

# **Mediator Lipidomics: dissecting the role of PUFA-derived metabolites**

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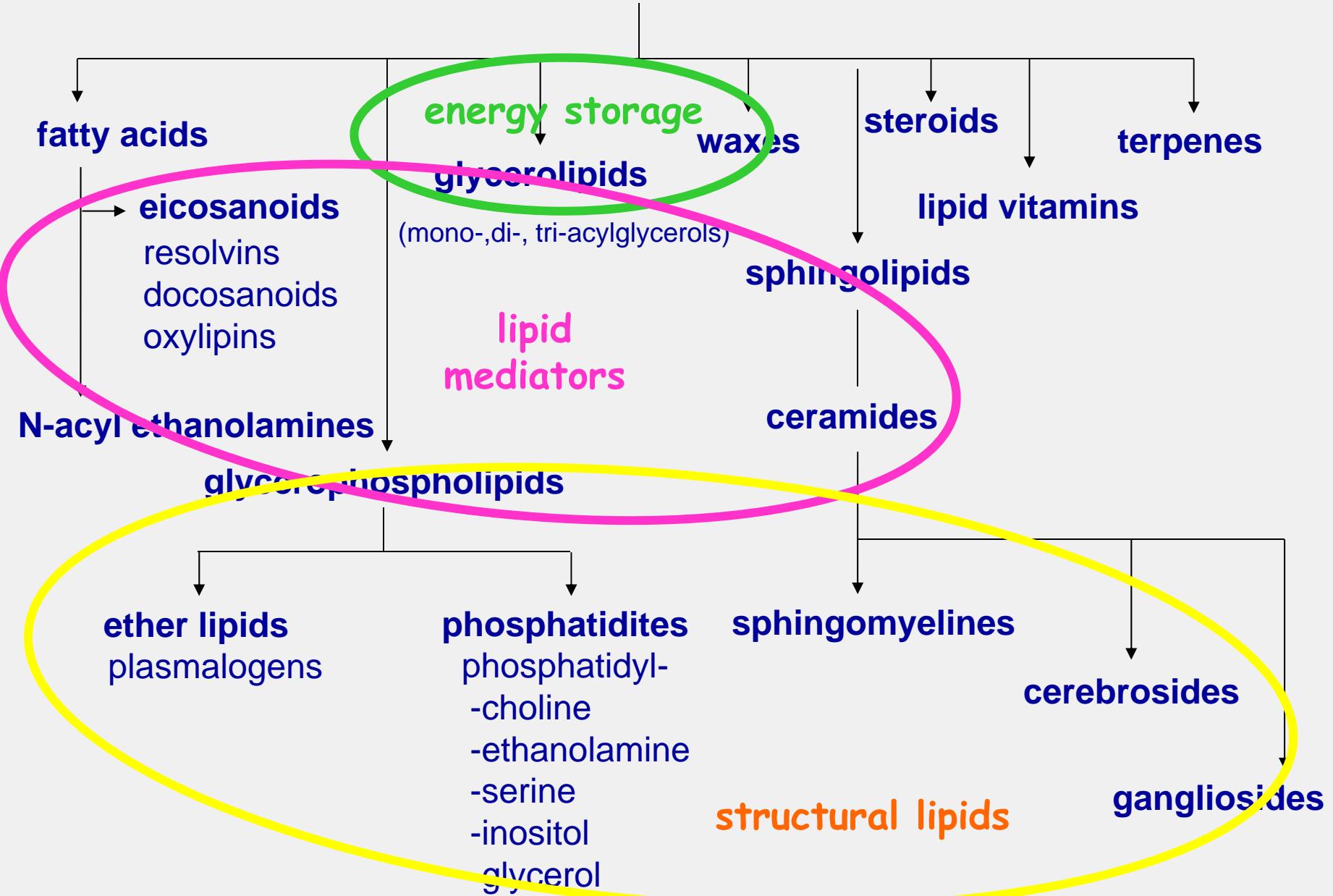
**Anna Nicolaou, PhD FRSC**  
**Professor of Biological Chemistry**

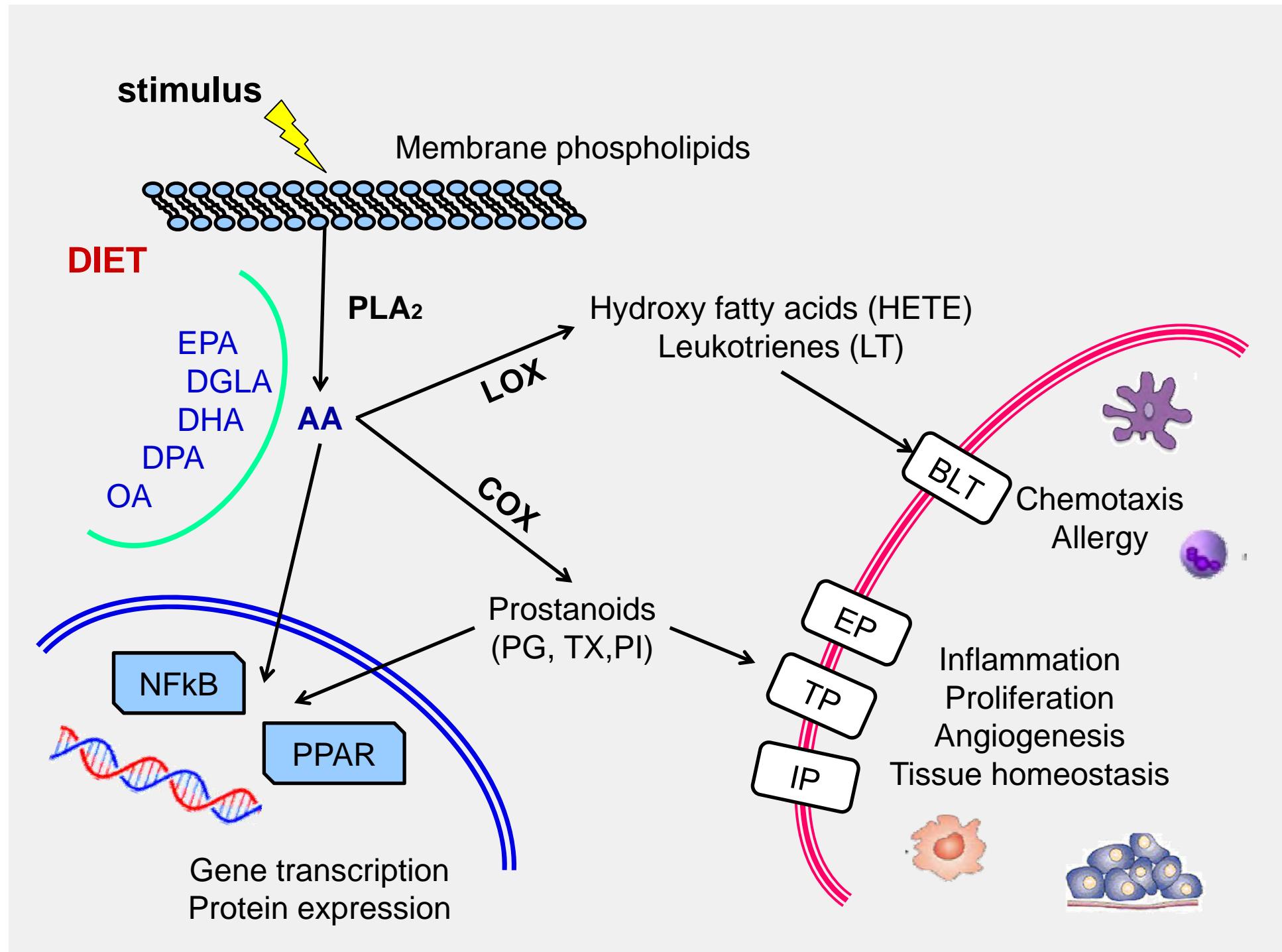


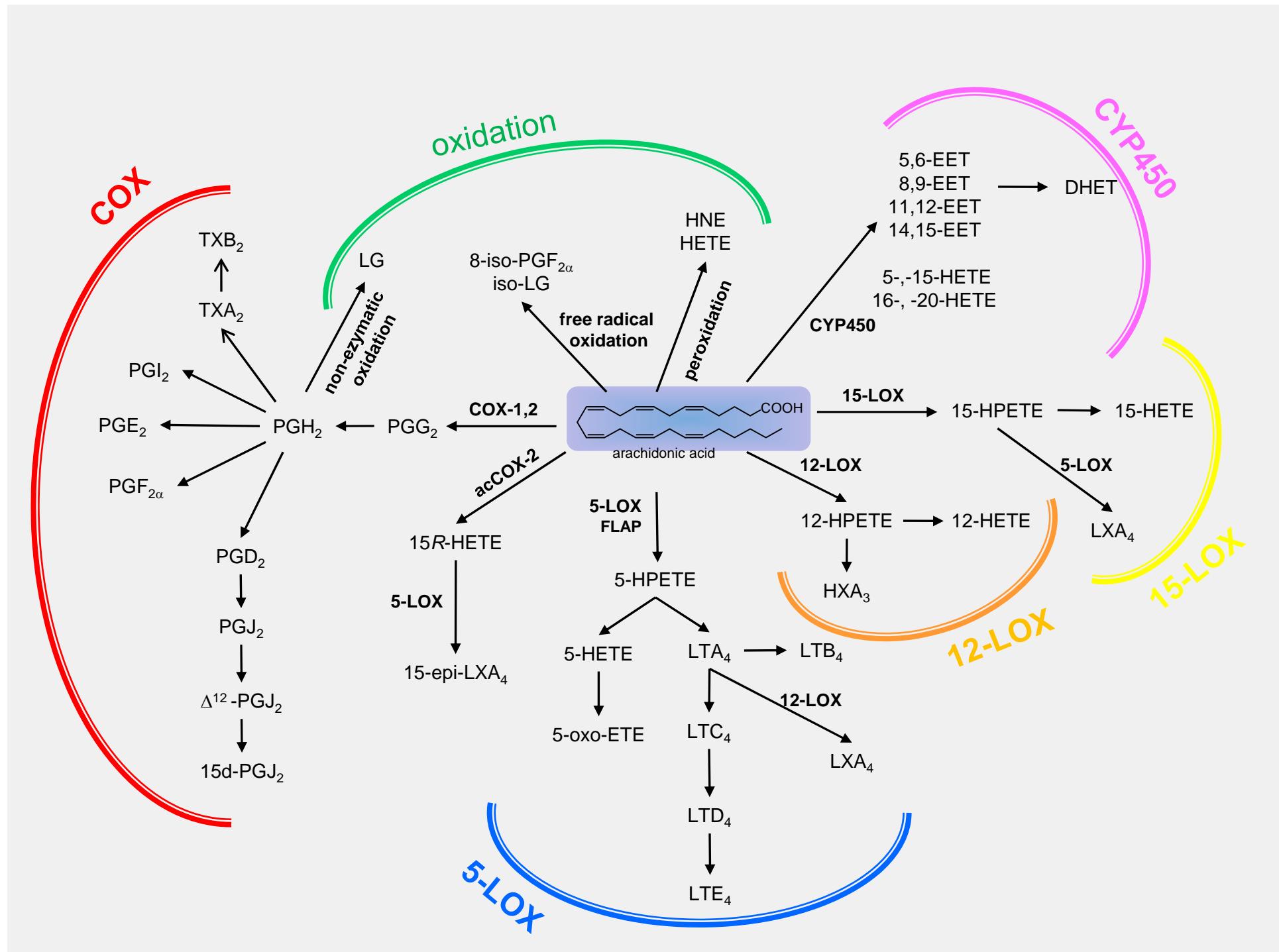
**Compositional Analysis of Lipids**  
**21 June 2013, Ghent**

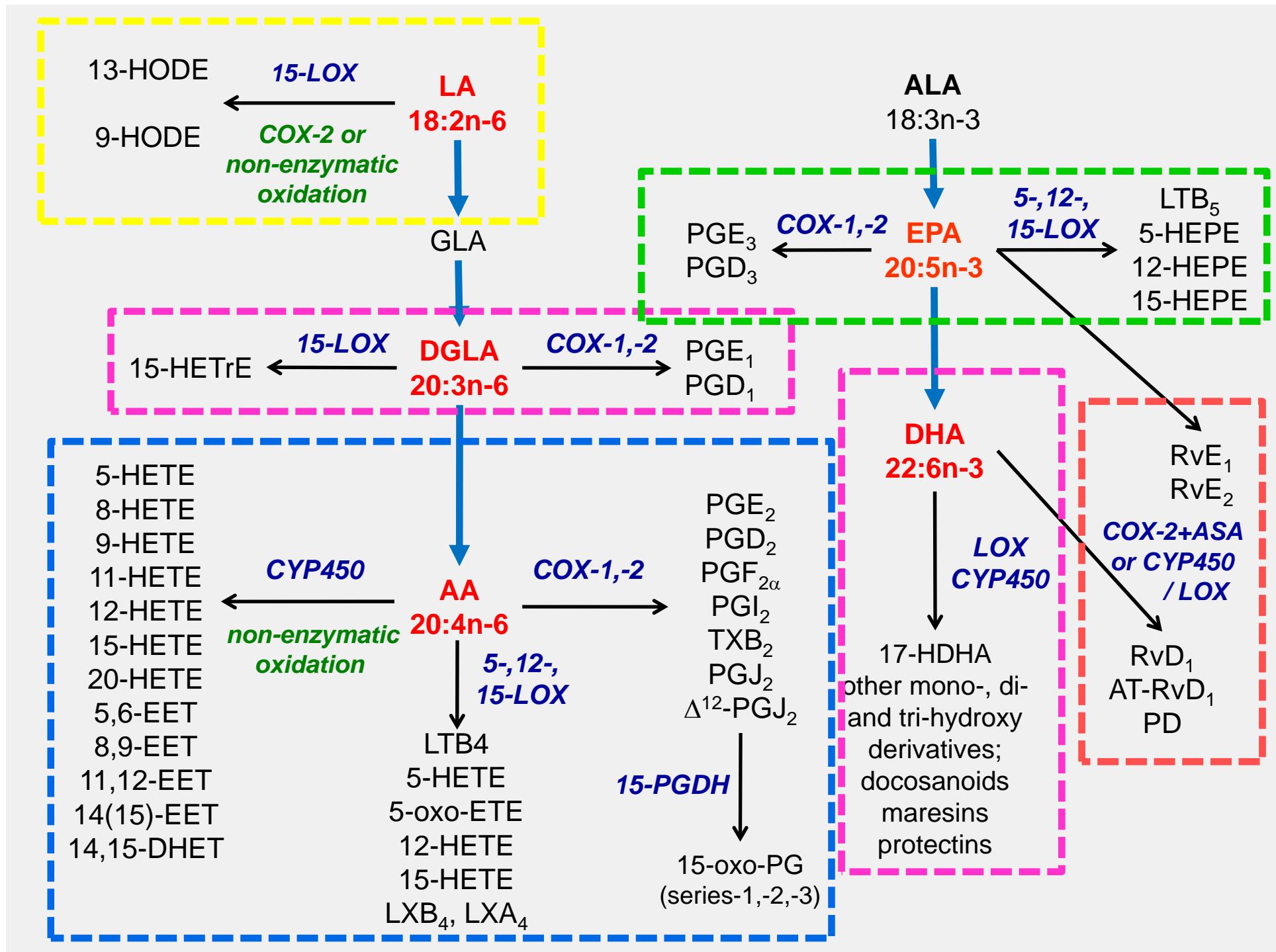


# BIOACTIVE LIPIDS

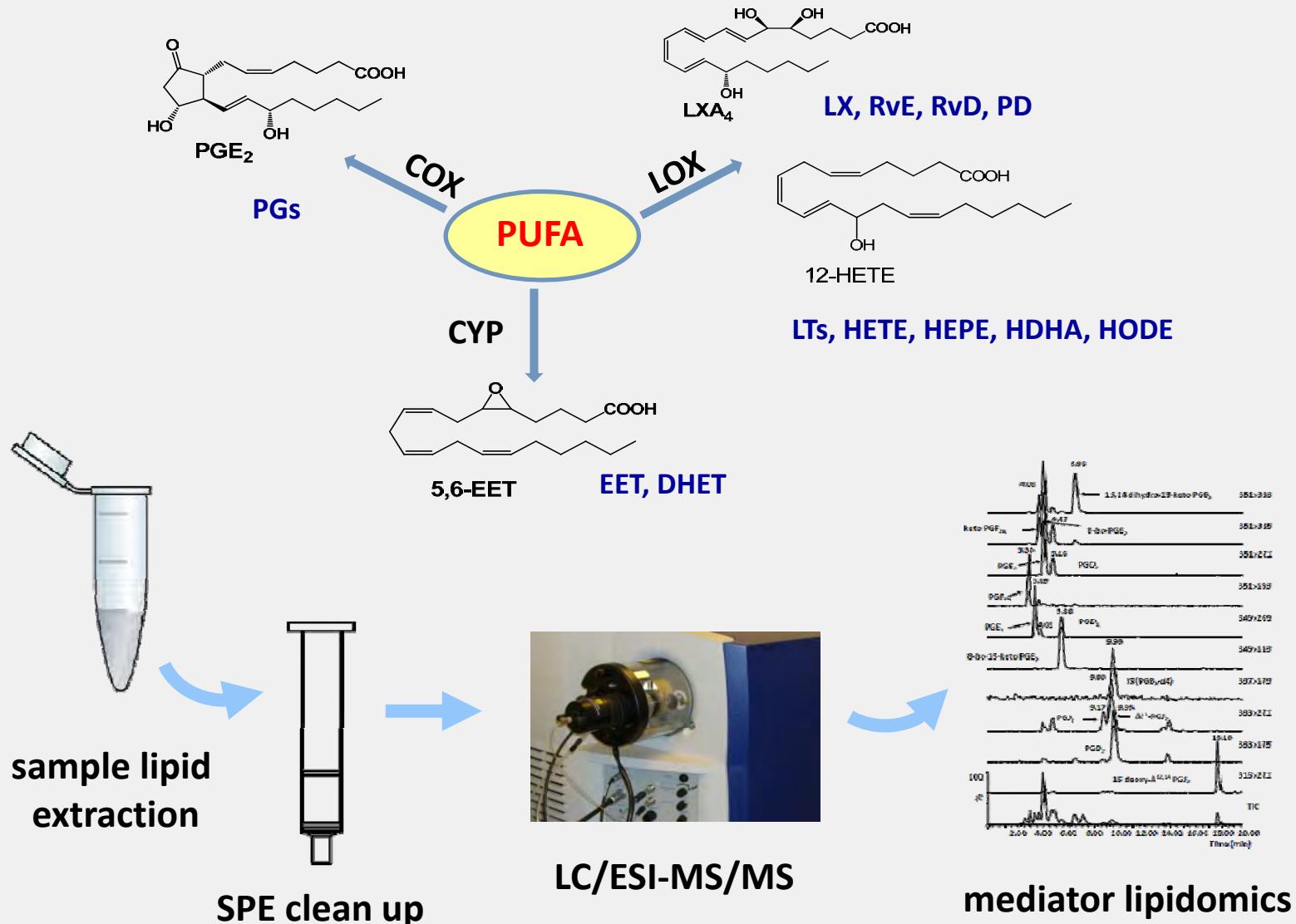








# Mediator Lipidomics: array of >80 lipids



# Mediator lipidomics workflow

## Sample preparation



**Solid:** Homogenise in 15 % (v/v) methanol in water  
**Liquid:** Adjust to 15 % (v/v) methanol  
Add internal standard(s)

## Solid phase extraction



Remove precipitated proteins  
Acidify sample (pH 3.0)  
Load on C18 SPE cartridge  
Elute lipids with methyl formate

## Lipid extract preparation & storage



Remove solvents under N<sub>2</sub>  
Reconstitute extract in small solvent volume  
If needed, store at -20 °C and for up to 1 week

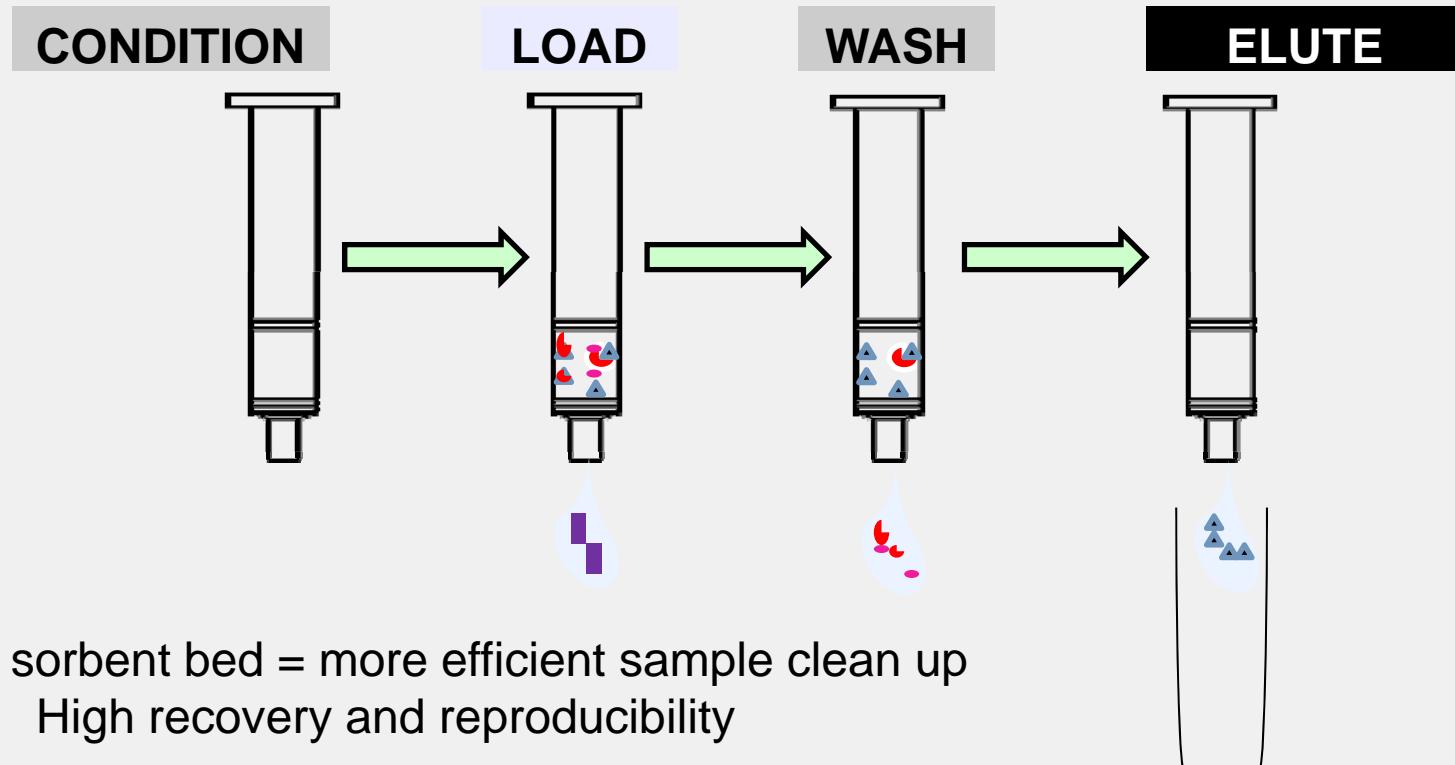
## LC/ESI-MS/MS



Prepare fresh calibration standards  
Check baseline and instrument sensitivity  
Run COX assay or  
Run LOX/CYP assay or  
Run chiral assay

## Data processing and calculations

# Solid phase extraction (SPE) clean up



1. **Condition SPE:** Prepare C18 sorbent bed

2. **Load extract:**

3. **Wash:** Remove weakly bound impurities, then wash with hexane to remove water

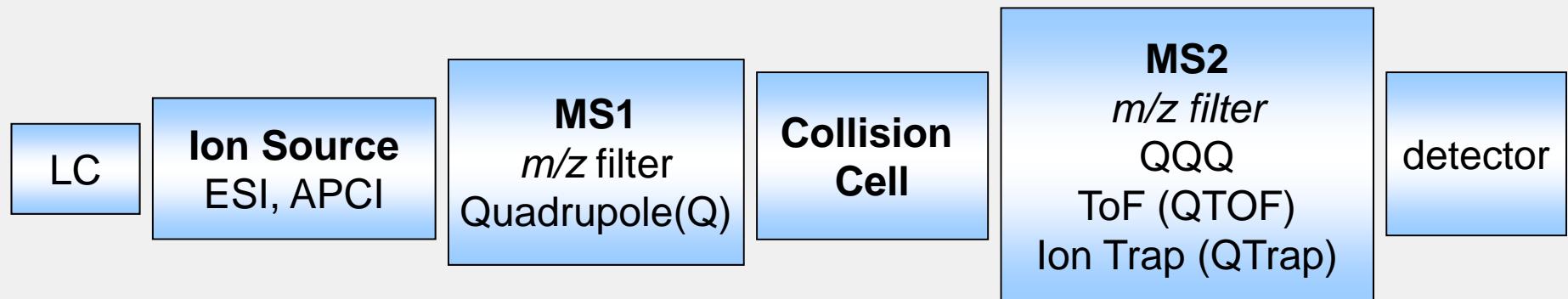
Reduce matrix effects

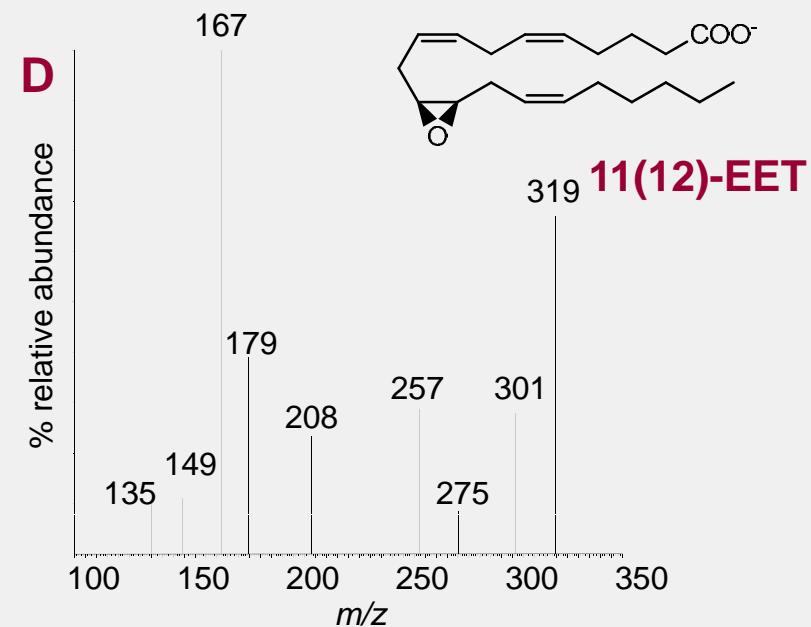
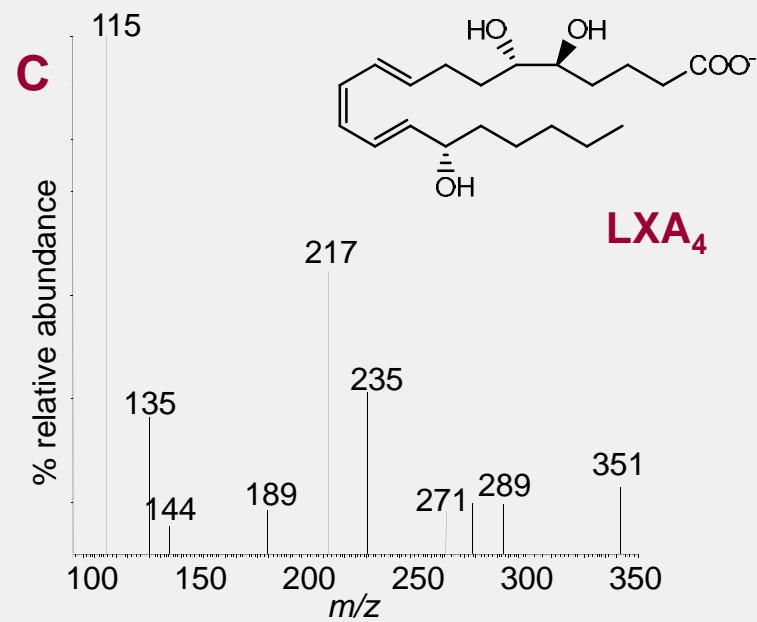
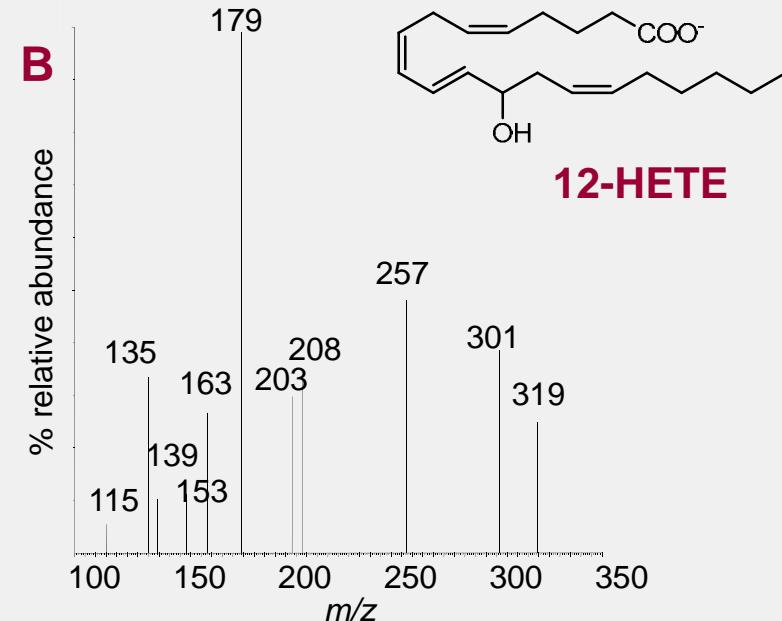
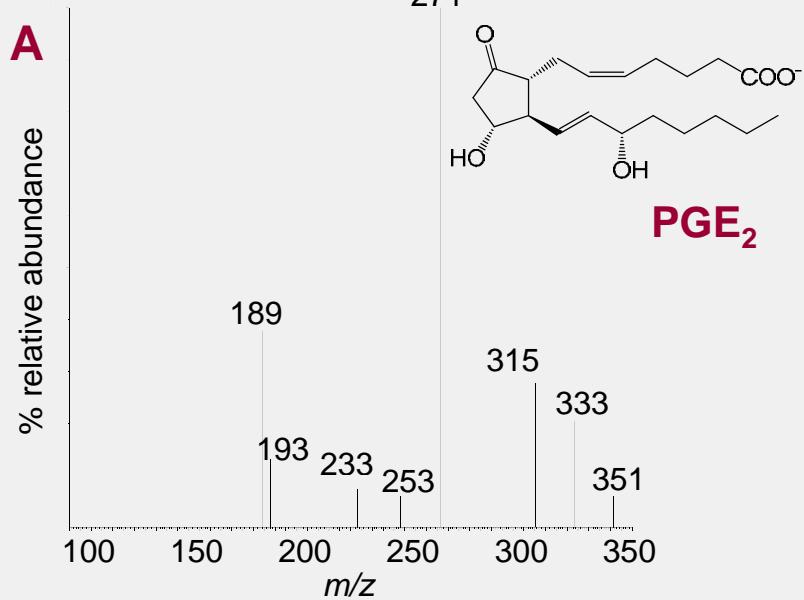
4. **Elute:** Selectively desorb lipid mediators

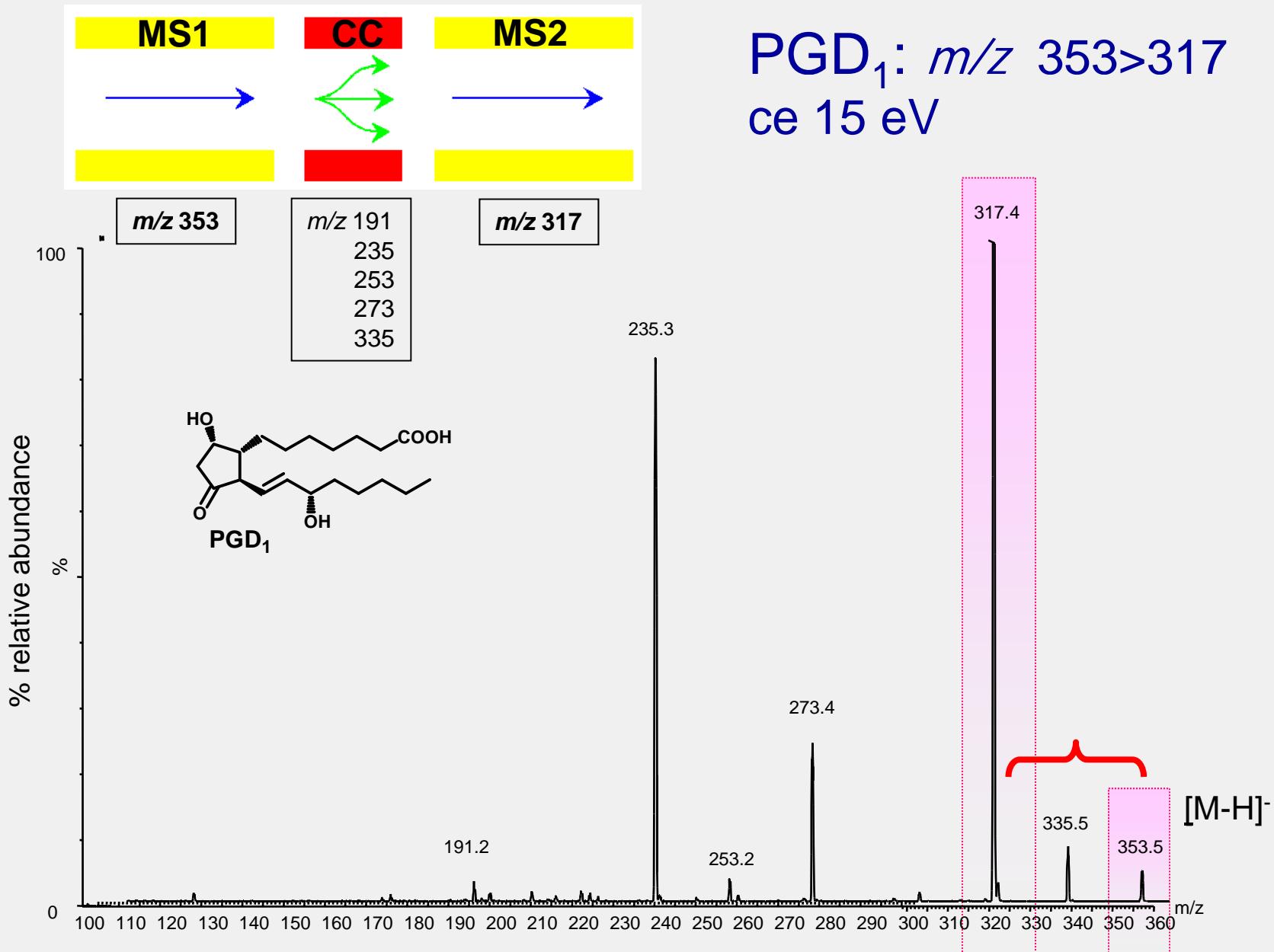
# tandem mass spectrometer

## typical lipidomics platform

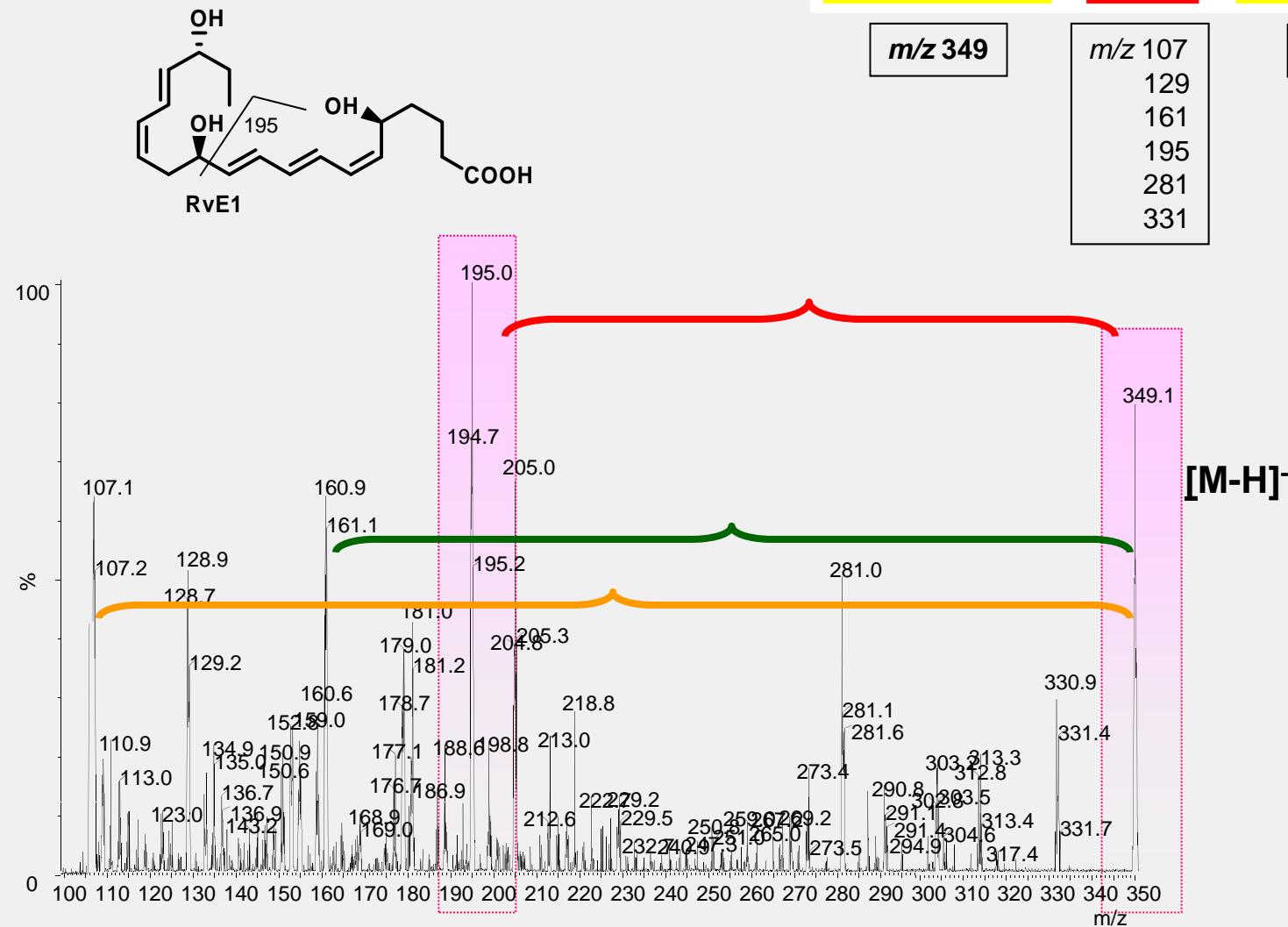
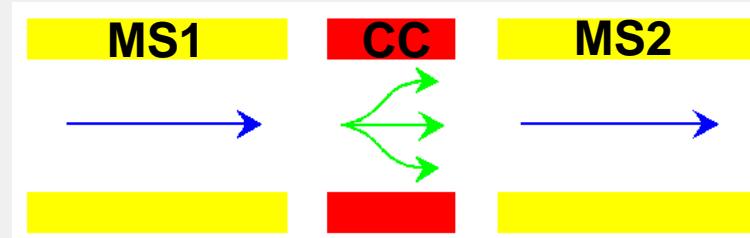
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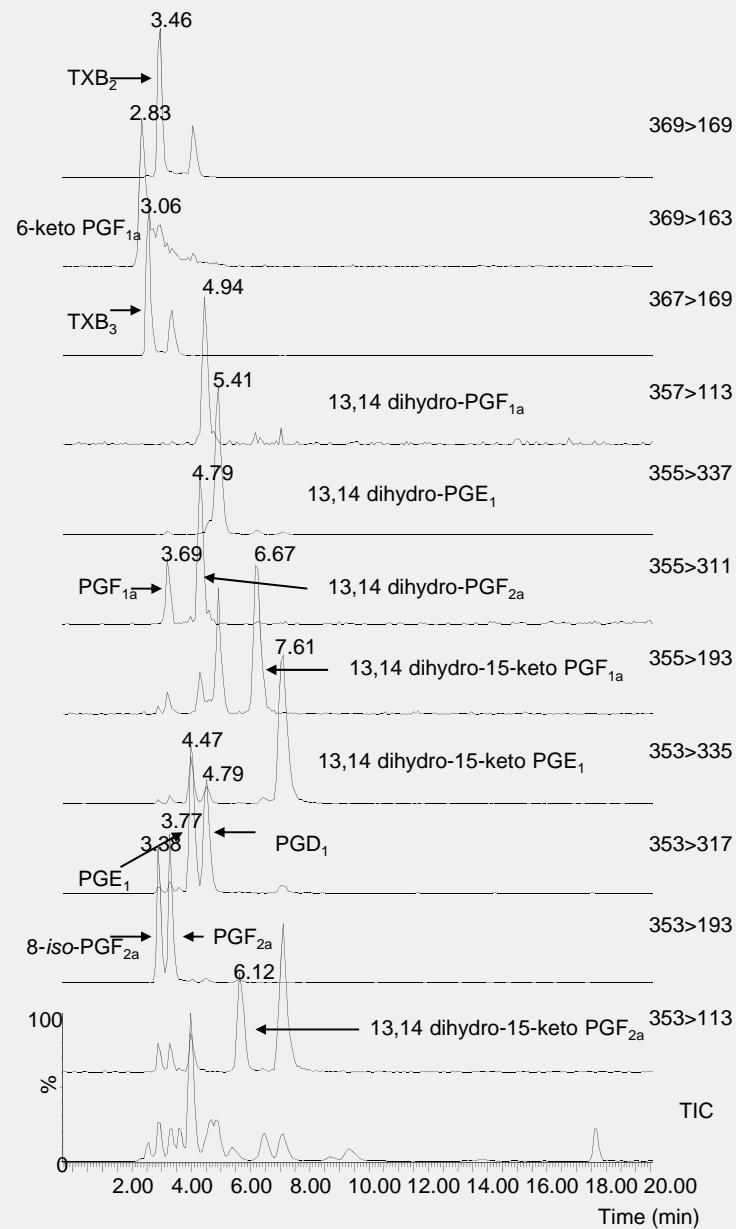




RvE1:  $m/z$  349>195  
ce 17 eV



## LC/ESI-MS/MS (ESI-)

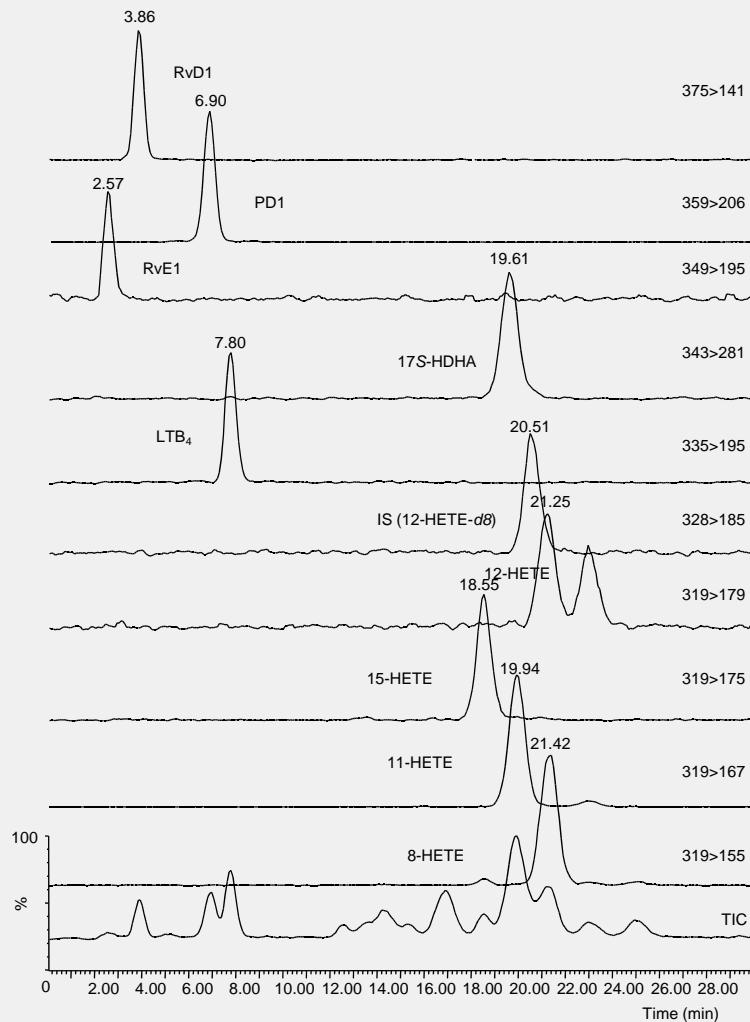


# Liquid chromatography: reverse phase

Lipid mediators typically separated by hydrophobic moiety (C18, e.g. Luna ®)

**Prostanoids:**  
isobaric species e.g. PGE and PGD  
optimal separation: acetonitrile-based gradient elution system

## LC/ESI-MS/MS (ESI-)



# Liquid chromatography: reverse phase

Lipid mediators typically separated by hydrophobic moiety (C18, e.g. Luna ®)

**Hydroxy fatty acids:**  
poor resolution with acetonitrile  
strong interaction with C18 column  
improved elution with methanol

**Core shell columns:** behave like UPLC columns (pore size 2.5μm)  
improved peak resolution and better sensitivity

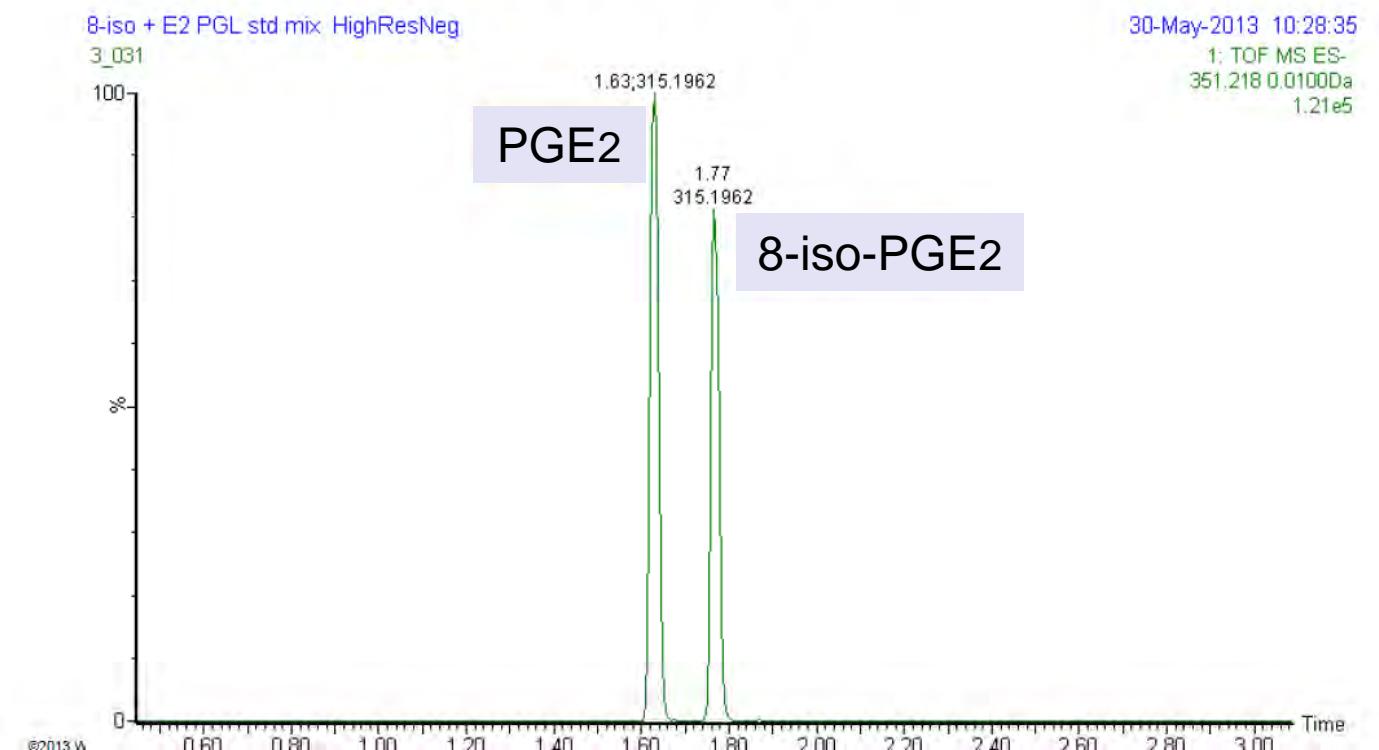
# Very fast even better separation of isobaric compounds with different RP UPC2 column

## PGL-E2 Chromatographyc separation BEH-2EP column

Waters

THE SCIENCE OF WHAT'S POSSIBLE.™

- Significantly better chromatographic resolution (1.65 measures as Rs half-height) was achieved on a BEH-2EP column:



# Chiral separation by LC

## Stationary phases:

Amylose: 18(S)-E Resolvins (Oh et al., J Clin Invest. 2011;121)

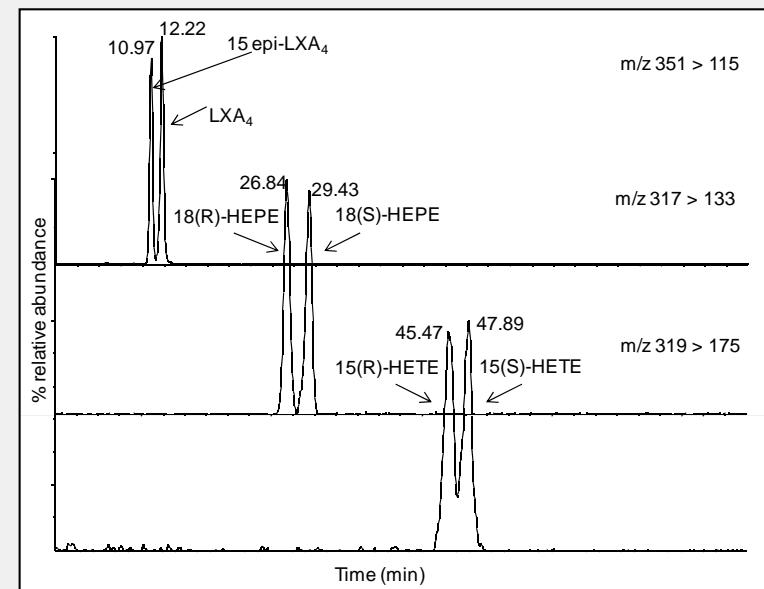
Cellulose:12(S)-HETE in blister fluid (Massey and Nicolaou, FRBM. 2012)

Reverse or normal phase solvents

## Cellulose (Lux-1)

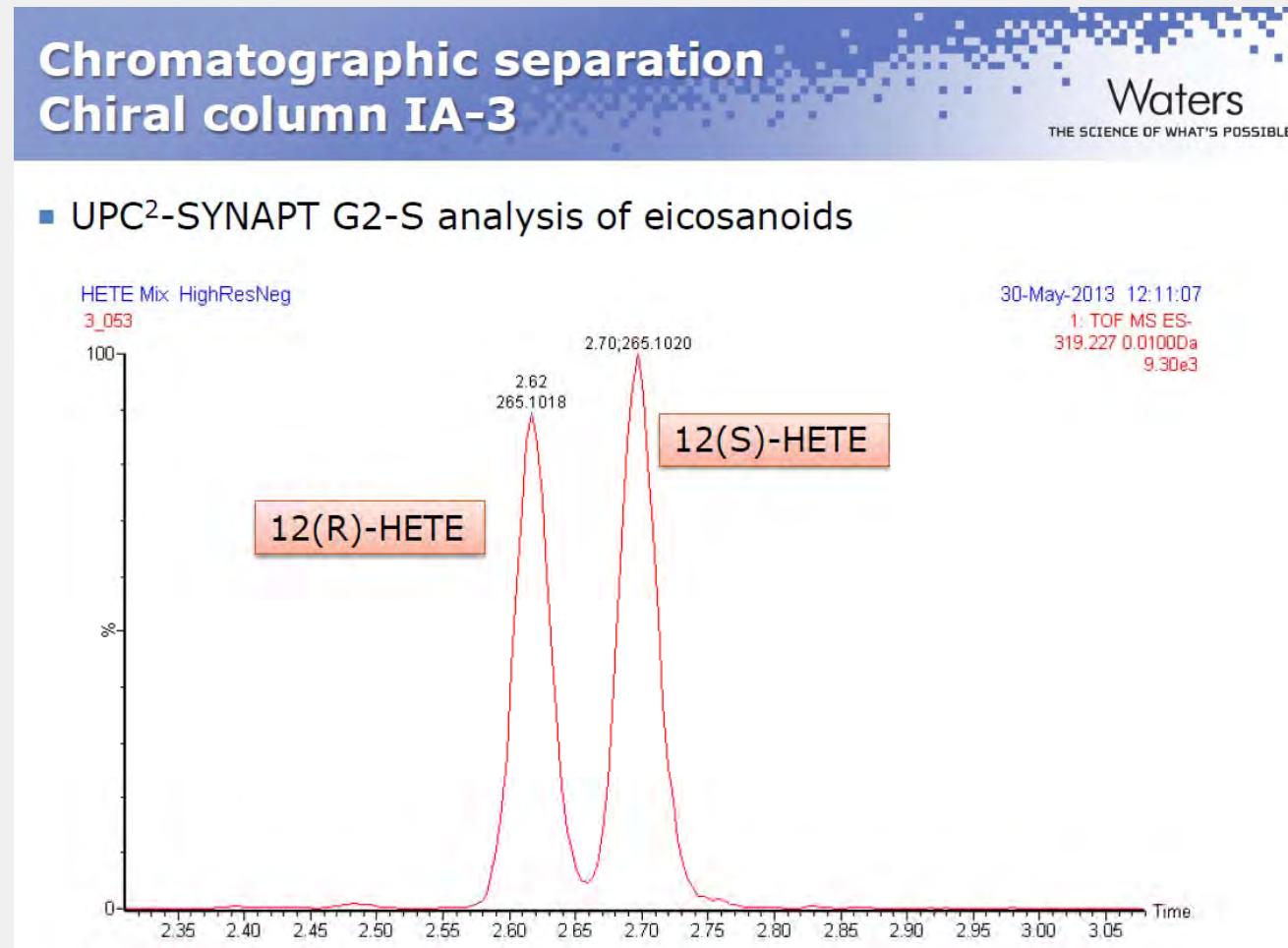
more stable stationary phase

improved separation of enantiomers



Massey& Nicolaou FRBM 2012

# Very fast separation of isobaric compounds with chiral UPC2 column



# mediator lipidomics protocol

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- Solid phase extraction clean-up step (matrix effects).
- Multiple Reaction Monitoring (MRM) assays.  
for > 80 lipid mediators; LoD/LoQ 1-10 pg.
- LC/ESI-MS/MS (Q<sup>3</sup>); calibration lines; *d*-internal standards.

## Biological material

- Solids: skin, tumours, liver, brain, uterine, ocular, nerve tissues, cells, etc.
- Liquids: plasma, urine, seminal plasma, follicular fluid, blister fluid, cell culture media, etc.
- Samples snap-frozen; extracted/run within days; dark/cold.

# **lipid mediators in skin inflammation: the sunburn response**

# inflammation in cutaneous disease

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psoriasis



atopic dermatitis



wound healing



photoageing



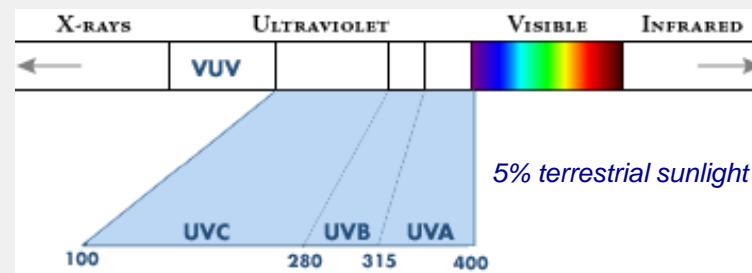
sunburn



squamous cell  
skin cancer

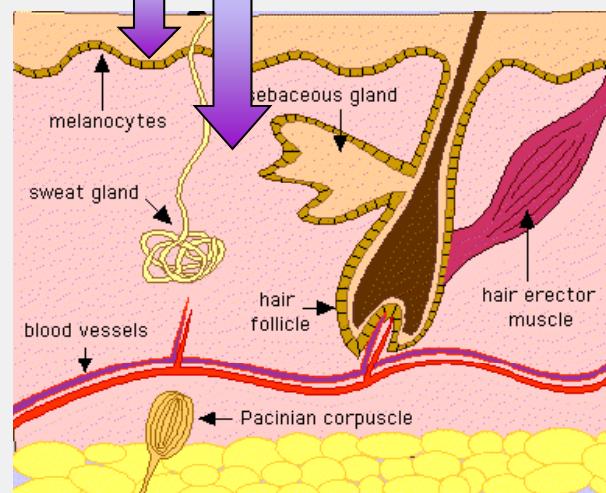


# UV radiation and human skin



**UVA: 320-400 nm**  
**dermis**

**UVB: 280-320 nm**  
**epidermis: *inflammation***



**epidermis**  
protection: radiation, xenobiotics, trauma

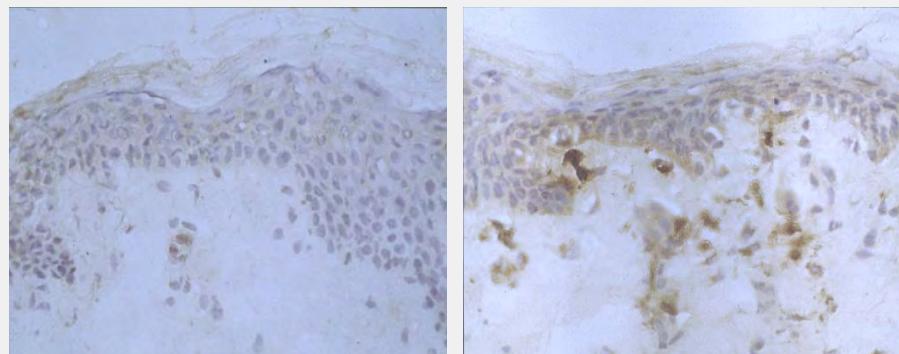
**dermis**  
elasticity; immuno- & biochemical support

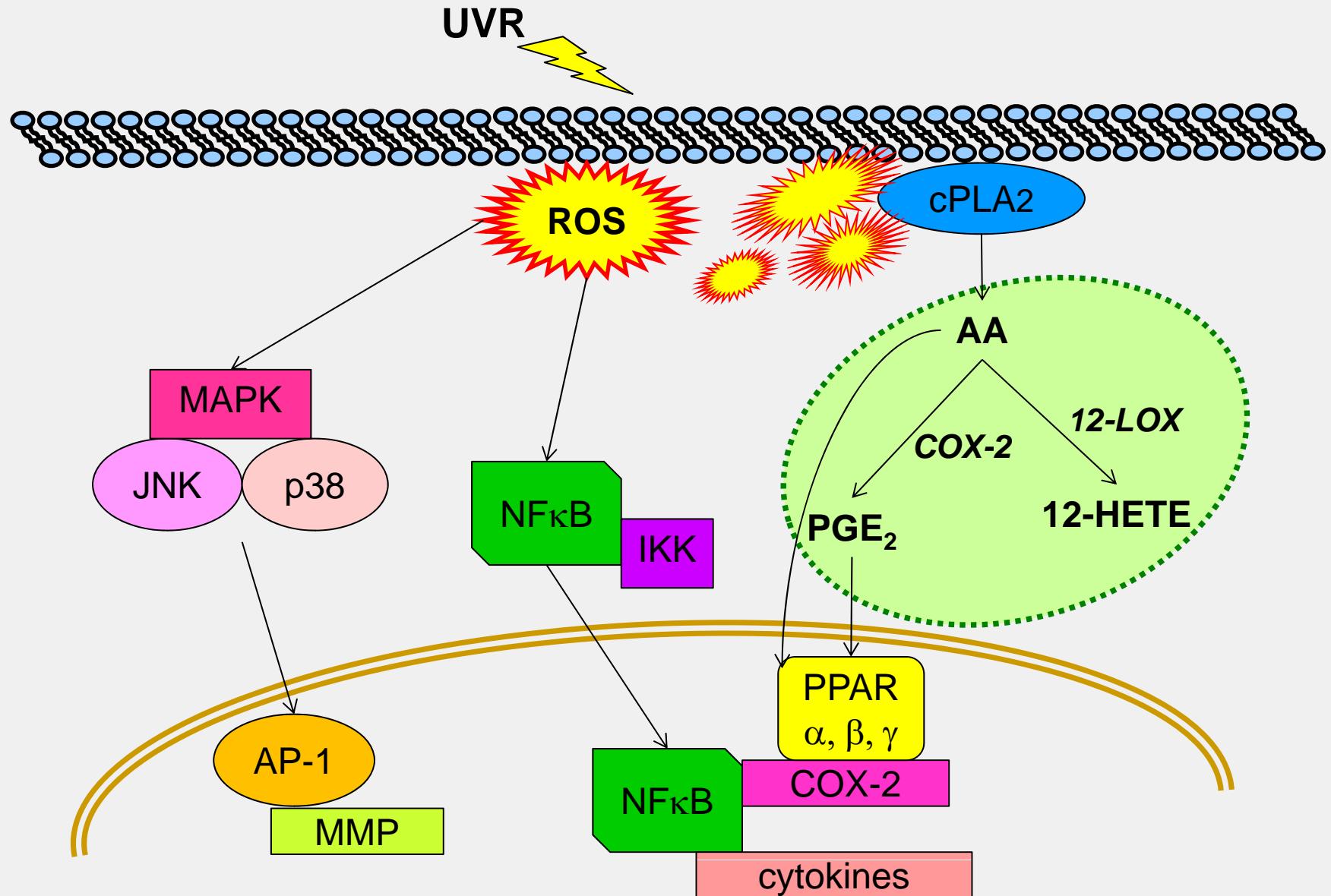
**UVR:** immunosuppression; photosensitivity; photoageing; photocarcinogenesis

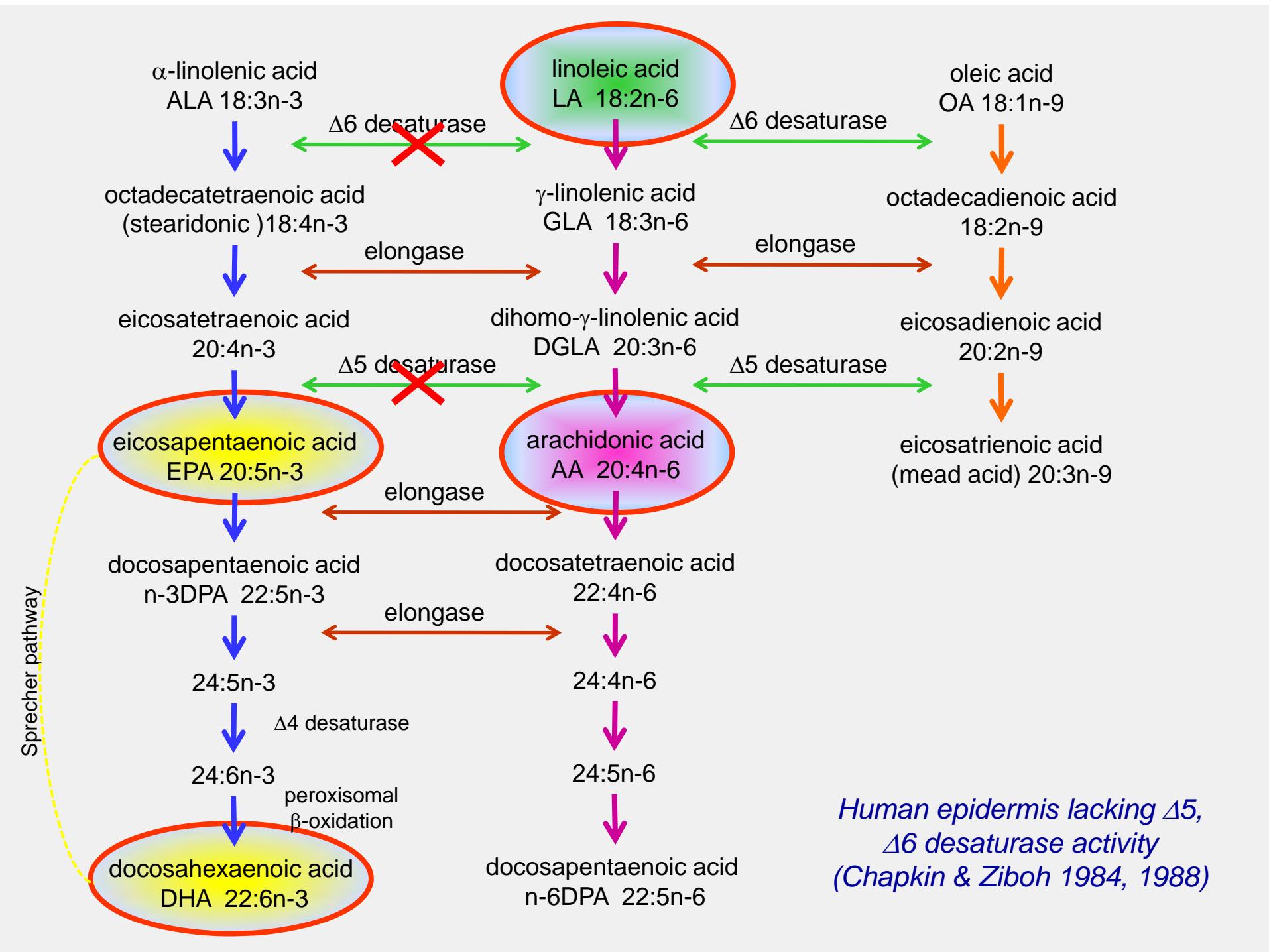
# UVR-induced skin inflammation (sunburn)

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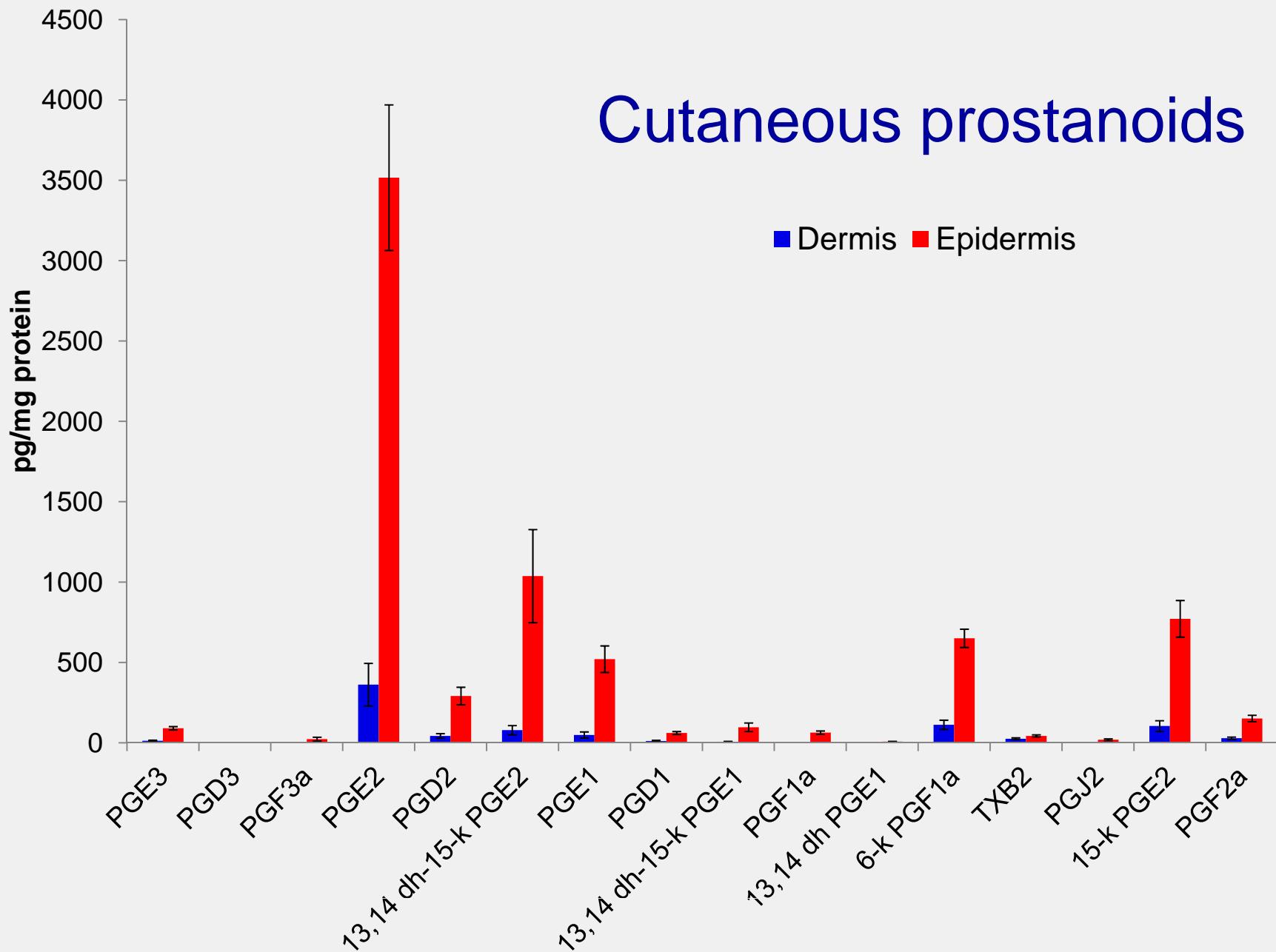
- Acute inflammatory response
- Erythema, pain, oedema (vasodilatation)
- Leukocytic infiltration
- Sunburn (apoptotic) cells





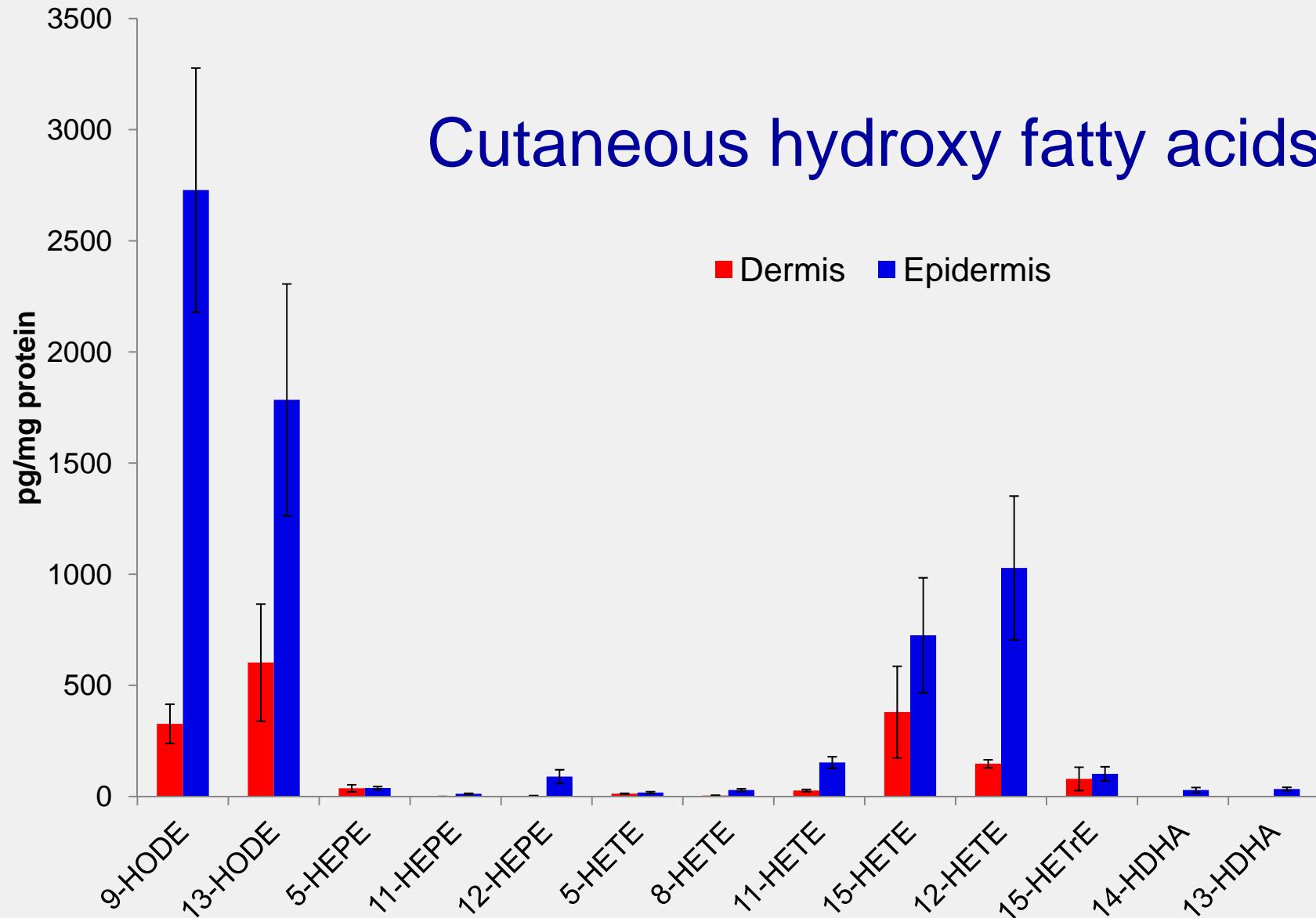


# Cutaneous prostanoids



human skin samples: ethical tissue; 3 mm punch biopsies; ~20 mg; n=8; LC/ESI-MS/MS

# Cutaneous hydroxy fatty acids



human skin samples: ethical tissue; 3 mm punch biopsies; ~20 mg; n=8; LC/ESI-MS/MS

# sunburn: experimental model

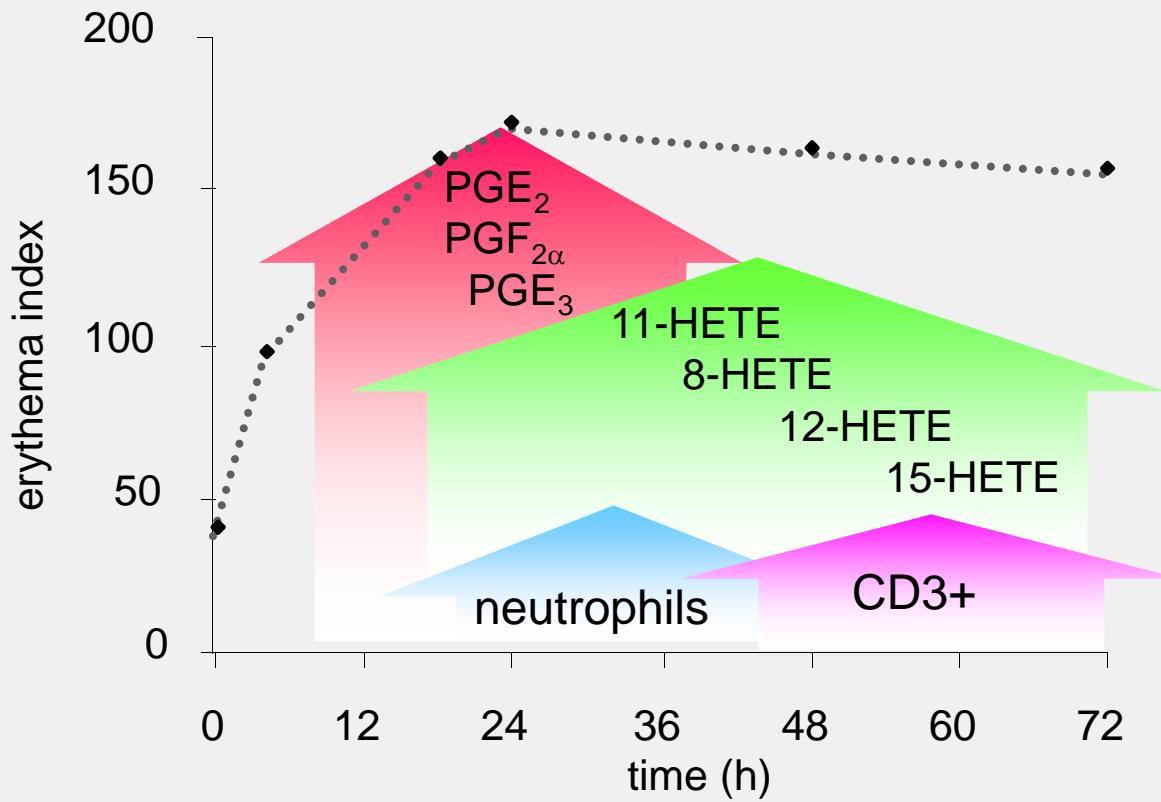
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- healthy adult volunteers, skin type I-IV
- skin exposed to UVR  
(UV6; 280-400 nm; 23% UVB :77% UVA)
- 3-4 minimal erythema doses (MED)



Suction blisters and skin sections (0 -72 h post UVR)

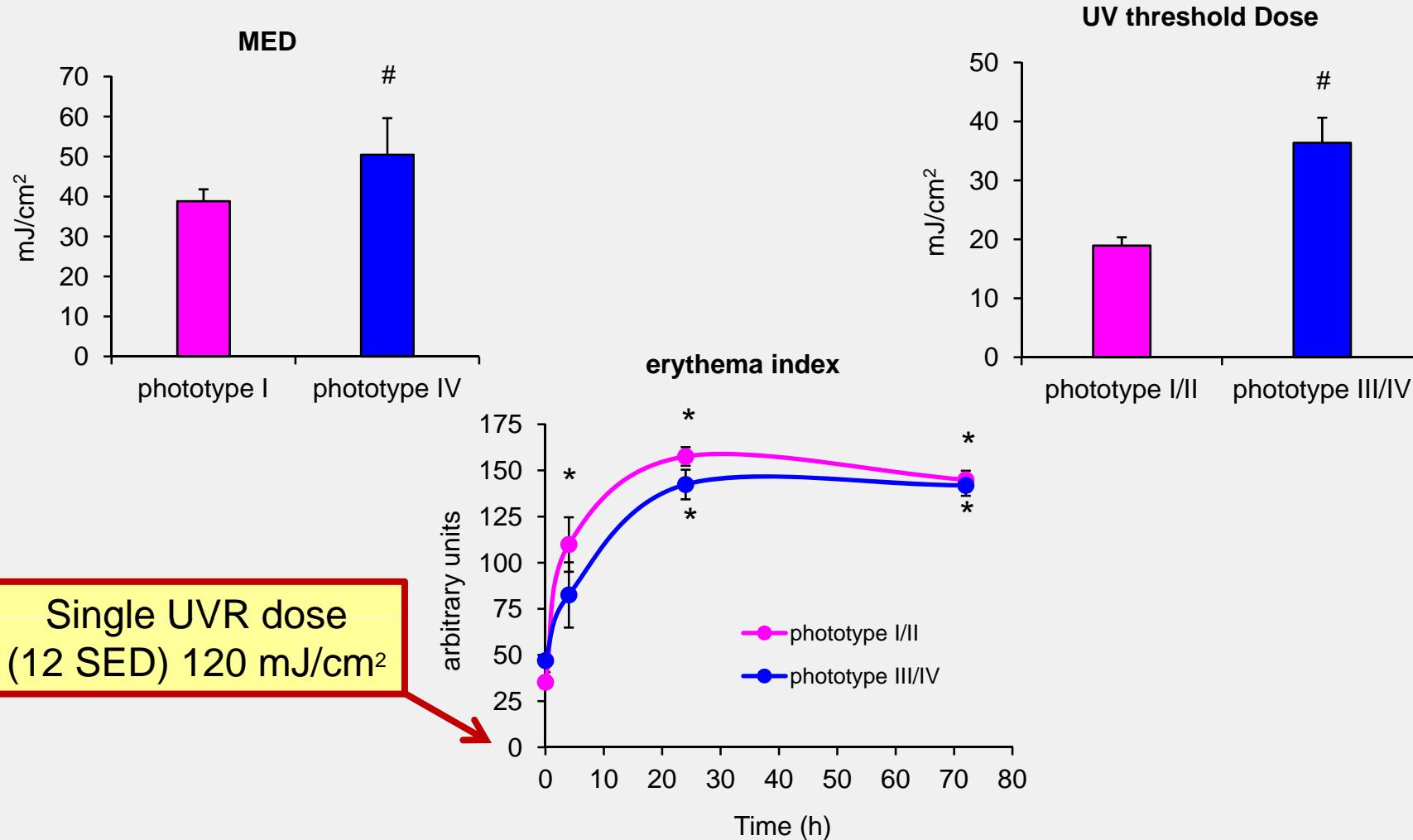
# Overlapping sequential eicosanoid profiles may mediate the early and late phases of sunburn response



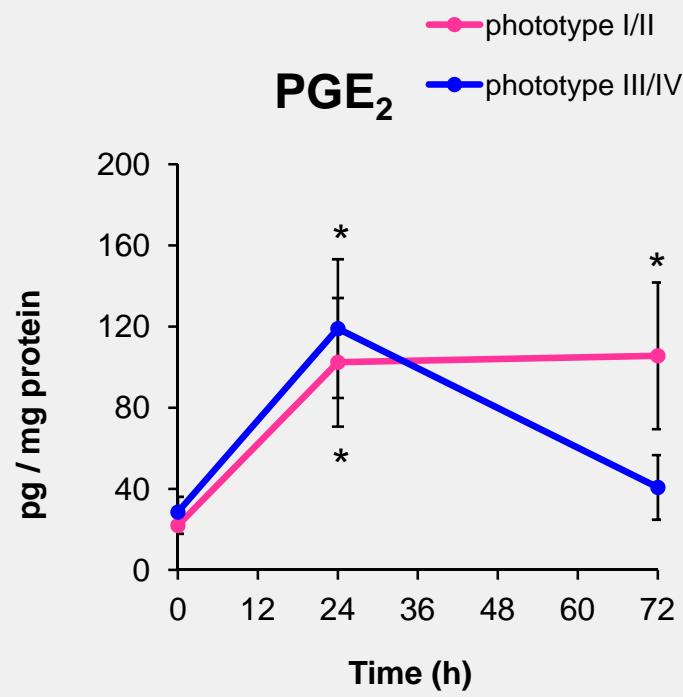
**early:** inflammation  
vasodilatation, chemotaxis

**late:** resolution  
repair

# erythema in skin types I/II and III/IV post single high UVR dose (12 SED)

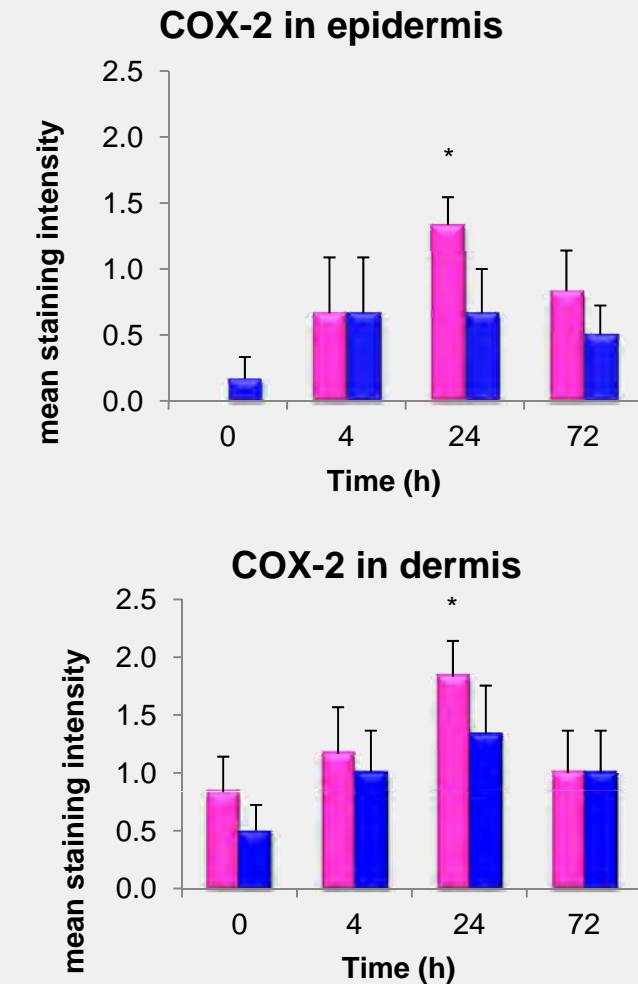


# PGE<sub>2</sub> higher in subjects prone to sunburn



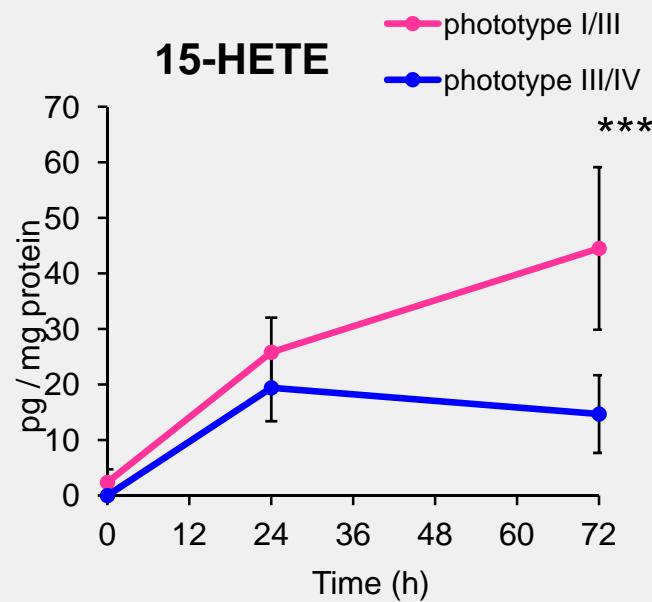
pro-inflammatory  
vasodilatation

n=9; p<0.05



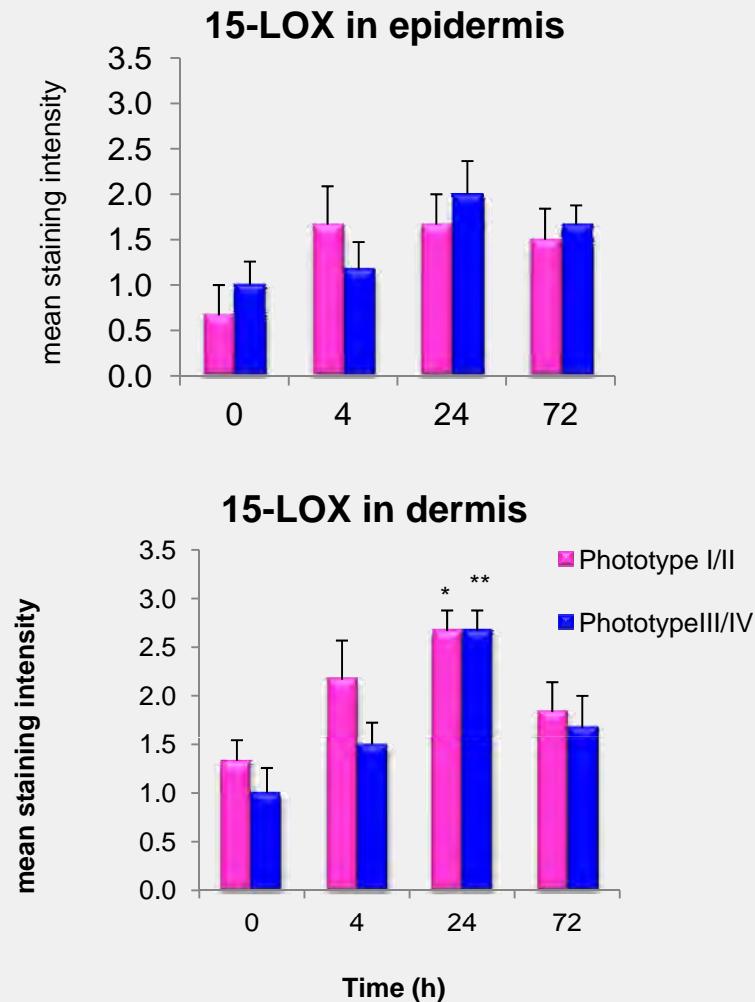
Nicolaou et al *Photochem Photobiol Sci* 2012

# 15-HETE higher in subjects prone to sunburn



anti-inflammatory  
precursor to lipoxins

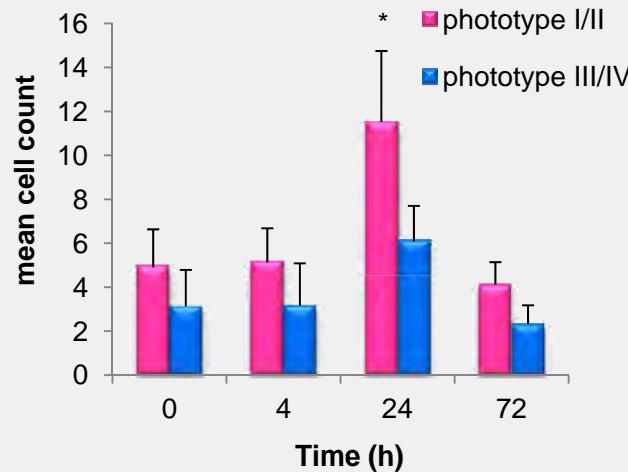
n=9; \*p<0.05, \*\*\*p<0.001



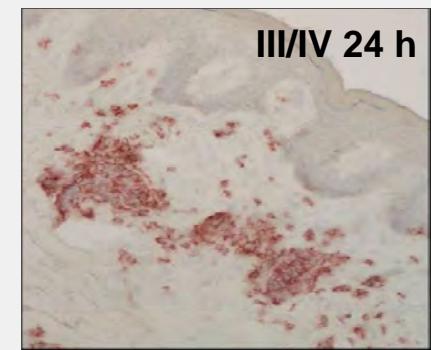
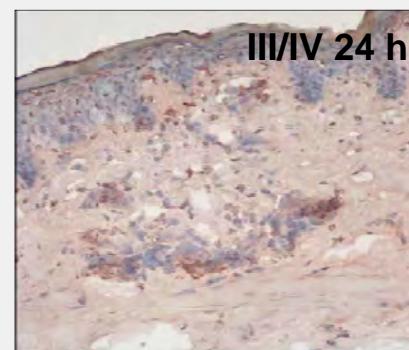
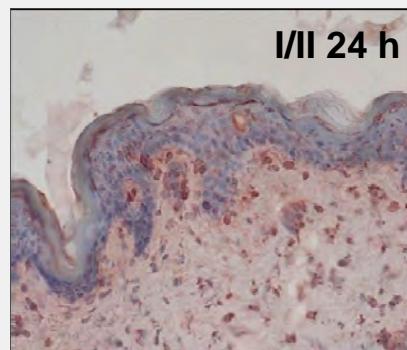
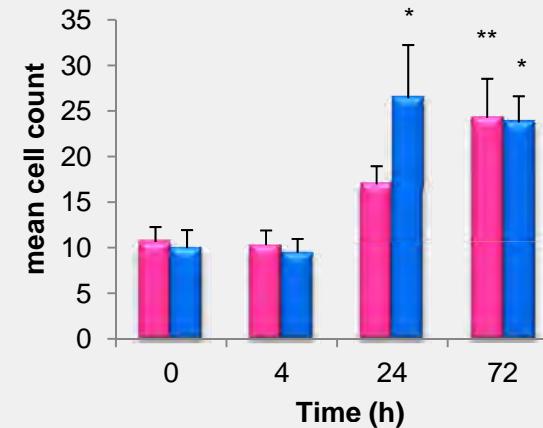
Nicolaou et al *Photochem Photobiol Sci* 2012

# higher neutrophil infiltrate in subjects prone to sunburn

neutrophils in dermis



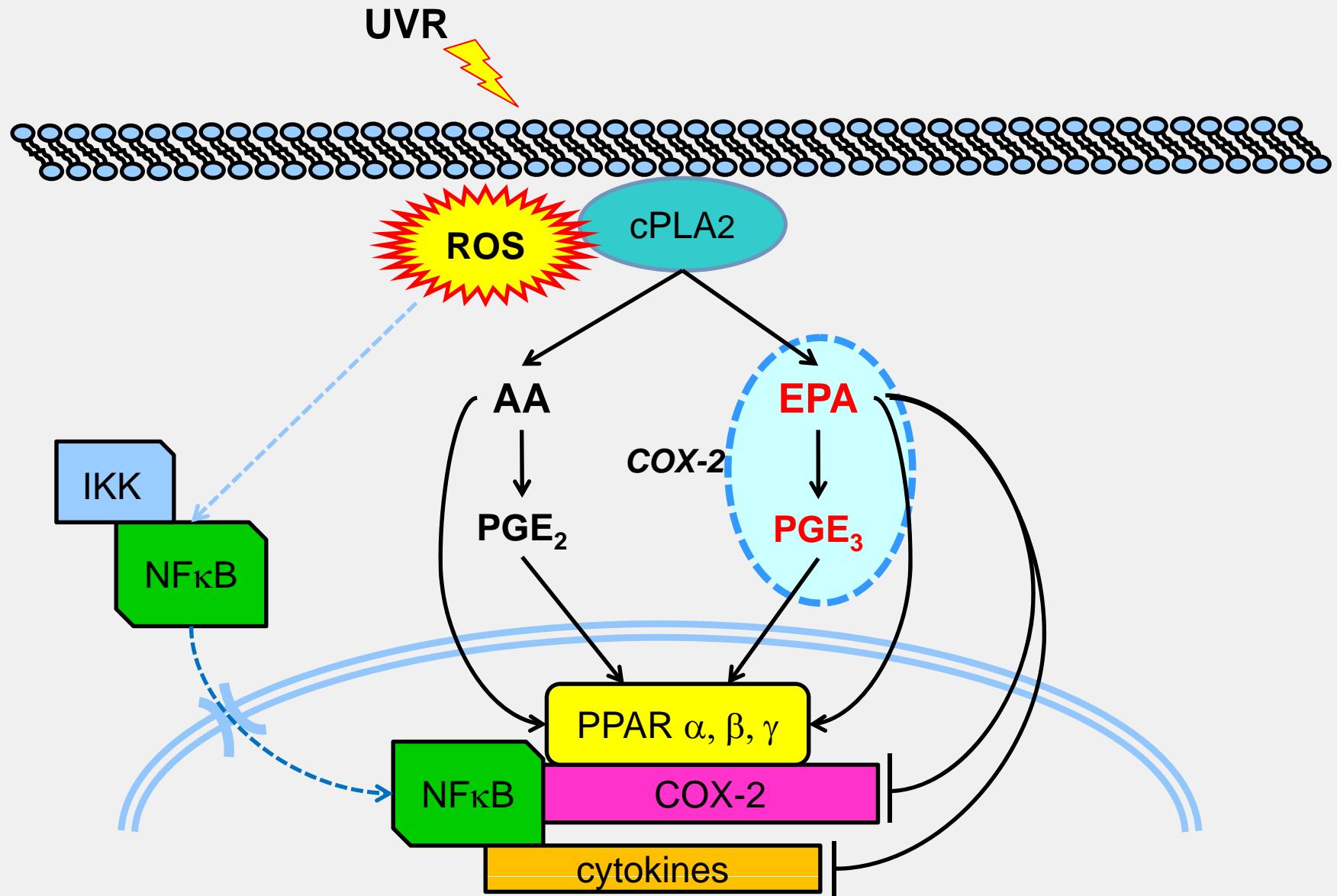
CD3+ cells in dermis



n=6; \* p<0.05; \*\* p<0.01

Nicolaou et al *Photochem Photobiol Sci* 2012

# **lipid biomarkers of skin inflammation in human nutritional studies**



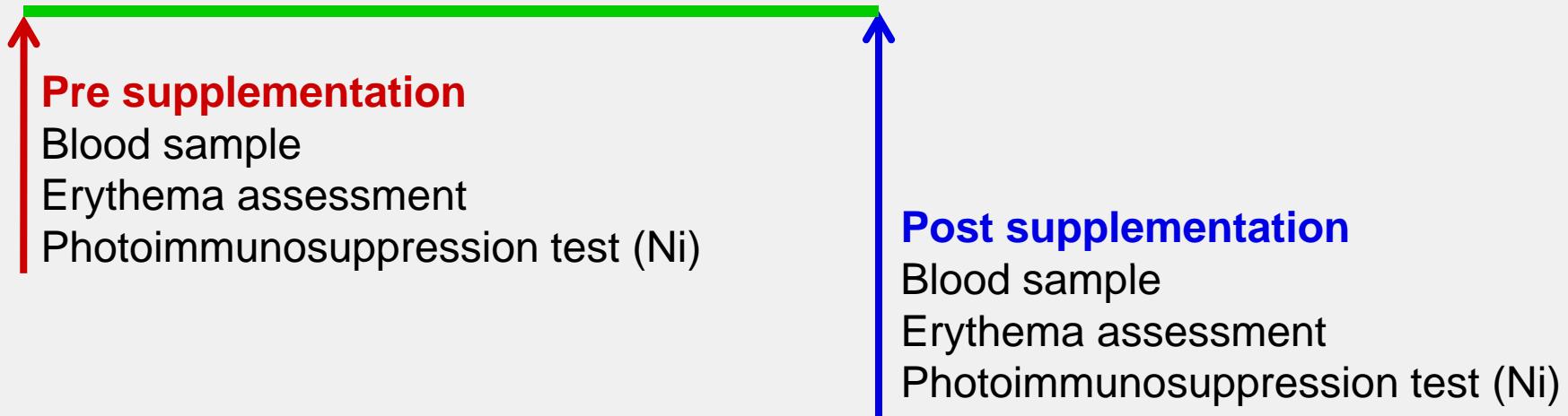
# n-3PUFA in skin inflammation and immunity: photoimmunosuppression

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Randomised double-blind study (n=79 subjects)

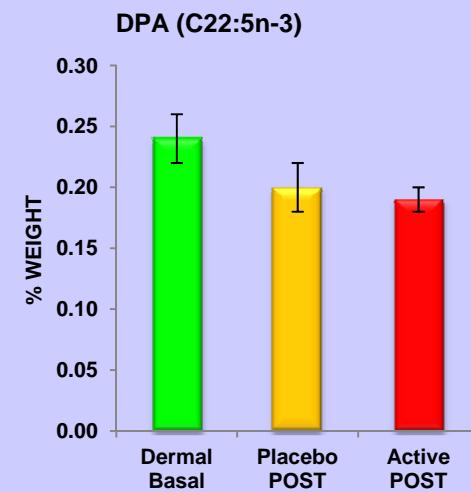
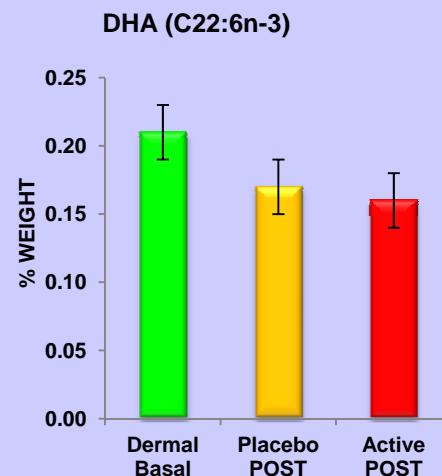
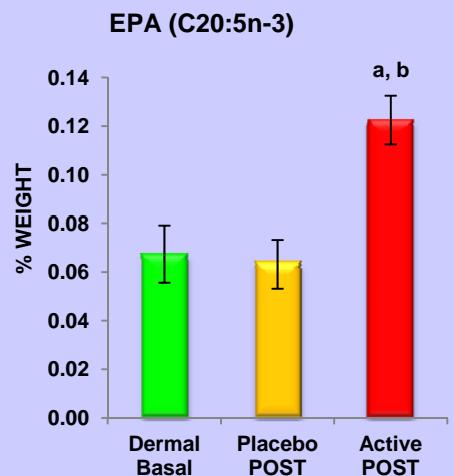
**control:** GTCC

**active:** 1g capsule~70% EPA&10% DHA; 5 cps/day; 12 weeks

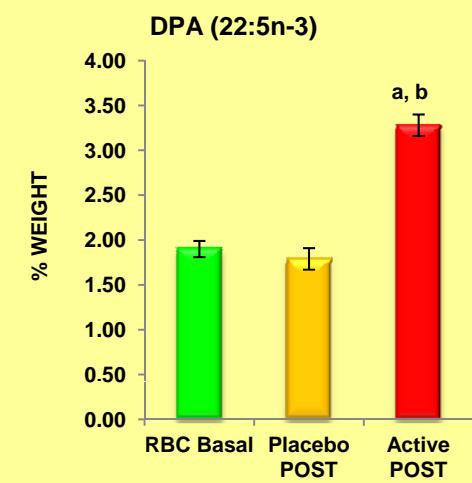
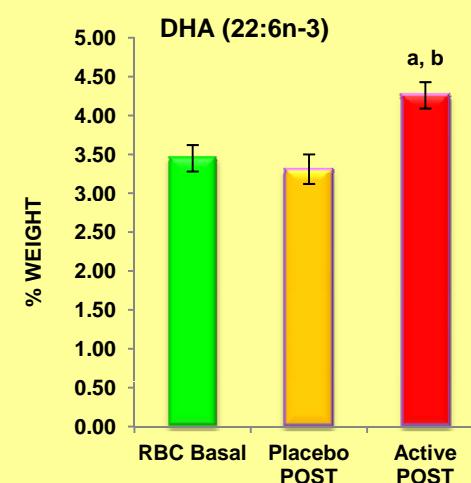
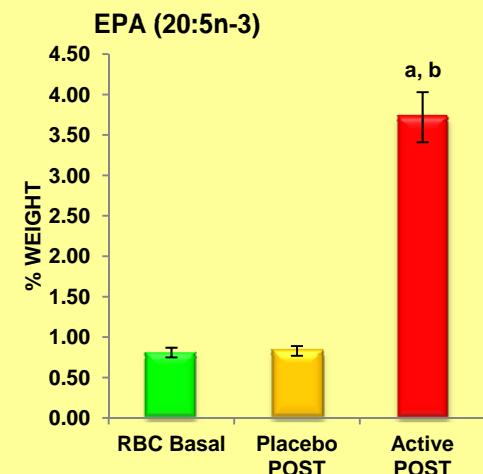


# EPA supplementation did not increase skin DPA or DHA levels

skin



r.b.c.

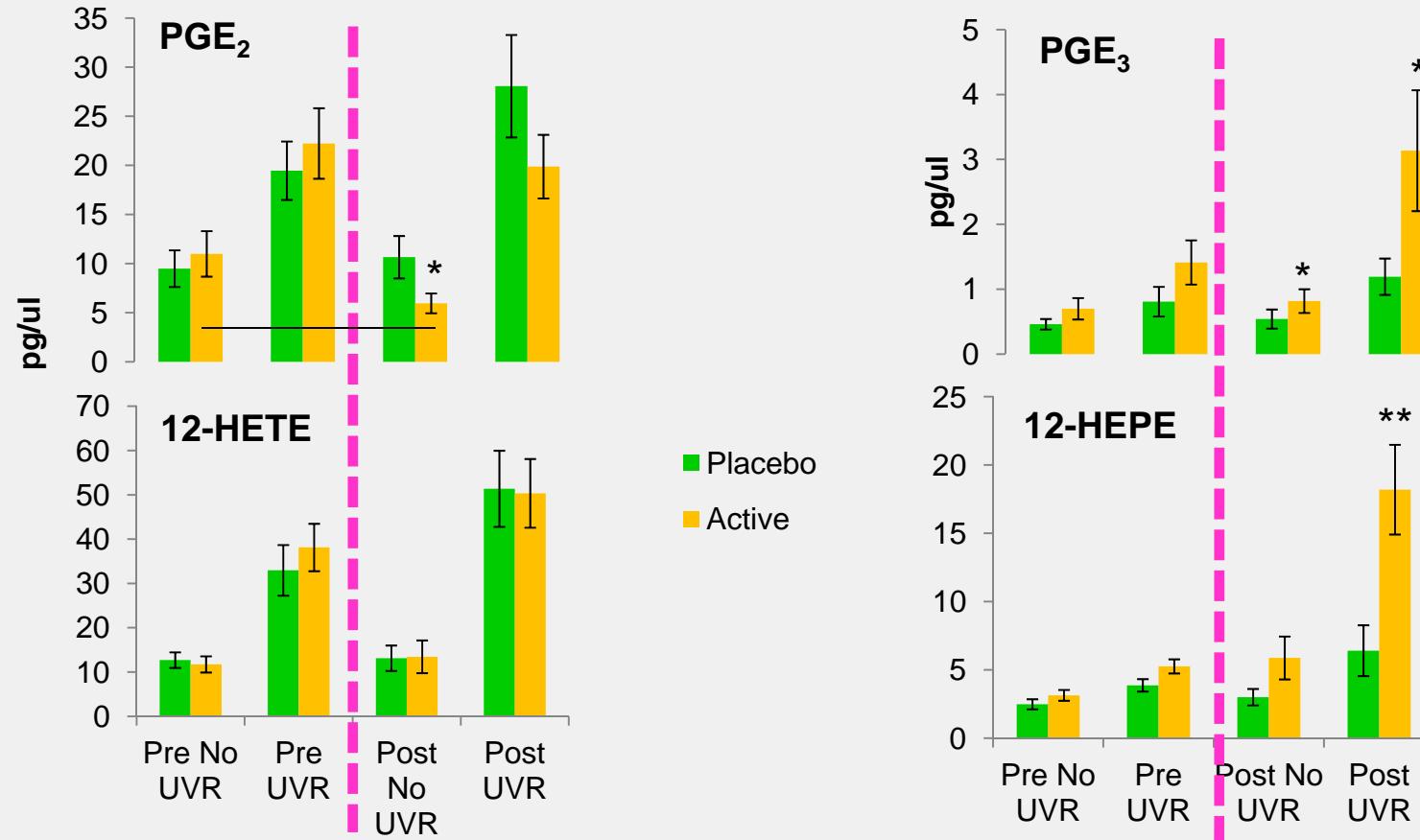


a: p<0.001 comparing to basal; b:p<0.001 comparing to placebo

# AA, EPA, OA mediators in cutaneous blister fluid

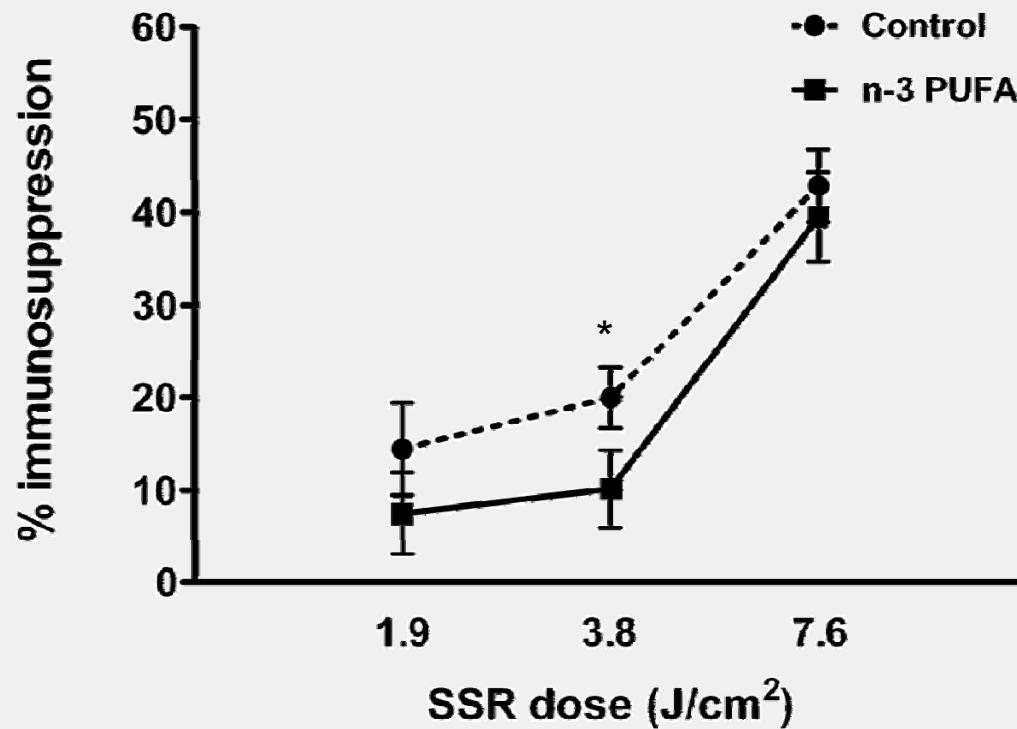
	Mean (SEM) (pg/μl)							
	Baseline				12 weeks			
	Control (n=19)		EPA (n=17)		Control (n=19)		EPA (n=17)	
	Unexposed	UVR-exposed	Unexposed	UVR-exposed	Unexposed	UVR-exposed	Unexposed	UVR-exposed
<b>PGE<sub>2</sub></b>	9.5 (1.9)	19.5 (3.1)†††	11.0 (2.4)	22.2 (3.8)†	10.7 (2.2)	28.1 (5.4)††	6.0 (1.1)*	19.9 (3.4)†††
<b>PGE<sub>3</sub></b>	0.5 (0.1)	0.8 (0.2)	0.7 (0.2)	1.6 (0.4)†	0.6 (0.2)	1.2 (0.3)†	0.8 (0.2)	3.1 (1.0)†
<b>PGE<sub>1</sub></b>	2.7 (0.7)	6.2 (1.2)†††	2.6 (0.6)	7.0 (1.2)††	3.5 (1.4)	8.7 (2.0)††	1.6 (0.4)	6.7 (1.4)†††
<b>13,14 dh-15k-PGE<sub>2</sub></b>	4.6 (1.1)	1.2 (0.4)†††	8.1 (2.2)	1.5 (0.4)†††	4.8 (1.3)	1.4 (0.4)†††	4.9 (1.4)	1.9 (0.4)
<b>12-HETE</b>	12.7 (1.8)	33.0 (5.7)†††	11.7 (1.9)	38.1 (5.7)	13.1 (2.9)	51.4 (8.6)†††	13.4 (3.9)	50.3 (8.2)†††
<b>11-HETE</b>	1.6 (0.2)	3.7 (0.6)†††	1.6 (0.2)	4.3 (0.5)†††	1.4 (0.2)	4.8 (0.5)†††	1.3 (0.3)	4.3 (0.6)†††
<b>15-HETE</b>	3.4 (0.5)	4.6 (0.6)	3.3 (0.5)	6.0 (0.7)††	3.0 (0.5)	6.3 (1.3)††	4.5 (0.9)	6.1 (0.9)†
<b>15-HETrE</b>	0.9 (0.1)	2.4 (0.5)††	1.3 (0.3)	2.2 (0.5)†	0.9 (0.1)	5.4 (2.4)††	0.9 (0.2)	1.9 (0.3)††
<b>12-HEPE</b>	2.5 (0.4)	3.9 (0.5)†	3.1 (0.4)	5.3 (0.5)	3.0 (0.6)	6.4 (1.9)†	5.9 (1.7)	18.2 (3.5)†††**
<b>11-HEPE</b>	ND	0.4 (0.14)a	1.7 (0.9)b	1.7 (0.6)f	7.4 (4.5)b	0.4 (0.05)c	0.6 (0.3)c	4.1 (2.0)g
<b>15-HEPE</b>	ND	ND	ND	ND	ND	ND	3.4 (0.9)d	5.0 (2.2)e
<b>9-HODE</b>	34.3 (5.6)	46.3 (9.6)	45.9 (10.6)	63.7 (7.7)†	26.1 (4.8)	51.1 (9.0)†	32.6 (11.0)	56.1 (9.3)††
<b>13-HODE</b>	36.6 (7.0)	32.6 (5.2)	32.3 (4.4)	55.5 (8.4)†	30.5 (5.9)	33.2 (4.7)	26.3 (5.2)	38.5 (5.9)

# systemic EPA alters skin eicosanoids



RCT: n=16-19 volunteers per group; skin type I/II;  
n-3 LC-PUFA (EPA: 70%; DHA: 10%) 5 cps/d, 3 months

# oral n-3 PUFA supplement protects against UVR-induced immunosuppression



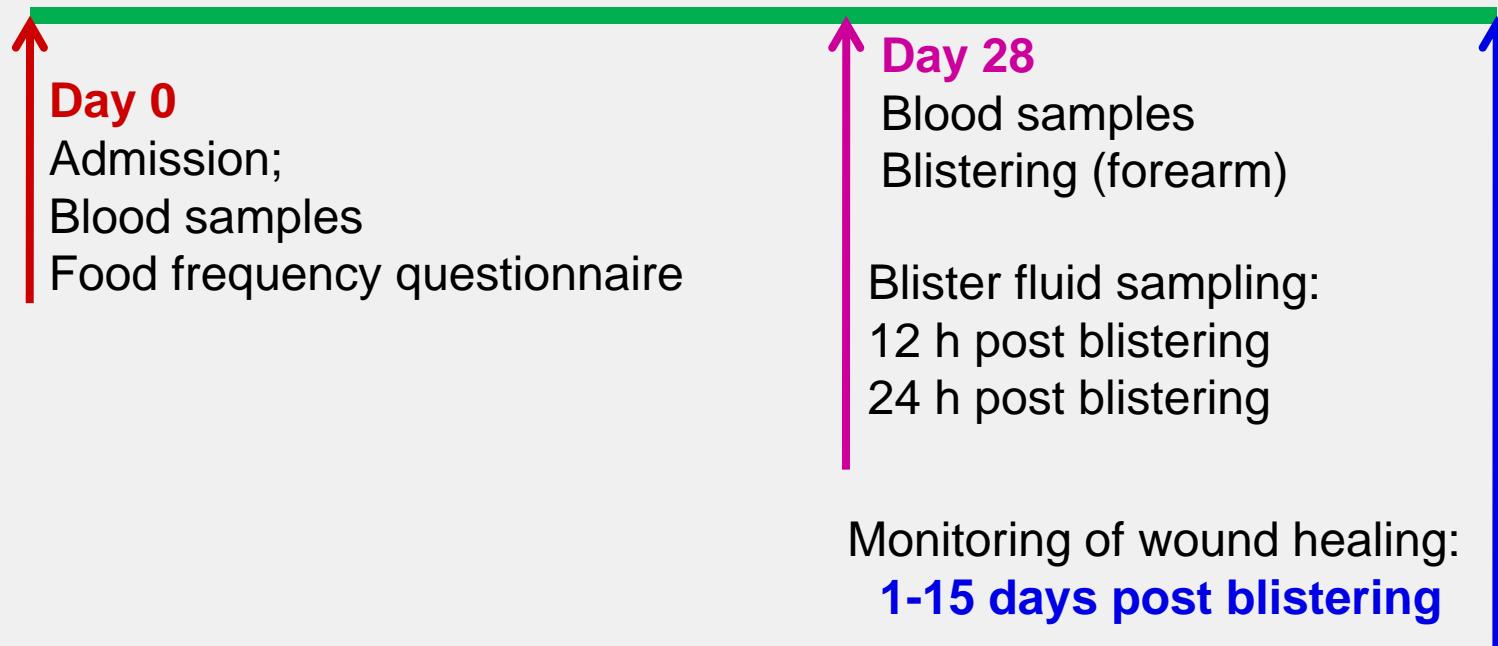
Protection at 3.8 J/cm<sup>2</sup> – 15 min summer midday sun at Manchester

SSR: Solar simulator; nickel allergy; n=33-36 per group; p=0.04

# n-3PUFA in wound healing

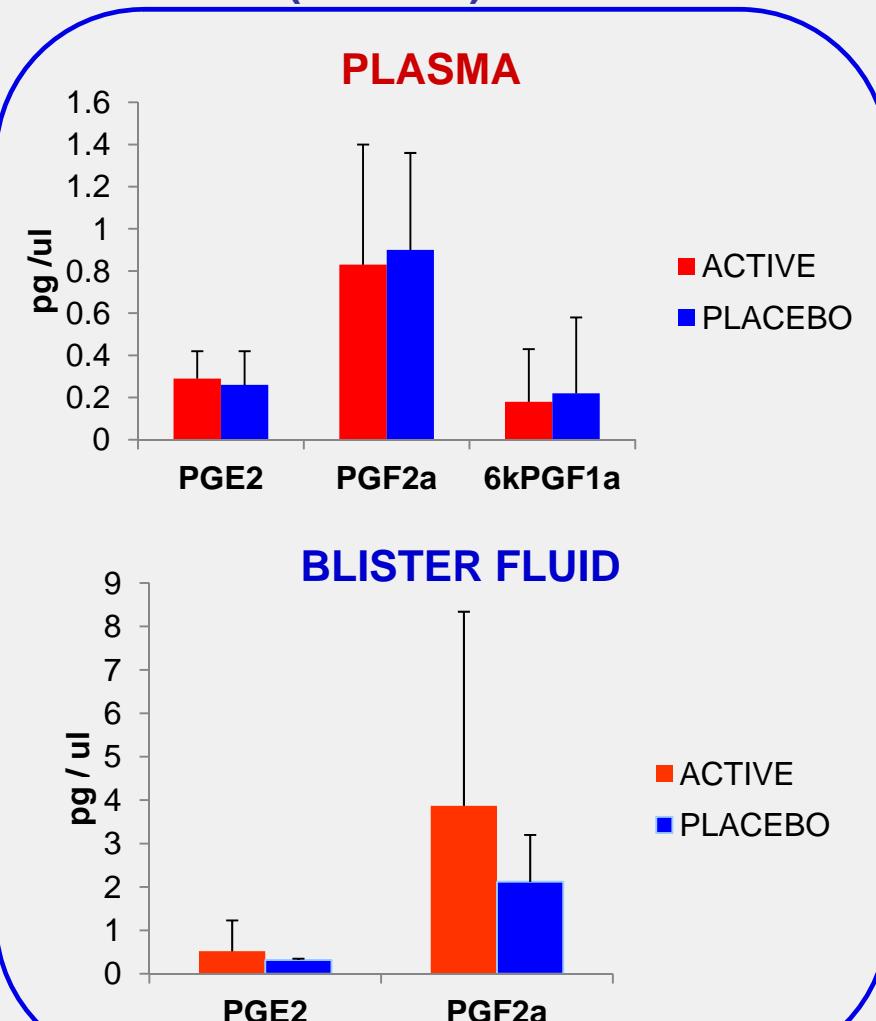
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- Randomised double blind study (n=18 subjects)
  - **placebo** (mineral oil) or
  - **active** (1.6 g EPA+1.2 g DHA/day, 81 mg aspirin) 28 days

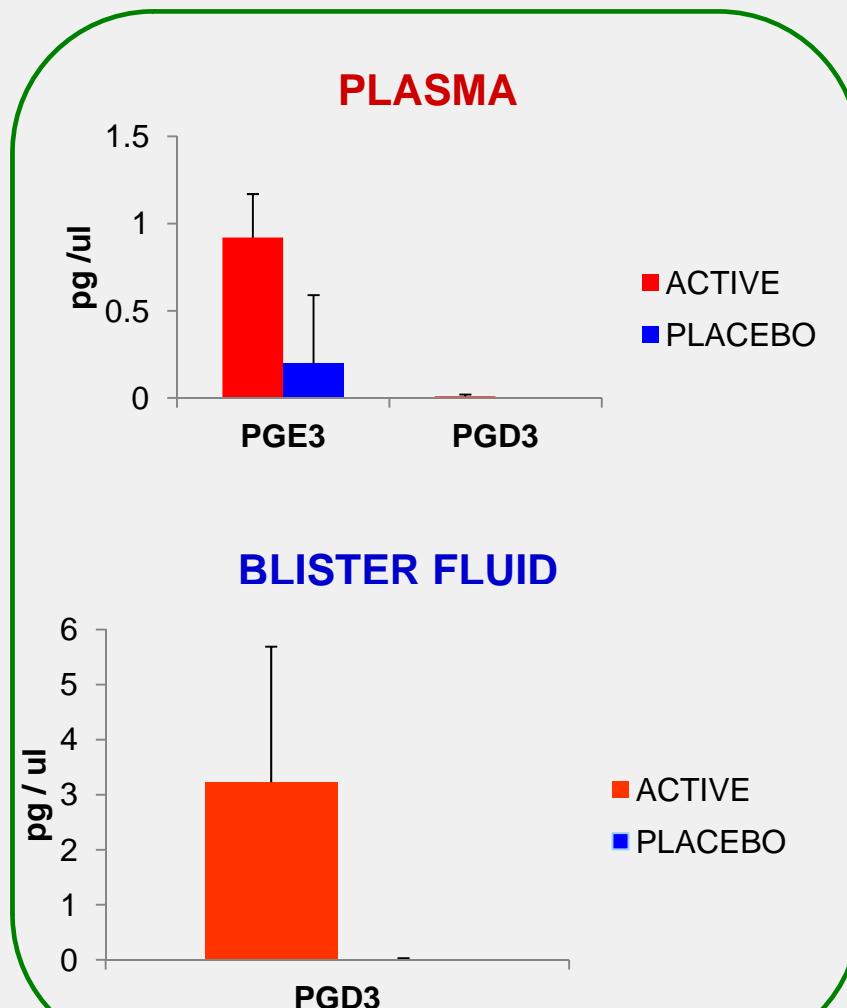


# n-3PUFA supplement and COX-mediators

AA (20:4n-6)-derived

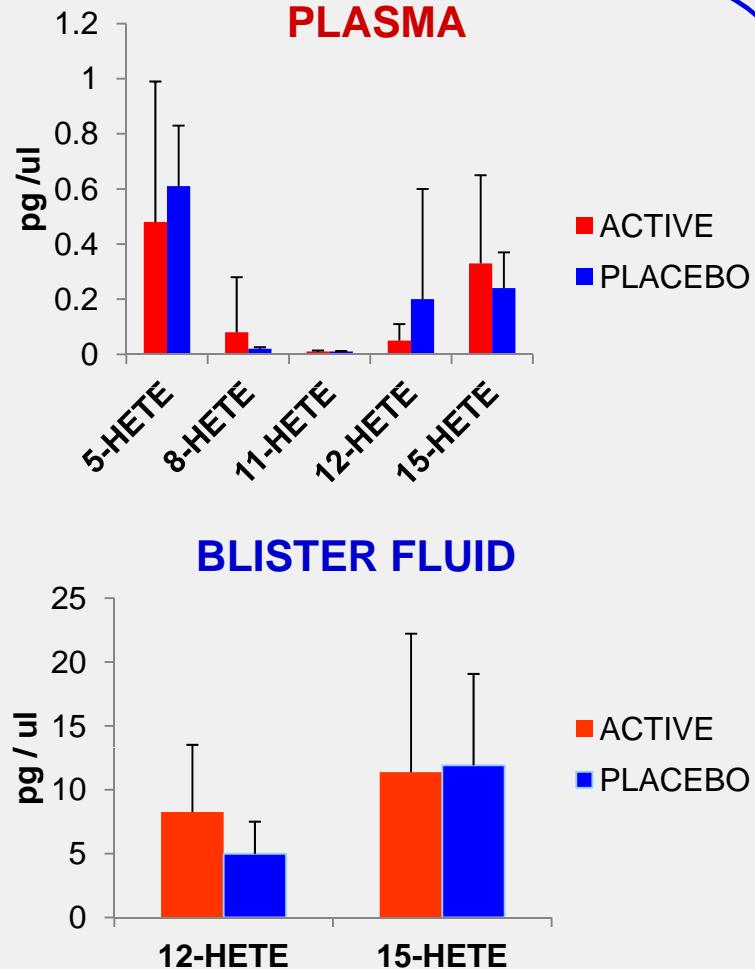


EPA(20:5n-3)-derived

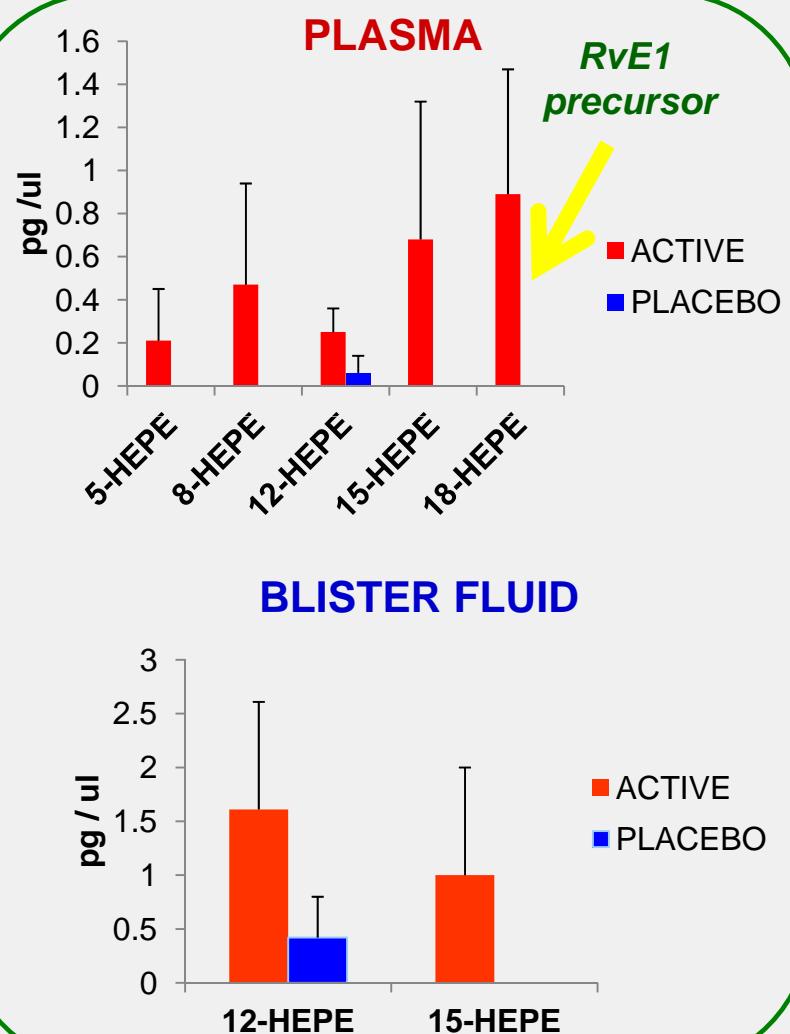


# n-3PUFA supplement and LOX-mediators

AA (20:4n-6)-derived

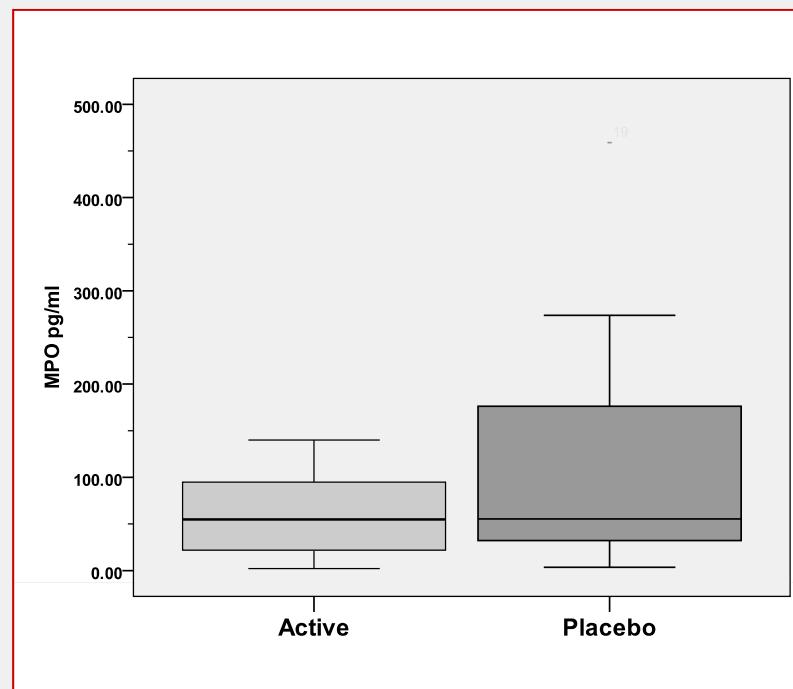


EPA(20:5n-3)-derived

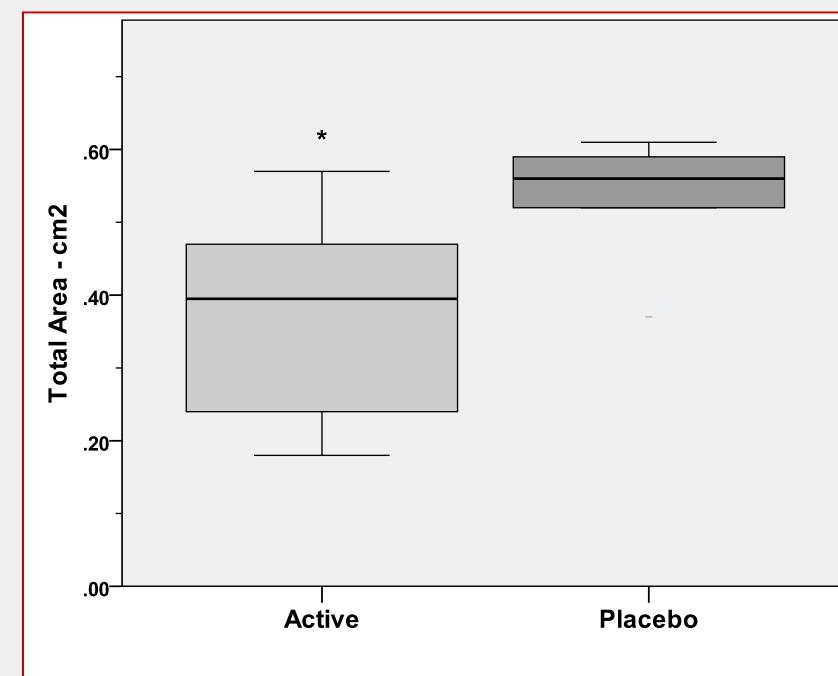


## N-3 PUFA reduced wound area (improved healing)

**myeloperoxidase (MPO) levels,**  
(leukocyte marker enzyme)



**Area of wound (blisters) remaining  
to re-epithelialize on Day 5**

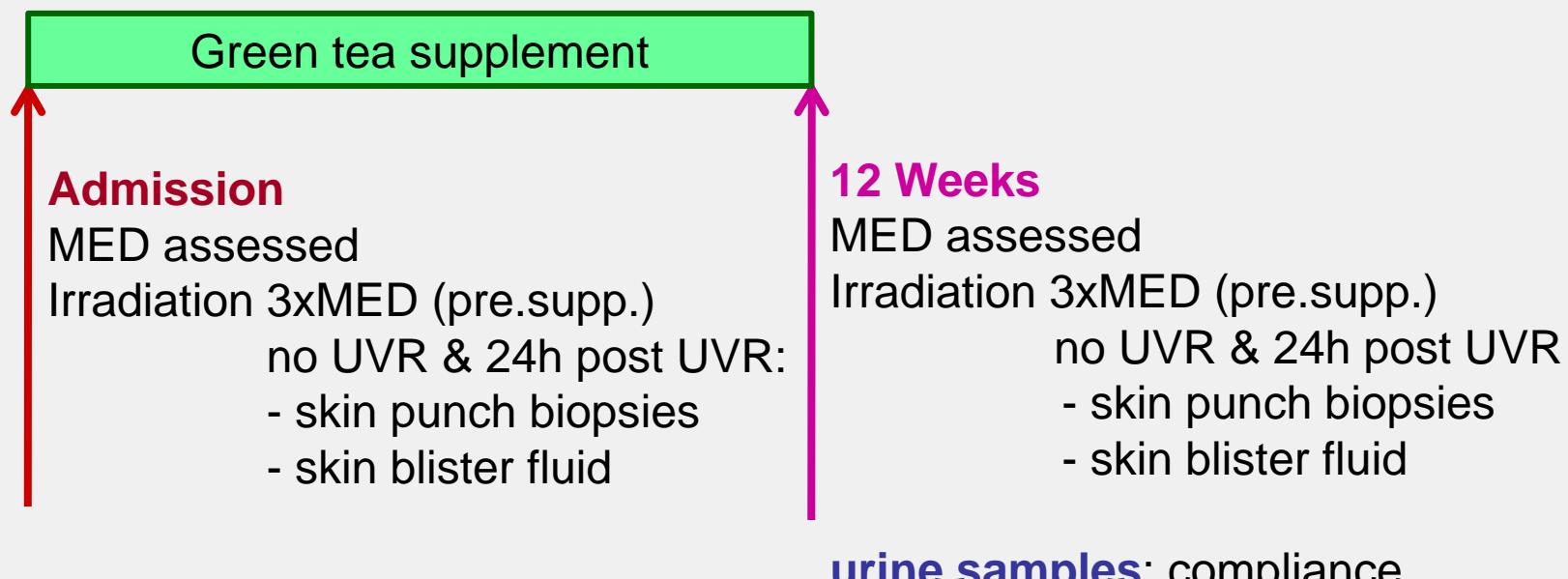


# Green Tea Catechins (GTC) and UVR-induced cutaneous inflammation

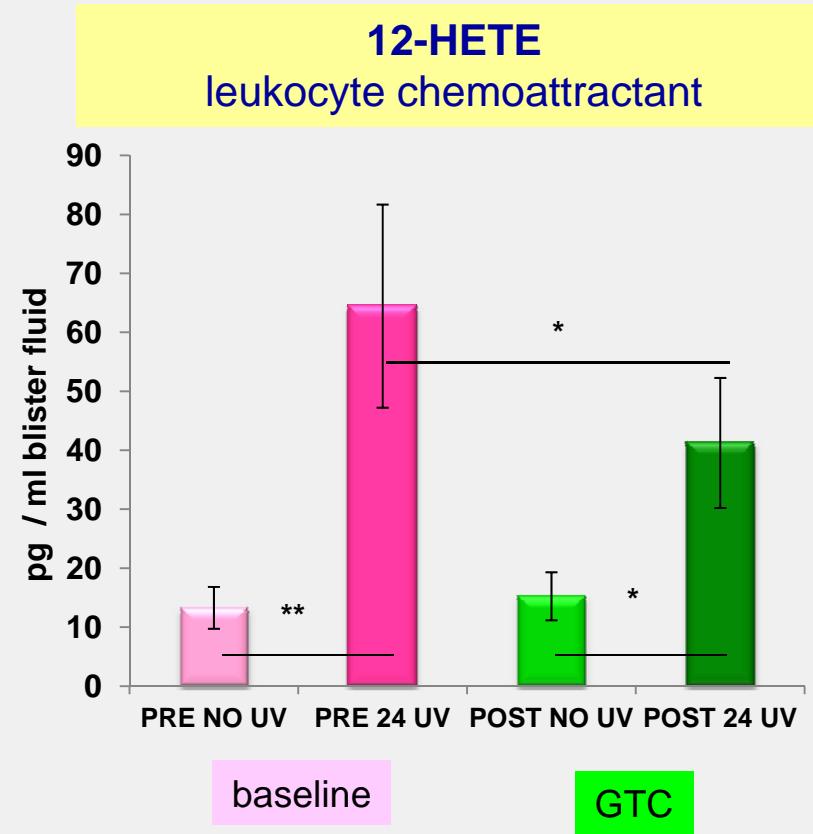
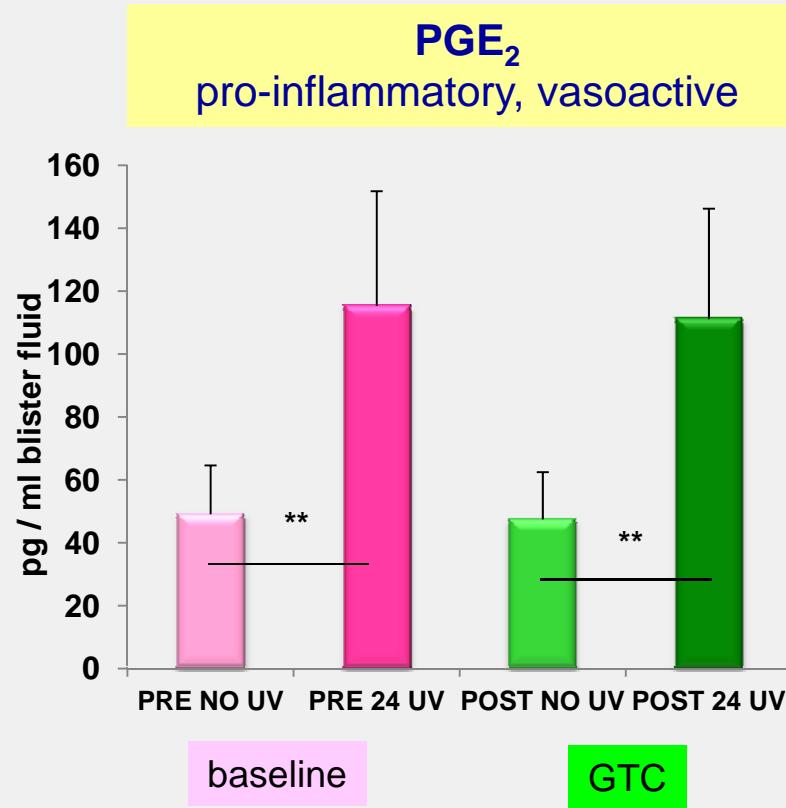
Healthy human volunteers: n=14, 27-56 yrs; all female;  
phototype I/II (tend to burn not tan)

Supplement: GTC 550 mg/day + 50mg/day vit.C

Study period: 12 weeks



# Effect of low dose GTC on cutaneous eicosanoids



\* p<0.05 \*\* p<0.001

Rhodes et al BJR 2013

# A Consideration of Biomarkers to be used for Evaluation of Inflammation in Human Nutritional Studies

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Table 2. Lipid mediators associated with inflammation\*

Class	Mediator	Substrate	Receptor(s)
Prostanoids	PGD <sub>2</sub>	Arachidonic acid via COX	DP1, DP2
	PGE <sub>2</sub>	Arachidonic acid via COX	EP1, EP2, EP3, EP4
	PGF <sub>2α</sub>	Arachidonic acid via COX	FP
	PGI <sub>2</sub>	Arachidonic acid via COX	IP
	TXA <sub>2</sub>	Arachidonic acid via COX	TP
	PGE <sub>1</sub>	Dihomo-γ-linolenic acid via COX	EP1, EP2, EP3, EP4
	PGD <sub>3</sub>	EPA via COX	DP1, DP2
	PGE <sub>3</sub>	EPA via COX	EP1, EP2, EP3, EP4
	5-HETE	Arachidonic acid via 5-LOX	BLT2
	5-HPETE	Arachidonic acid via 5-LOX	OXE
Leukotrienes	LTB <sub>4</sub>	Arachidonic acid via 5-LOX	BLT1, BLT2
	LTC <sub>4</sub> , D <sub>4</sub> , E <sub>4</sub> (termed cys-LT)	Arachidonic acid via 5-LOX	CysLT1, CysLT2
	15-HETE	Arachidonic acid via 15-LOX	BLT2
	15-HPETE	Arachidonic acid via 15-LOX	BLT2
	12-HETE	Arachidonic acid via 12-LOX	BLT2
	LTB <sub>5</sub>	EPA via 5-LOX	BLT1, BLT2
	LXA <sub>4</sub>	Arachidonic acid via 15-LOX and 5-LOX or 5-LOX and 12-LOX (transcellular)	FPR2/ALX
Endocannabinoids	2-Arachidonoylglycerol	1,2-Diacylglycerol with arachidonic acid at the <i>sn</i> -2 position	CB1, CB2
	Anandamide	<i>N</i> -arachidonoylphosphatidylethanolamide via phospholipase D; in turn, <i>N</i> -arachidonoylphosphatidylethanolamide is formed from phosphatidylcholine with arachidonic acid at the <i>sn</i> -1 position and phosphatidylethanolamine	CB1, CB2
Resolvins, protectins and maresins	RvE1	EPA via acetylated COX-2 and 5-LOX (transcellular)	RvE1 (ChemR23), BLT1
	RvD1	DHA via acetylated COX-2 and 5-LOX or via 15-LOX and 5-LOX (transcellular)	RvD1 (GPR32), ALX/FPR2
	PD1 (NPD1)	DHA via 15-LOX and LOX (transcellular)	Not yet known
Lysolipids	MaR1	DHA via 15-LOX and 12-LOX (transcellular)	Not yet known
	PAF	Phosphatidylcholine with diethyl ether link at the <i>sn</i> -1 position	PAF-R
	Lyso-PA	Phosphatidic acid, which in turn is synthesised from phosphatidylcholine	LPA1, LPA2, LPA3, LPA4, LPA5, LPA6
	Sphingosine-1-phosphate	Sphingosine, which in turn is synthesised from ceramide	S1P1, S1P2, S1P3, S1P4, S1P5

# Mediator Lipidomics

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- LC/ESI-MS/MS mediator lipidomics: versatile, sensitive approach.
- Role of lipid mediators in health and disease.
- Discovery of novel mediators and biomarkers; development of therapeutics.
- Contribution to systems biology.

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