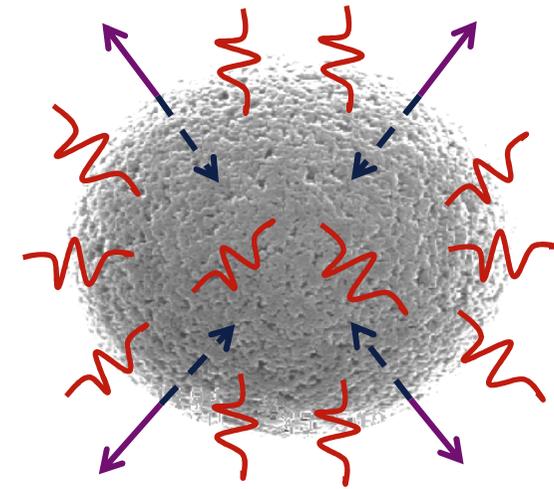
- Longer distances for mass transfer (diffusion)
- Good capacity require longer residence times
- Porosity impacts surface area and rigidity
- Pore size and porosity optimised for application

Perfusion/monoliths



- Shorter distances for mass transfer (convective flow)
- Good capacity shorter residence times
- Monoliths can be optimised for the binding of large molecules (e.g. viruses, plasmids)
- Structure can lead to limitations of scale

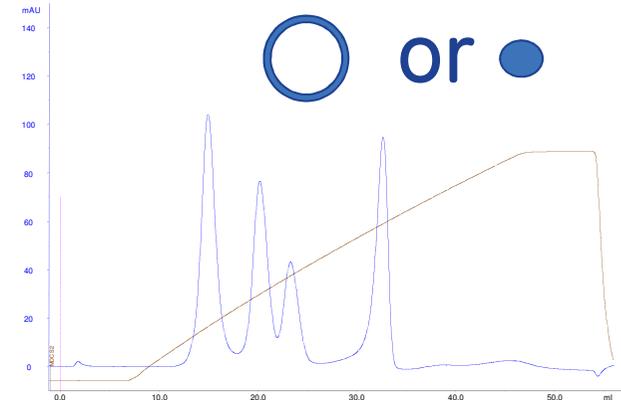
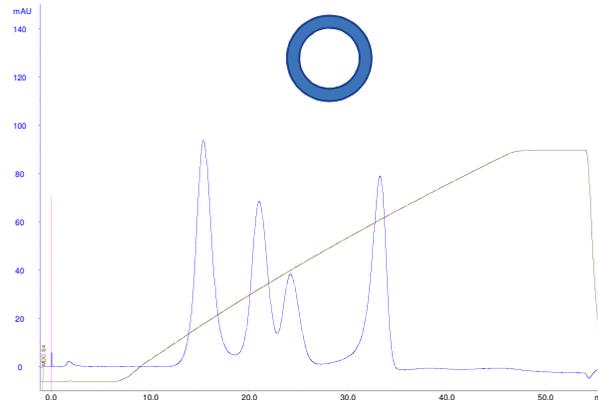
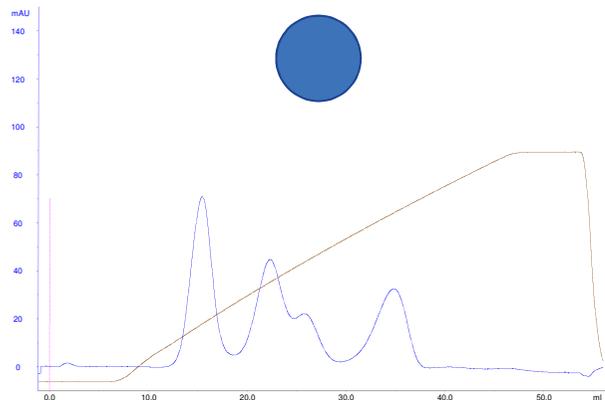
Surface Extenders



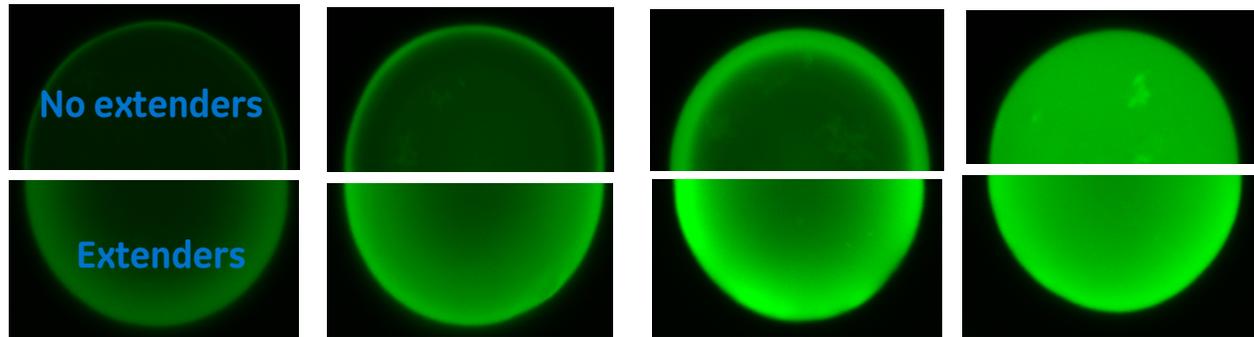
- Shorter distances for mass transfer
- Increased surface area without compromising rigidity
- High capacity even at short residence times
- Can demonstrate atypical IEX behaviour

Speed of mass transfer and resolution

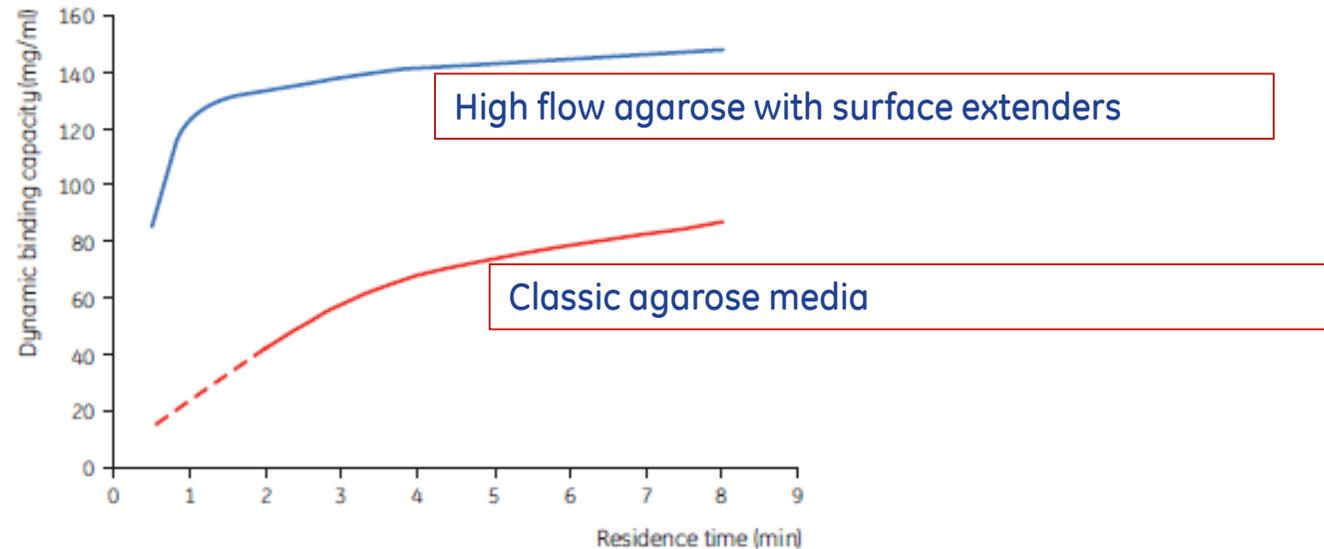
	Shell thickness (μm)	Fraction of bead volume functionalized (%)	BSA capacity (g/L)	BSA capacity (%)
	N/A	100	75	100
	11	65	61	81
	6	41	45	60



Effect of surface extenders on mass transfer and capacity



Confocal microscopy study showing enhanced protein uptake (increased mass transfer and equilibrium capacity) for IEX resin with ligand extenders



Porosity and surface extenders

Importance of critical chromatography resin properties in the design of a high productivity immunoglobulin process

Joseph Bertolini, Chor Sing Tan, Karl McCann

CSL Biotherapies, 189 Camp Road, Broadmeadows VIC 3047, Victoria, Australia

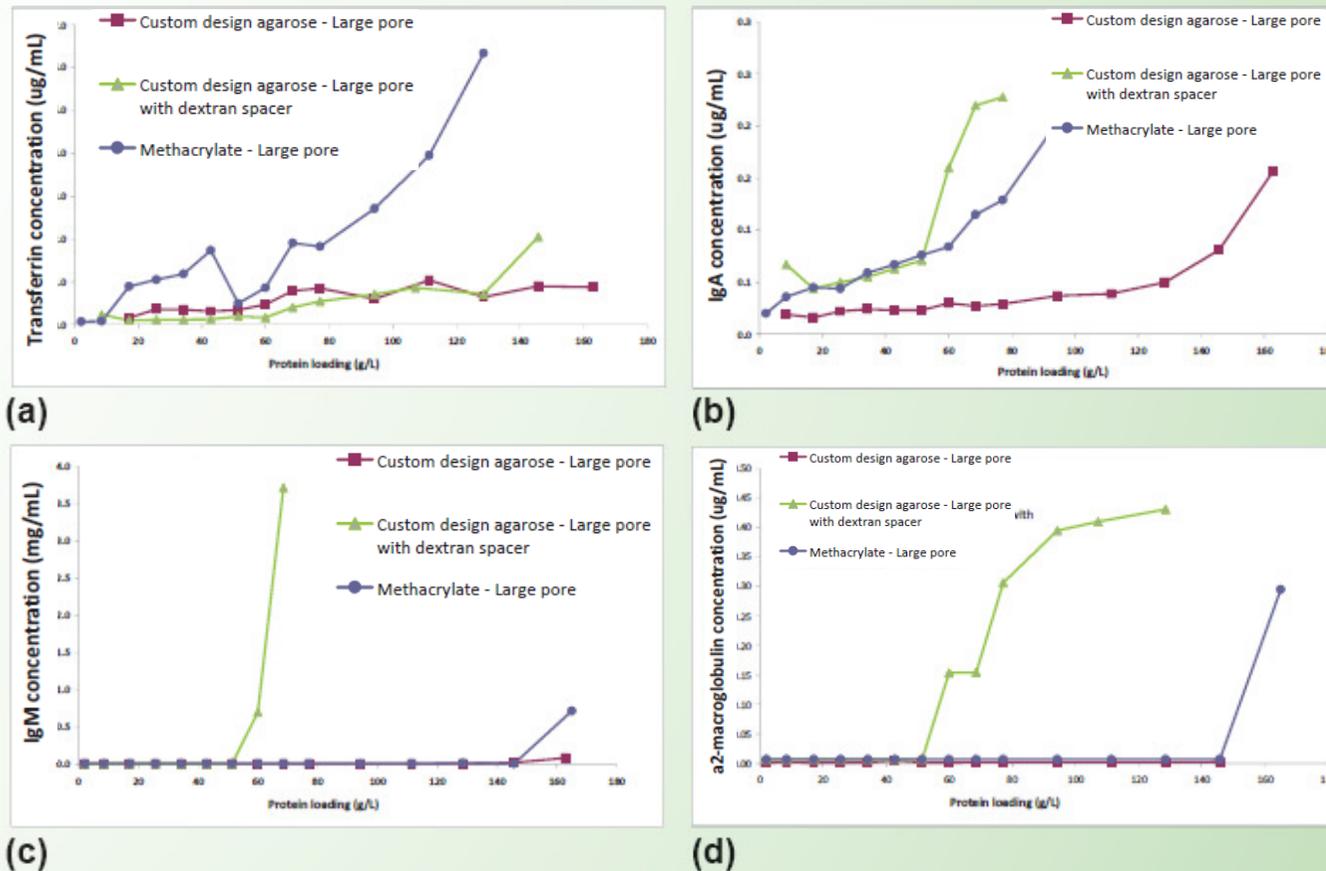


Figure 5 Impurity concentration of Pure IgG fractions derived from strong anion exchange resins loaded at 160 g/L of Crude IgG (a) transferrin (b) IgA (c) IgM (d) α_2 -macroglobulin

Bead size/distribution and resolution

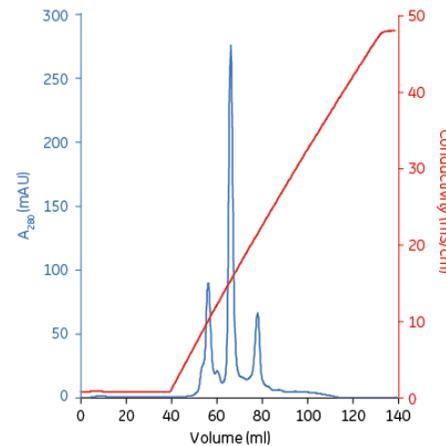
Small beads

- Fastest mass transfer
- Highest efficiency
- Best resolution
- Step and linear gradients used
- Higher back pressure
- Equipment and economy implications

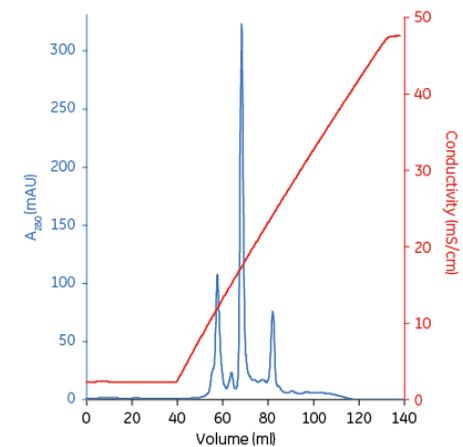
Large beads

- Good mass transfer
- Higher capacity
- Lower efficiency
- Less resolution
- High flow/capacity and step gradients for best productivity
- Low back pressure
- Most economical

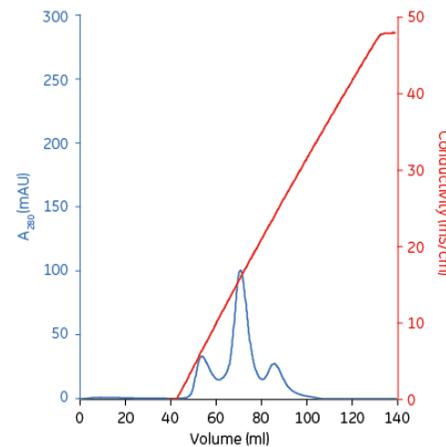
40um



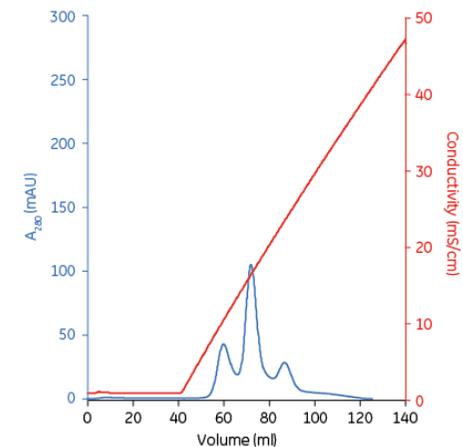
34um



90um surface extenders



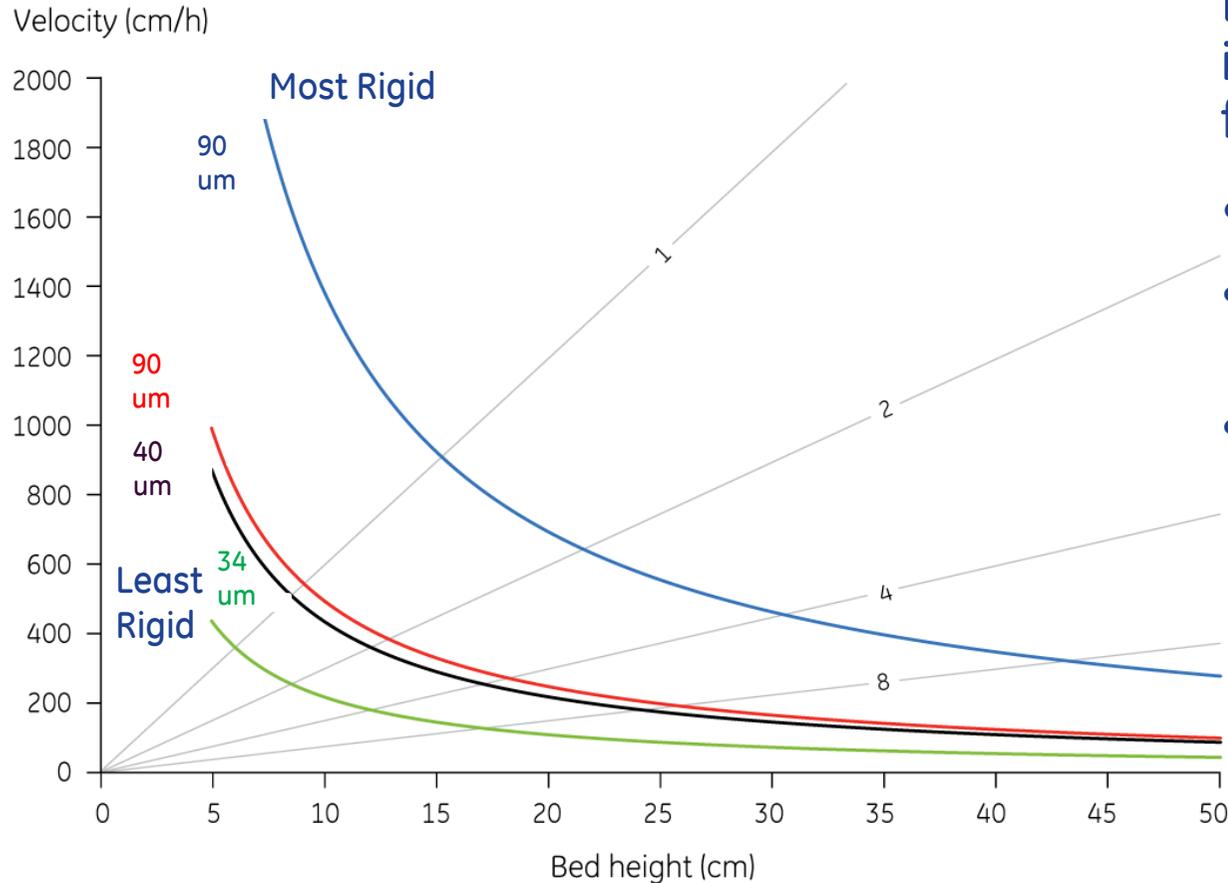
90um



imagination at work

Rigidity

Improving pressure/flow properties for modern media



Large operational window in terms of bed heights and flow rates

- Facility fit
- Higher bed heights – reduced footprint
- Higher flow rates – increased throughput



Functional groups used on ion exchangers

Cation exchangers

Carboxymethyl (CM)

Sulphopropyl (SP)

Methyl sulphonate (S)

Functional group

$-\text{OCH}_2\text{COO}^-$

$-\text{CH}_2\text{CH}_2\text{CH}_2\text{SO}_3^-$

$-\text{CH}_2\text{SO}_3^-$

Anion exchangers

Diethylaminopropyl (ANX)

Diethylaminoethyl (DEAE)

Quaternary aminoethyl (QAE)

Quaternary ammonium (Q)

Functional group

$-\text{CH}_2\text{CHOHCH}_2\text{N}^+\text{H}(\text{CH}_2\text{CH}_3)_2$

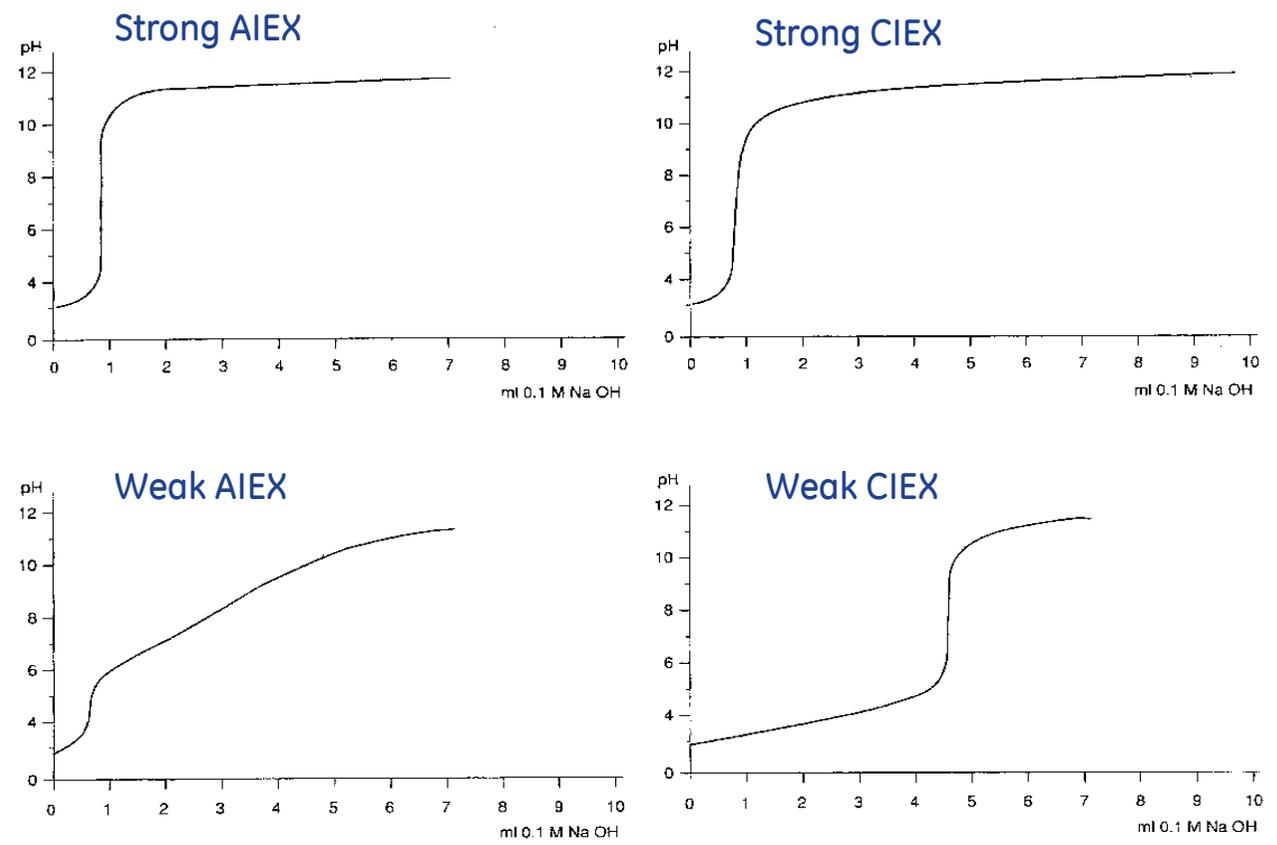
$-\text{OCH}_2\text{CH}_2\text{N}^+\text{H}(\text{CH}_2\text{CH}_3)_2$

$-\text{OCH}_2\text{CH}_2\text{N}^+(\text{C}_2\text{H}_5)_2\text{CH}_2\text{CH}(\text{OH})\text{CH}_3$

$-\text{CH}_2\text{N}^+(\text{CH}_3)_3$

Strong and weak ion exchangers

strong ion exchangers: capacity is constant over a wide range of pH

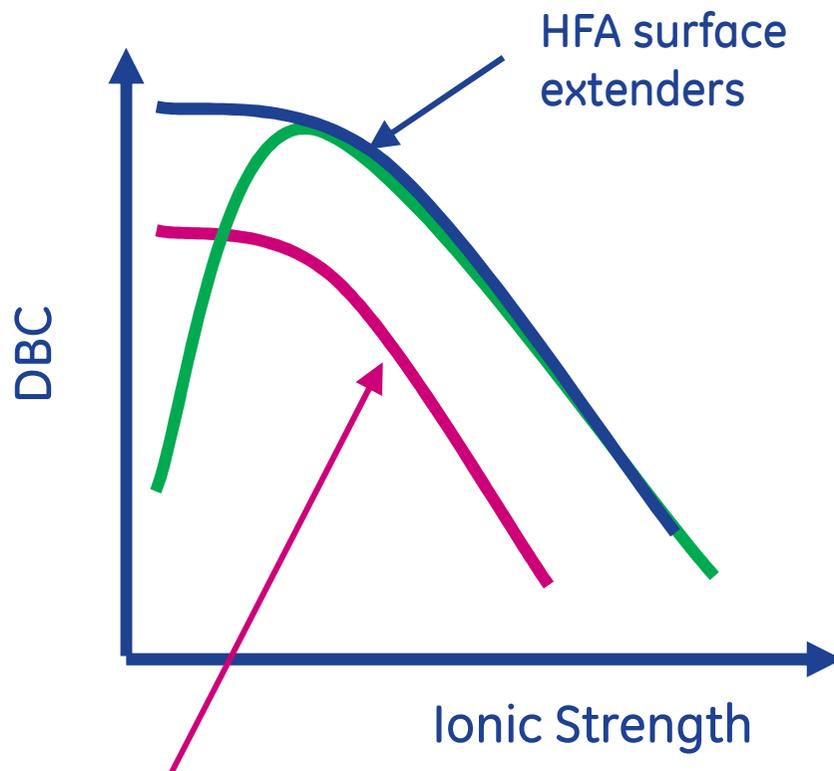


weak ion exchangers: capacity varies with pH



Surface extenders and non-traditional AIEX behavior!

Effect of conductivity on DBC



Classic agarose



- Normally the highest DBC is obtained at low ionic strength (conductivity)
- Sometimes there is an optimal ionic strength for absorption of a certain sample under certain loading conditions (pH) so called non-traditional behavior
- Non-traditional behavior seems to occur more often on surface enhanced ion exchangers.

Multimodal chromatography

Multimodal benefits:

- Unique selectivity
- Use with complex feeds
- Purify challenging targets
- De-colourise feed stocks

Different techniques on one matrix by:

- Chemically different ligands
- One ligand with several characteristics

Utilizes many types of interactions:

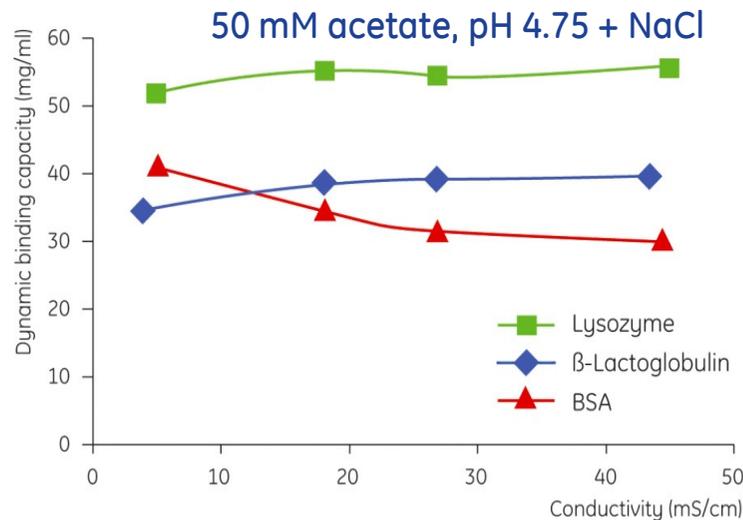
Ionic
Hydrophobic
Hydrogen bonding
Thiophilic
Pseudo-affinity

Examples:

N-benzyl-n-methyl ethanolamine
N-Benzoyl-DL-homocysteine
Octylamine
4-Mercapto-Ethyl-Pyridine
Hexylamine
Phenylpropylamine
Hydroxyapatite

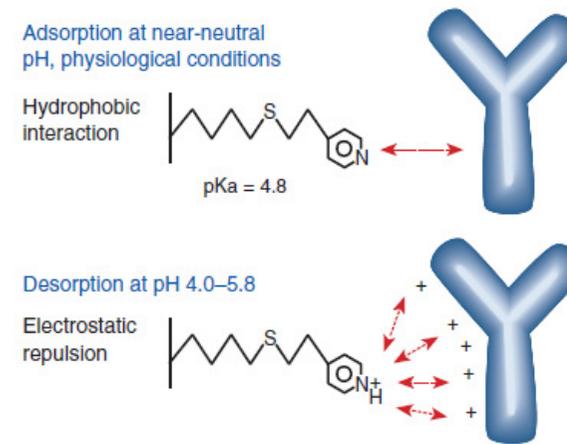
MM designed for specific applications

Capture in high salt (N-Benzoyl-DL-homocysteine)



- Capture from undiluted feedstocks.
- Remove requirement for dilution or diafiltration
- Increase productivity and process economy

Purification of antibodies (4-Mercapto-Ethyl-Pyridine)*



- A no-salt/low-salt alternative to Hydrophobic Interaction Chromatography (HIC)
- Monoclonal and polyclonal IgG capture and intermediate purification (aggregate, DNA and HCP removal)
- Enhanced process economics

But gaining widespread applicability and used as a development platform

Table 1. The use of mixed-mode materials in the downstream processes of various therapeutic proteins

Protein	Source	Resin	Process step	Binding	Elution	Load (mg/mL)	Purity after process step (%)
Cytokine mutein	<i>E. coli</i>	CHT type I	Intermediate	5 mM K-phosphate, pH 6.5	5–500 mM K-phosphate gradient, pH 6.5	5–18	70–80
Recombinant protease inhibitor	Yeast supernatant	Lewatit CNP105	Capture	Supernatant adjusted to pH 6	0.1 N HCl	7	35–40
Recombinant protease inhibitor mutein I	Yeast supernatant	Capto MMC	Capture	20 mM Na acetate, pH 5	20 mM Tris, 2 M NaCl, pH 8.5	9–17	90
Recombinant protease inhibitor mutein I	Yeast supernatant	HEA Hypercel	Intermediate	20 mM Tris, 2 M NaCl, pH 8.5	40 mM Na citrate, pH 2.7	15–27	>90
Recombinant protease inhibitor mutein II	Yeast supernatant	Capto MMC	Capture	20 mM Na citrate, pH 3	20 mM Tris, pH 8.5	10	85–90
Human protease inhibitor	<i>E. coli</i> lysate	Capto MMC	Capture	50 mM Na citrate, pH 6.5	50 mM Tris, 0.5 M NaCl, pH 7.5	5	85
Antibody fragment	Yeast supernatant	MEP Hypercel	Capture	25 mM Tris, pH 7.5	25 mM Na citrate, 150 mM NaCl, pH 3.7	15–26	72
Antibody fragment	Mammalian cell line	CHT type I	Polishing	15 mM K-phosphate, pH 7	15 mM K-phosphate, pH 7, NaCl gradient	10	>98

Mixed-Mode Chromatography in Downstream Process Development

Salt-tolerant adsorption and unique selectivity are the major advantages of mixed-mode materials over single-mode resins. Mar 2, 2010

By: [Felix Oehme, PhD](#), [Joerg Peters, PhD](#) data from Bayer Pharma, Biotech Development

BioPharm International Supplements



imagination at work

Complete process using only MM media for challenging target proteins

Figure 2. Flow scheme of the purification process for the recombinant protease inhibitor mutein I

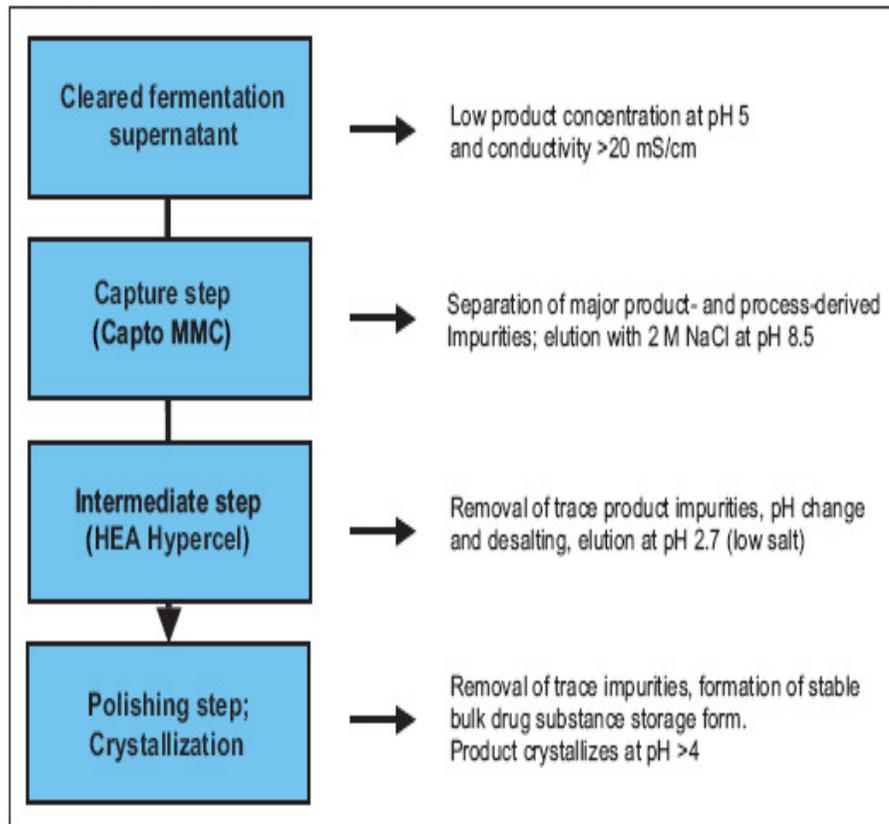


Table 3. Purification data from a typical Capto MMC capture run of the recombinant protease inhibitor mutein I

Step	Volume (L)	Mutein concentration (mg/L)	Total amount mutein (g)	Yield (%)	Purity (%)
Starting material	30	267	8.0	-	<10
Flow through	30	0	0	0	-
Wash 1 + 2	10	0	0	0	-
Wash 3	7.5	933	0.7	10	36
Elution	2.8	2,600	7.3	90	90

Table 5. Purification data from a typical HEA Hypercel intermediate step of the recombinant protease inhibitor mutein I

Step	Volume (L)	Mutein concentration (mg/L)	Total amount mutein (g)	Yield (%)	Purity (%)
Starting material	1.15	2,600	3.0	-	90
Flow through	1.15	0	0	0	-
Wash	0.8	0	0	0	-
Elution	0.35	7,372	2.6	87	91

Mixed-Mode Chromatography in Downstream Process Development

Salt-tolerant adsorption and unique selectivity are the major advantages of mixed-mode materials over single-mode resins. Mar 2, 2010

By: [Felix Oehme, PhD](#), [Joerg Peters, PhD](#) data from Bayer Pharma, Biotech Development

BioPharm International Supplements

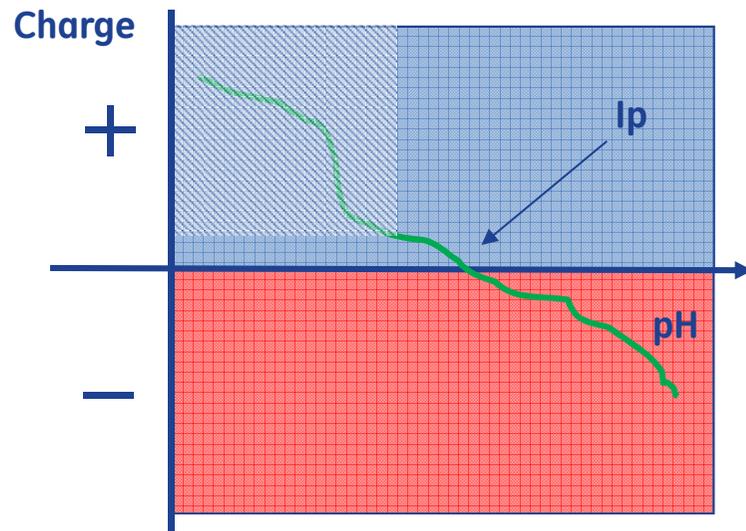


imagination at work

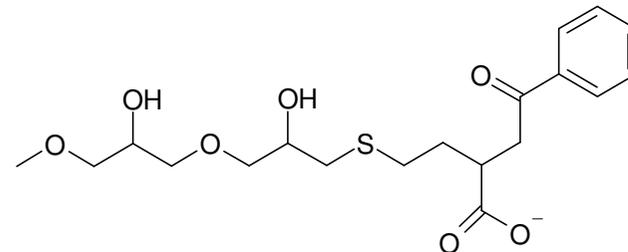
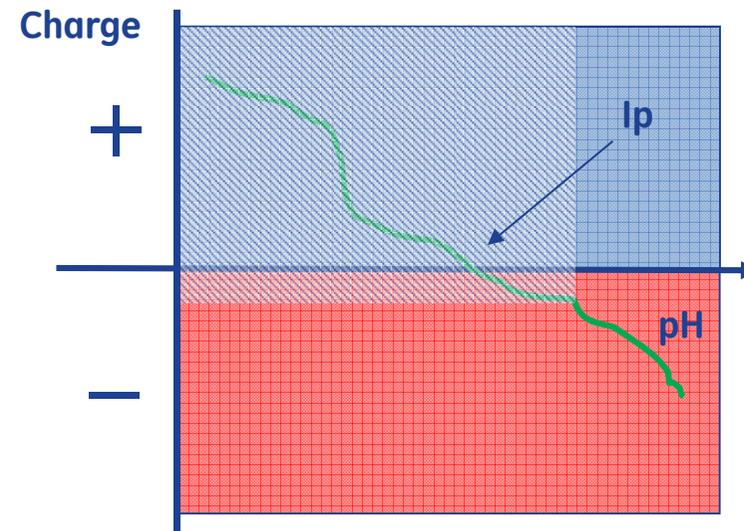
Cation exchangers vs. Multimodal

Isoelectric point vs. loading pH

Traditional cation exchange



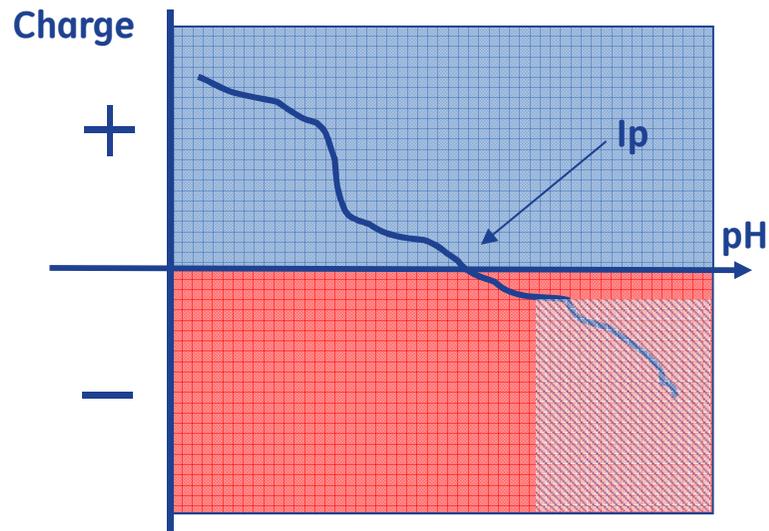
N-Benzoyl-DL-homocysteine



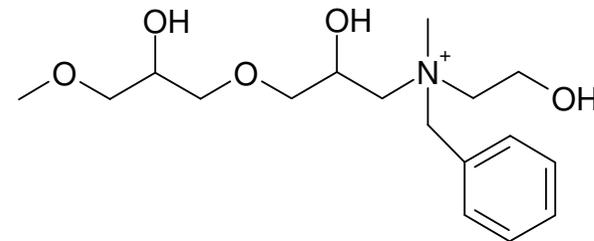
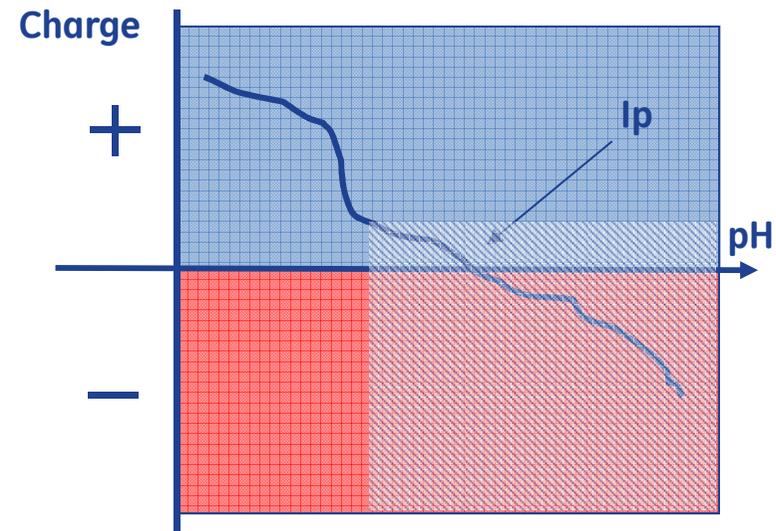
Anion exchangers vs. Multimodal

Isoelectric point vs. loading pH

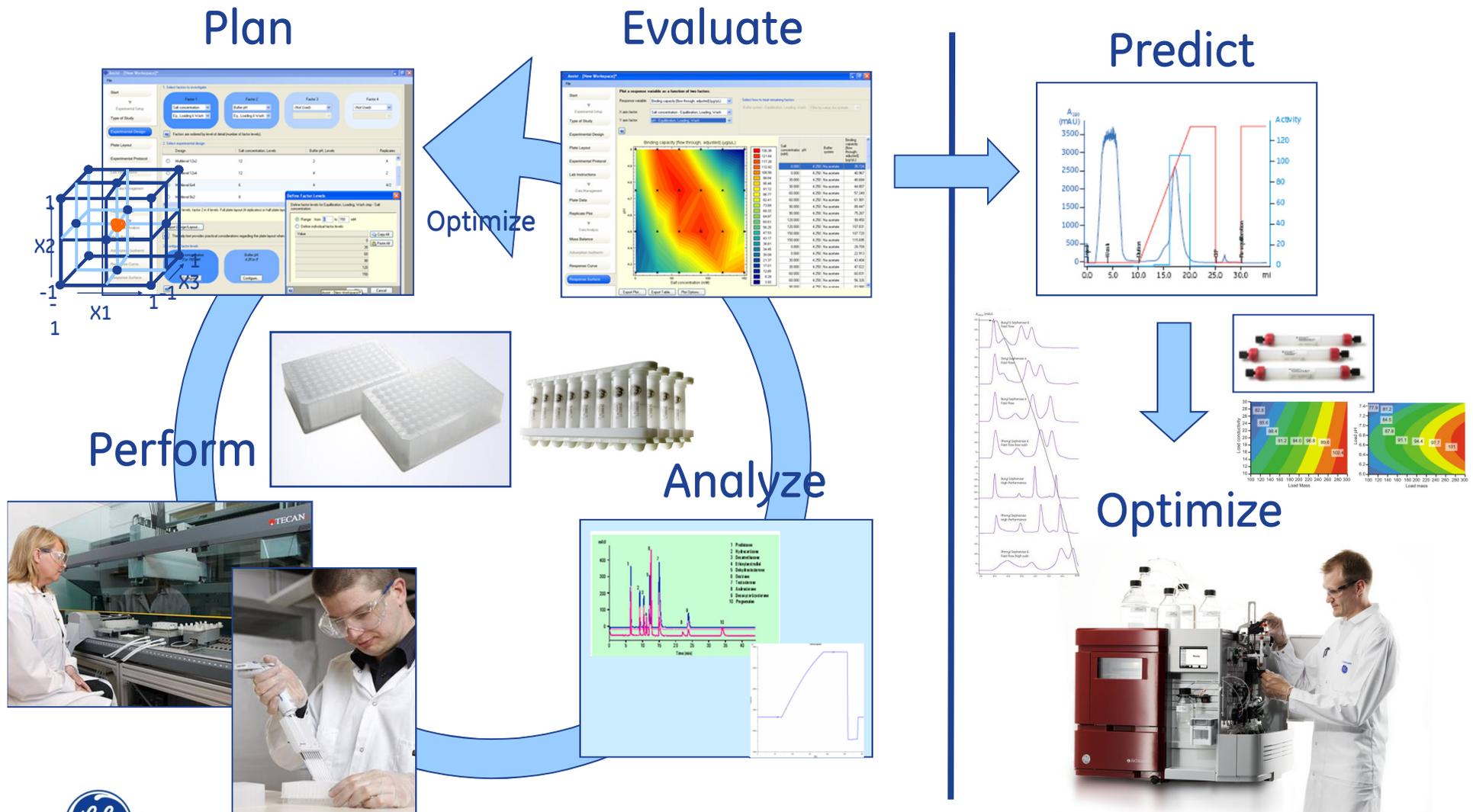
Traditional anion exchange



N-benzyl-n-methyl ethanolamine

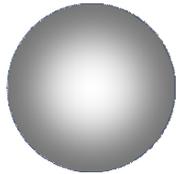


HTPD & DoE ideal for optimisation

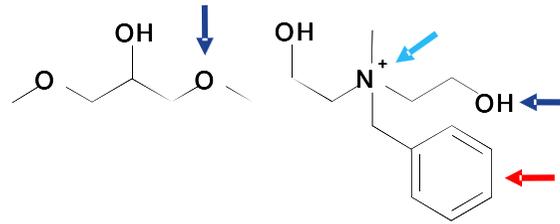


DoE ensures a more robust process

Multimodal media



+



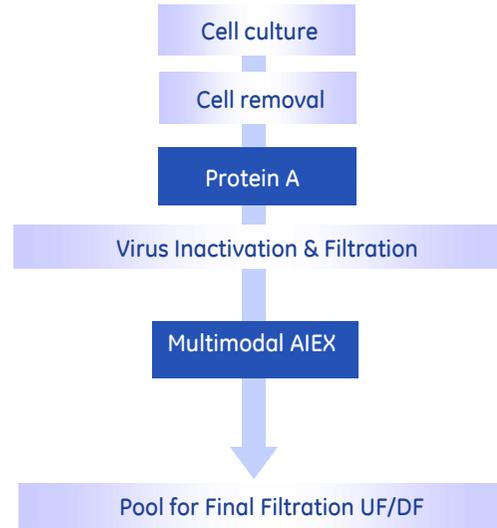
Base matrix

High flow agarose
75 µm (average)

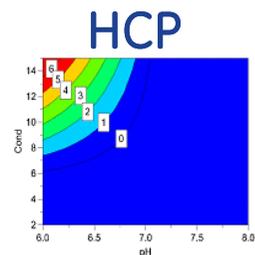
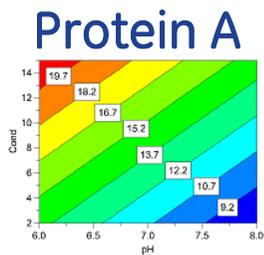
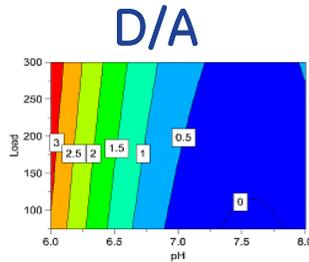
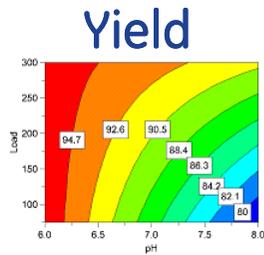
N-benzyl-n-methyl ethanolamine

Multimodal strong anion ligand

Two-chromatographic step process



Viral clearance

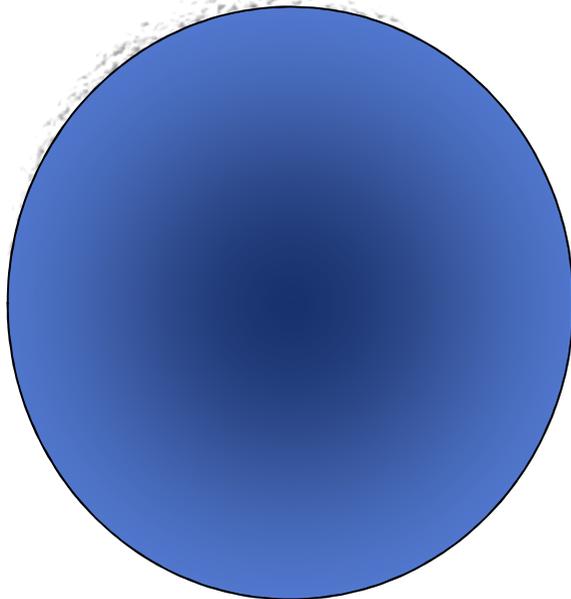


Virus	Conductivity	Log ₁₀ red. factor
MVM	10	5.8
MVM	30	5.9
MuLV	10	4.5
MuLV	30	3.6



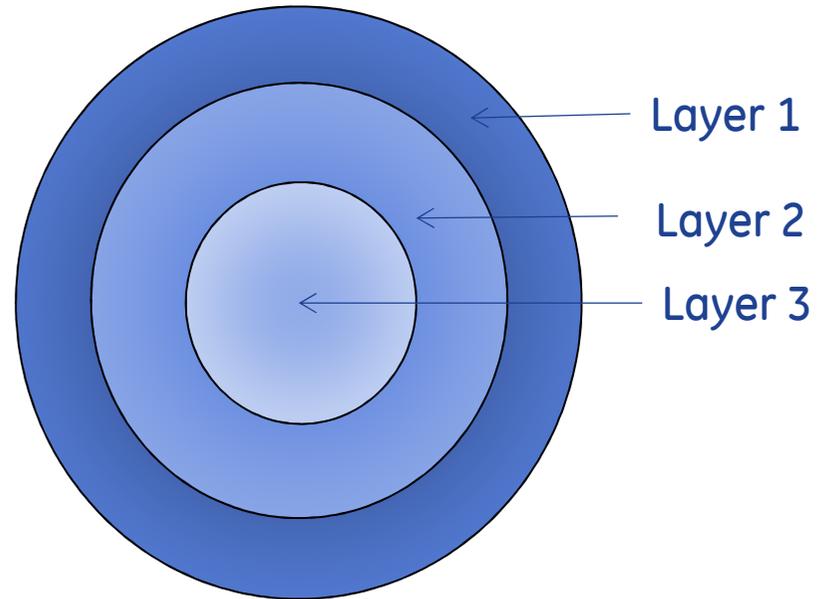
What's next for AIEX?

Concept
Layered beads
More dimensions to each bead



Design variables

- Particle size
- Pore size
- Type of ligand
- Ligand density



Additional design variables

- Number of layers
- Layer thickness
- Porosity modification
- Type of ligand

CORE BEADS combining IEX and SEC

5 μm inactive shell

Viral
particle



Octylamine - Multimodal ligand
Very strong protein binding

Host cell proteins (HCP)
DNA fragments
Endotoxins
Detergents
Benzonase™
Etc...

85 μm particle size

Selecting an appropriate IEX resin

- **Where is the step in the downstream process?**
 - Capture, Intermediate purification, polishing
 - Influence bead size, flow rates, capacity, salt tolerance
- **What's the goal of the step?**
 - Bulk separation, concentration of target, removal of contaminants (binding or flow through mode), resolution.
 - Influence bead size, flow rates, capacity (target or contaminants) ligand selectivity
- **What are the characteristics of the target molecule vs. contaminants?**
 - pI and charge, stability, size
 - Influence choice of ligand, porosity, capacity, bead size
- **What are the characteristics of the feed?**
 - Clarity, viscosity, composition
 - Influence choice of ligand, porosity, capacity, bead size and distribution, rigidity, flow rates
- **Compatibility with other techniques in the process (AF, HIC, SEC etc.)?**
 - Minimise sample handling, diafiltration, and or dilution,
 - Influence number of U/O, process economy, robustness
- **Scalability of process conditions within facility?**
 - Flow rates, capacity, pressure, packing, buffers
 - Influence column dimensions, system and column requirements, process tanks



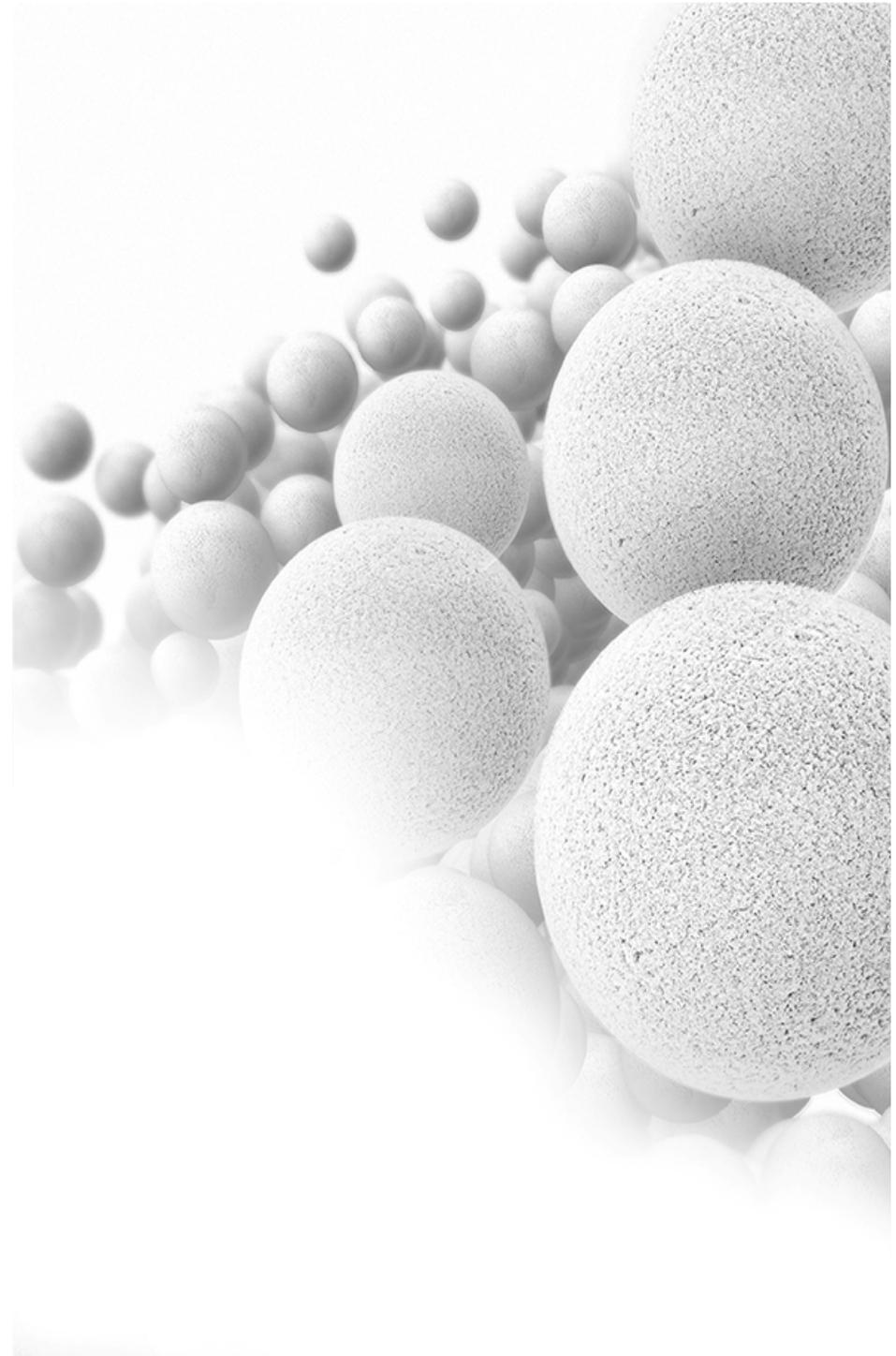
imagination at work

Handbooks from GE Life Sciences



imagination at work

Thank you!



GE, imagination at work and GE Monogram are trademarks of General Electric Company.

ÄKTA, ÄKTA pilot, ÄKTA process, AxiChrom, BioProcess, Capto, Chromaflow, HiScreen, HiTrap, MabSelect, MabSelect SuRe, MabSelect Xtra, MacroCap, Media Wand, PreDicator, RoboColumn, ReadyToProcess, Sephacryl, Sephadex, Sepharose, SOURCE, Superdex and UNICORN are trademarks of General Electric Companies.

All third party trademarks are the property of their respective owners.

All goods and services are sold subject to the terms and conditions of sale of the company within GE Healthcare which supplies them. A copy of these terms and conditions is available on request. Contact your local GE Healthcare representative for the most current information.

© 2011 General Electric Company – All rights reserved.

GE Healthcare Bio-Sciences AB, a General Electric Company.

GE Healthcare Bio-Sciences AB, Björkgatan 30, SE-751 84 Uppsala, Sweden.